

Benefits of liquid urea and a microbial catalyst on biomass and the nutritional value of Mombasa grass

Beneficios de la urea líquida y un catalizador microbiano en la biomasa y el valor nutricional del pasto Mombasa

Shirley Lorena Alquichire Rojas ^{1,3}, Elide Valencia-Chin ^{2,4}.

¹Universidad Católica de la Santísima Concepción. Concepción, Chile. ²Universidad de Puerto Rico. Mayaguez, Puerto Rico.

³ ✉ shirley.alquichire@upr.edu, ⁴ ✉ elide.valencia@upr.edu



<https://doi.org/10.15446/acag.v71n1.99814>

2022 | 71-1 p 64-72 | ISSN 0120-2812 | e-ISSN 2323-0118 | Rec.: 2021-11-30 Aprob.: 2022-03-25

Abstract

During 2019, an experiment was conducted at the Agricultural Experiment Station in Isabela (Puerto Rico) on an Oxisol with previously well-established stands of cv. Mombasa. This experiment assessed the effects of a microbial catalyst (MC) and liquid urea 22-0-0 (LU) at a rate of 168 kg ha⁻¹ (in split applications), a mixture of LU+MC and a control on aboveground biomass, root biomass, nutritional value, nitrogen use efficiency and soil parameters on cv. Mombasa at 35-day (d) harvests during six harvests. The study was established in a completely randomized design with four replicates. The effects of LU and MC on belowground (root) biomass were determined by collecting samples in 1 m² to determine the yield and chemical composition. Soil samples were collected at 15 cm depth using a soil corer at the first and 6th harvest from each plot to assess organic carbon (OC), total nitrogen (TN), pH, macronutrients, and cation exchange capacity (CEC). The results showed that aboveground biomass doubled using LU (2369 kg DM ha⁻¹) compared to the control and MC (1100 kg DM ha⁻¹). Crude protein (CP) was 10.1 % using LU. Neutral Detergent fiber was 70 % for the control, 74.2 % when LU was used, and around 40 % for acid detergent fiber (ADF) for any treatment. Overall, there were no significant effects of treatments on OC and organic matter percentages, P, N, Ca, Mg, and CEC. In conclusion, LU is an excellent source of N for Mombasa, but shorter harvest frequencies may be required to improve the fiber quality of Mombasa.

Keywords: biofertilizer, bromatological variables, nitrogen fertilizer, organic carbon, tropical forage.

Resumen

Durante el año 2019 se llevó a cabo un experimento en la Estación Experimental Agrícola de Isabela (Puerto Rico) en un Oxisol con un pastizal previamente establecido con el cv. Mombasa. El objetivo de este experimento fue evaluar el efecto de un catalizador microbiano (CM) y la urea líquida 22-0-0 (UL) a una tasa de 168 kg ha⁻¹ (en aplicaciones subdivididas), una mezcla de MC+UL y un control sobre la biomasa aérea, biomasa de la raíz, el valor nutricional, la eficiencia de uso de nitrógeno y parámetros del suelo sobre el cv. Mombasa a los 35 días de cosecha durante seis periodos de cosecha. El estudio fue establecido en un diseño completamente aleatorizado con cuatro replicas. Los efectos de CM y UL sobre la biomasa aérea y la producción de raíces fueron determinadas mediante la recolección de muestras en 1 m² para determinar el rendimiento y la composición química. Las muestras de suelo fueron recolectadas a 15 cm de profundidad usando un barreno en la 1^{era} y 6^{ta} cosecha en cada unidad experimental para evaluar el carbono orgánico en el suelo (CO), el nitrógeno total (NT), el pH, los macronutrientes y la capacidad de intercambio catiónico (CIC). Los resultados mostraron que el rendimiento de la biomasa aérea fue mayor cuando se usó UL (2369 kg MS ha⁻¹) comparado con el control y el CM (1100 kg MS ha⁻¹). La proteína cruda (PC) fue de 10.1 % usando UL. La fibra detergente neutra (FDN) fue de 70 % para el control y 74.2 % cuando se usó UL, y alrededor del 40 % para la fibra detergente ácida (FDA) durante todos los tratamientos. No se encontraron efectos significativos de los tratamientos sobre el porcentaje de CO y materia orgánica, P, N, Ca, Mg y CIC. En conclusión, la UL es una excelente fuente de N para el pasto guinea local, pero es posible que se necesiten frecuencias de cosecha más cortas para mejorar la calidad de la fibra.

Palabras clave: biofertilizante, carbono orgánico, fertilizante nitrogenado, forraje tropical, variables bromatológicas.

Introduction

Forage grasslands are the main source of ruminant feeding in the tropics (Estrada *et al.*, 2015). However, perennial grasses (*Poaceae*) and legumes (*Fabaceae*) can be affected by weather and rainfall. Nowadays, much of the literature emphasizes that rainfall is the most critical condition to produce dry matter (DM) (Perera *et al.* 2019; Norman *et al.* 2021). Widely evidence suggests that plant fertilization is a key to reaching high-level production (Martínez-Dalmau *et al.*, 2021). Hence, studies argue the needs for better nutrient management and planning to recover the rates after harvesting or grazing (Norton *et al.*, 2013; Teague *et al.*, 2013). Specifically, nitrogen (N) is an essential compound for constructing amino acids, proteins, and chlorophyll (Islam *et al.*, 2022). Moreover, it is responsible for the development of roots and the proper establishment of pastures (Sakiroglu *et al.*, 2020). A possible way to achieve good levels of N in soil is through the biological fixation by specialized microorganisms or abiotic fixation (Hodge, 2005; Prosser, 2005). Because of the scarce N supply on both fixation ways, N inorganic fertilizer input is an important source of N for plants.

Likewise, soils play an important role in water-nutrient retention, carbon (C) reservoirs, and agricultural productivity. However, Oxisols (which represent 23 % of the soils in the intertropical zone) and Ultisols (18 %) are predominant in the island of Puerto Rico and are widely used as pastures for livestock; these soil orders are equivalent to 10.8 % and 19.8 % of the area of Puerto Rico, respectively (Muñoz *et al.*, 2018). Also, 2.1 % of the total land area is used for hay and grazing forages (USDA, 2014). These predominant soil groups are highly weathered soils with low fertility, as well as naturally acidic and highly erodible. Additionally, these soils have a big lack of N. Furthermore, their intensive use also causes deterioration of their physicochemical properties such as compaction, and decreases cation exchange capacity (Beinroth, 2000). This situation justifies the use of N fertilizer, alternative fertilizers, and more productive grasses to improve soil fertility and yield production.

Tropical perennial grasses, such as the *Megathyrsus* spp. (cv. Mombasa), have a high breeding potential for digestible energy. In Puerto Rico, local guinea grass at 50-d harvest reported crude fiber of 36.9 % (Vicente-Chandler *et al.*, 1964). Likewise, in a Latossolo Vermelho-Amarelo distrófico in Matto Grosso, Brazil, 100 kg of urea-N ha⁻¹ was effective increasing the mean of dry matter yield (DMY) by three consecutive harvests of 3399, 1856, and 1488 kg ha⁻¹, which was significantly different (Olivastro *et al.*, 2018). Thus, rates from 60 to 100 kg N ha⁻¹ every 40-45 days increased CP (> 7 %) and leaf-stem ratio. With a total rate of 180 kg N-urea ha⁻¹, DMY was on average 5704 kg ha⁻¹ when harvested every 40-45 days in the established year (Hare *et al.*, 2015). Mombasa grass is of great relevance for tropical agriculture systems due to its high biomass production and nutritional value (Caldas *et al.*, 2020).

On tropical soils, nutrient release may be limited by microbial flora since their interaction with the decomposition of organic matter (OM) and enables nutrient mineralization and its accessibility to forages (Zhang *et al.*, 2021). For this reason, a microbial catalyst (MC) is of great interest, because an improvement in the microbial population is expected. A rapidly absorbed N fertilizer such as liquid urea (LU) contributes to the nutritional value and vigor of regrowth, in addition to root growth in tropical forages. Afterall, low losses under field conditions of liquid urea fertilizer are expected to mitigate ammonia losses and increase nitrogen use efficiency (NUE) and crop yield.

Urea fertilizer undergoes hydrolysis after its application and reacts under the presence of the urease enzyme. Then, ammonification process releases NH₄⁺ and, by nitrification, produces NO₃⁻ ions; finally, after plant uptake, the N source presents losses by volatilization (Sun *et al.*, 2008) Mariano. *et al.* (2019) found that urea remained on the soil surface four to six days after fertilization, thus reducing ammonia losses. Galindo *et al.* (2017, 2018) found advantages when 300 kg N ha⁻¹ was used because N fertilization is more efficient and does not cause damage at high doses (1200 kg N ha⁻¹) in the tropics. At low dosages, Castagnara *et al.* (2011) observed that Mombasa treated with doses ranging from 40 to 160 kg urea-N ha⁻¹ showed acceptable CP, NDF and ADF percentages (8.7 %, 73 % and 42 %, respectively).

N fertilization has been limited or non-existent due to prohibition and high costs of N fertilizers in Puerto Rico. However, little is known so far about the use of LU as N fertilizer or MC and its effects on the yield of cv. Mombasa. For the tropical and sub-tropical area, the use of these nutrient sources would assist in the adoption of a better N fertilizer use management. The LU will be potentially useful in N plant nutrition with positive effects on shoot and root biomass and shoot nutritional value. In this respect, further research has suggested that LU or urea and micronutrients improve shoot biomass production and increase the nutritional value (Barros *et al.*, 2018; Caldas *et al.*, 2020; de Oliveira *et al.*, 2020; Escarela *et al.*, 2017; Galindo *et al.*, 2017). The aim of this study was to compare the effect of a MC and LU on dry matter yield (DMY) and the nutritional value on five-week regrowth of well-established stands of Mombasa from March to October 2019, as well as to determine the biomass of root tissues, the percentage of soil organic carbon (OC) and soil nutrients.

Materials and methods

The study was conducted at the Agricultural Experiment Station of Isabela (AES) (17° 27' 32" N and 67° 02' 53" W; average annual rainfall of 1451 mm) of the University of Puerto Rico, located in Mayagüez, northwest of Puerto Rico. Well-established Mombasa on an Oxisol of Coto series (very fine, kaolinic, isohyperthermic, Typic Hapludox) (Muñoz *et al.*, 2018) was used for this study from March 18 to October 28, 2019. The climate was tropical humid (data provided by AES) (Table 1).

The experimental design was completely randomized. Mombasa crop was established on October 31, 2018, and a standardized cut (10-cm height) was conducted on March 18, 2019, using a flail-chopper. A 40 x 37 m plot area was digenervated into 16 experimental units. Treatments: control (without liquid urea or microbial catalyst), microbial catalyst (MC) (Table S1), liquid urea (LU) (Table S2) and MC+LU were randomly assigned to each plot in split applications (on March 19 and July 13, 2019). Both applications of LU were of 84 kg N ha⁻¹ yr⁻¹; both applications of MC were of 1.0 L ha⁻¹; while the MC+LU consisted in two doses applied through foliar spraying for each experimental unit. Four of the experimental units were left as control. Treatments were applied on the same day of the standardized cut and after the 3rd harvest. The experimental variables measured were DMY, ADF, NFD and CP in the aboveground biomass and DMY and dry matter percentage (DM %) in root biomass for each treatment after the sixth 5-week harvests, as well as micro and macronutrients, pH, CEC and OM on soil samples.

Aboveground biomass was harvested at 15 cm above the soil in 1 m² every 5 weeks from the center of each experimental plot (May 6, June 10, July 15, August 19, September 23, and October 29, 2019). The samples were weighed and then placed in a forced air oven at 60 °C for 48 hours. Afterwards, the dried samples were weighed again and the moisture percentage and DMY were obtained from fresh and dry weight, calculated using the equations [1] and [2]. Dried samples were obtained individually using a Thomas-Wiley Laboratory Mill Model 4, passed through a 1-mm screen and stored in a plastic bag for ADF, NDF and N analysis. Ground tissue samples were analyzed at the Dairy One Forage Testing Laboratory in Ithaca, New York, for the calculation of CP (calculated with equation [3]), NDF and ADF, as suggested by Van Soest (1994).

$$\% DM = \frac{\text{dry weight}}{\text{dry weight}} \times 100 \quad (\text{Eq. 1})$$

$$DMY = \frac{DFM \times \%DM}{1000 \text{ kg ha}^{-1}} \quad (\text{Eq. 2})$$

$$\% CP = \% N \times 6.25 \quad (\text{Eq. 3})$$

NUE was suggested by Dobermann (2005) and calculated in this research to provide a general view of plant performance because the objective of these indexes is to provide evidence about the dose of fertilizer and its economically optimum plant nourishment while minimizing losses.

The estimated weight of root biomass assumes the C potential sequestered on soils. Therefore, roots and crown biomass samples were collected in 0.25 m² (after final harvests). Roots were transported

to the laboratory, washed with water to remove soil, air-dried for one day, and weighed. The root samples were stored in paper bags, dried at 60 °C for 72 hours in a forced air oven and then weighed again.

Soil samples were collected from the experimental plot at a depth of 15 cm on two sampling dates, March 19, before treatment, and October 26, 2019, using a grid and a soil corer for the basic soil fertility analysis. The soil samples collected from the last harvest were made in 0.25 m² and 15-cm depth in each treatment in the center of the plots. Collected soil samples were placed in paper bags and air-dried for five days, then, they were grounded and sieved. Soil nutrients and cation exchange capacity (CEC) were measured by Mehlich III extraction (Sparks, 2007) and filtrated for P, K, Ca, Fe, Mn and Zn in an ICP-OES 7300 DV (Optical Emission Spectrometer). Soil pH was measured in a 1:1 soil, water slurry and CaCl₂ solution. Soil OC was analyzed using the dichromate-oxidation Walkley and Black as described by Nelson and Sommers (1982). The results of OC were calculated using the equations [4] and [5].

$$\%OM = \%OC \times 2.24 \quad (\text{Eq. 4})$$

$$OC = 0.58 \times OM \quad (\text{Eq. 5})$$

The data was examined using a two-way ANOVA corresponding to the mathematical model $Y_{ij} = \mu + \alpha_i + \beta_j + \alpha\beta_{ij} + \varepsilon_{ij}$, with the fixed factors "treatment" (α) and "harvest" (β), and a one-way ANOVA was made for DMY and soil data analysis using "treatment" as a fixed factor. The Tukey's test and Least significant difference (LSD) method (level of significance $p < 0.05$) were used as post hoc tests in order to reveal significant differences among treatments. Assumptions of homogeneity and normality of variance were tested using the tests Levene's and Shapiro-Wilks, respectively. The whole statistical analysis was performed using the SAS system[®] (SAS, 2021).

Table 1. Mean monthly rainfall (mm), minimum, medium and maximum temperature from March to October, 2019 at the Agricultural Experiment Station Isabela, Puerto Rico

Month	Rainfall (mm)	T min. °C	T med. °C	T max. °C
March	205	18.4	23.6	29.2
April	56	19.6	25.3	31.5
May	296	20.9	25.1	30.9
June	53.6	20.5	26.1	30.5
July	91.8	21.2	26.5	31.2
August	131	22.1	26.7	33.1
*September	-	-	-	-
October	144	21.8	26.1	31.3

*Data not available. T min.: minimum temperature, T med.: medium temperature, T max.: maximum temperature.

Table S1. Property specifications of Microbial catalyst

Characteristic	Value
Co	0.83 %
Cu	0.47 %
Mn	0.42 %
Mo	0.0003 %
Zn	0.40 %
pH	4.65
Density	1.07 g cm ⁻³

Note: *Information included in the product label.

Table S2. Property specifications of Liquid urea 22-0-0

Density kg/L	Density lb/gal	N total lb/gal	N kg/100 L	Minimum Biuret
5.63	9.70	2.1	121.8	≤ 2%

*Information included in product label.

Results

For DMY, there was a significant interaction between the treatments. The DMY decreased between each period of harvest (1st and 4th harvest) (Table 2). Yield mean values of six accumulated harvests (210 days after planting) showed significant differences. The lowest DMY was observed for the control and MC treatments, with 1091 and 1110 kg ha⁻¹, respectively. These results suggest that MC has no effect on increasing DMY of Mombasa. Dry matter yield doubled on average (>1.0 Mg ha⁻¹) with the application of LU. However, adding MC to the LU did not differ from the sole application of LU, confirming that there was no response of Mombasa to the microbial catalyst treatment MC (Table 2). Accumulated DMY (six 35-d harvests) of LU showed an increase in biomass of over 6 Mg ha⁻¹ in contrast with the control treatment (Table 2).

There was no significant interaction between fertilizer treatments at any harvests for CP, but there was a significant main effect for fertilizer treatments (Table 3). Crude protein concentration was similar for the control and MC, averaging 7.4 %. Crude protein percentage was 1.3 % higher in LU treatment compared with MC+LU treatment. This difference is not clearly understood, as a hypothesis was that MC should affect positively N concentrations in plant tissue. The LU applications were approximately 2.3 % higher than the control, indicating a positive effect of LU on N concentrations on tissue.

The apparent recovery efficiency by difference (Er), removal efficiency (RE), CP produced by kg of N applied, partial factor productivity (PFP) and agronomic efficiency (AE) are shown in the supplemental material (Table S3 and S4). The applied doses of DMY and N and the N concentration in

tissues were used to calculate these values. This index interpretation was made to provide useful information for growers and livestock farmers to keeping records of inputs, outputs and current status of NUE at a specific place, time and rate. In summary, the PFP index values in this study ranged from 40 to 70 with values > 49. The increase of AE was between 23 and 43 kg in yield per kg of N applied. The Er is better close to 100 %, as shown in the experiments over > 25 to 81 %, and for RE, it is better > 80 %; the values shown in this research were > 69 %. All indexes correspond to well-managed systems. Also, cumulative CP in DMY by kg of N applied was higher in the 4th harvest compared to the 1st one, where the range of CP biomass was 357 to 518 kg CP ha⁻¹. The index values were around 4.38 and 5.3 kg CP produced per kg of N applied (Table S4).

Table 4 presents the percentage values and analysis of variance for ADF and NDF in tissues of Mombasa on the 1st and 4th harvest. There was no interaction of fertilizer treatment by harvest on ADF, but differences per treatment were found. Mean percentage values for ADF were 39.9 % for MC, while those treated with MC+LU and LU averaged 42.2 % (two percentage points over the MC). An interaction was observed on fertilizer treatment by harvest for NDF; the average for the LU was 74.2 % in contrast to the control (71 %).

There were no significant differences between fertilizer treatments for root biomass (Table 5), which averaged 836 kg DM ha⁻¹ for LU and MC+LU and 560 kg DM ha⁻¹ for the control. The leaf-root ratio (LRR) was 3.7 for the LU treatment and 2.0 for the control. While N had a significant effect on aboveground biomass of Mombasa, it had little or none effect on root biomass.

As can be observed in Table 7, the K content was significantly different. The K value was higher in the control group (0.37 meq 100 g⁻¹) than MC and LU with 0.22 and 0.14 meq 100g⁻¹, respectively. As reported in this experiment, N fertilization, specifically N-NH₄⁺,

Table 2. Mean of the effects of the interaction of treatments on dry matter yield (kg ha⁻¹) in 6 35-d harvests of Mombasa at the Agricultural Experiment Station Isabela, Puerto Rico

FT	Harvest (kg ha ⁻¹)						Total
	1	2	3	4	5	6	
² Control	2088	906	733	1180	890	1000	6797b
³ MC	1953	1001	678	980	877	973	6462b
⁴ LU	3663	1582	909	4878	1400	1586	14018a
⁵ MC+LU	3269	1389	900	4927	1625	1380	13490a

¹Fertilizer treatment (FT), ²Control group, ³Microbial catalyst (MC), ⁴Liquid urea (LU), ⁵Microbial catalysts (MC) plus liquid urea (LU).

*Means followed by different letters in the same column are significantly different among treatments ($p \leq 0.0001$) using a one-way analysis of variance (ANOVA) and Tukey's test was used as a post hoc test.

Table 3. Fertilizer treatment effects and summary of analyses of variance for nitrogen (N) and crude protein (CP) percentages of Mombasa at the Agricultural Experiment Station Isabela, Puerto Rico

Variable	³ Control	⁴ Microbial catalyst	⁵ LU	⁶ MC+LU	Source of variation		
					⁷ FT	⁸ H	⁹ FT×H
¹ N %	1.24	1.19	1.62	1.40	0.0004*	0.0291*	0.5790
² CP %	7.77bc	7.44c	10.15a	8.79b			

¹Nitrogen percentage, ²Crude protein percentage, ³Control group, ⁴Microbial catalyst (MC), ⁵Liquid urea (LU), ⁶Microbial catalysts (MC) plus liquid urea (LU), ⁷Fertilizer treatment, ⁸Harvest, ⁹Interaction between fertilizer treatments, *harvest. Means followed by different letters in the same row are significantly different among treatments and harvest ($p \leq 0.05$) using a two-way analysis of variance (ANOVA) and Tukey's test was used as a post hoc test.

Table S3. Liquid urea and liquid urea plus microbial catalyst effect on N recovery in Mombasa (1st and 4th harvest) and apparent recovery efficiency by difference at the Agricultural Experiment Station Isabela, Puerto Rico

¹ FT	-----N (%)-----			N in biomass (kg ha ⁻¹)			*Apparent recovery efficiency by difference (Er)	
	Harvest		Mean	Harvest		Mean	kg increase absorption per kg of nutrient applied (%)	
	1 st	4 th		1 st	4 th		1 st	4 th
² Control	1.24	1.25	1.25	18	14.6	16.3	-	-
³ LU	1.56	1.69	1.62	58.8	83.0	70.9	48	81
⁴ LU+MC	1.27	1.54	1.40	41.9	75.5	58.7	28	72

¹Fertilizer treatment (FT), ²Control group, ³Liquid urea (LU), ⁴Liquid urea (LU) plus microbial catalyst (MC). * Two applications of 84 kg N ha⁻¹ each, and mean per treatment were made, and depending on the index, a control value was used.

Table 4. Fertilizer treatments and summary of analysis of variance for ADF and NDF of Mombasa at the Agricultural Experiment Station Isabela, Puerto Rico

Variable	³ Control	⁴ Microbial catalyst	⁵ LU	⁶ MC+LU	Source of variation		
					⁷ FT	⁸ H	⁹ FT×H
¹ ADF %	40.4ab	39.8b	42.2a	42.4a	0.0083*	0.3413	0.5673
² NDF %	71.0b	70.5b	74.2aa	74.2a	<0.0001*	0.3231	0.0055*

Note: ¹Acid detergent fiber percentage, ²Neutral detergent fiber percentage (NDF), ³Control group, ⁴Microbial catalyst (MC), ⁵Liquid urea (LU), ⁶Microbial catalysts (MC) plus liquid urea (LU), ⁷Fertilizer treatment, ⁸Harvest, ⁹Interaction between fertilizer treatments. Means followed by different letters in the same row are significantly different among treatments and harvest ($p \leq 0.05$) using two-way analysis of variance (ANOVA) and Tukey's test was used as a post hoc test.

Table S4. Removal efficiency (RE) and CP produced by kg of N applied on Mombasa at the Agricultural Experiment Station Isabela, Puerto Rico

¹ FT	*Removal efficiency		
	kg N in tissue	kg absorption per kg of applied nutrient	
	Harvest		Mean
	1 st	4 th	kg CP kg ⁻¹ N
² LU	70.9		0.84
³ LU+MC	58.7		0.69
	*Crude protein produced by kg of N applied		
LU	4.37	6.21	5.30
LU + MC	3.06	5.70	4.38

¹ Fertilizer treatment (FT), ²Liquid urea (LU), ³Liquid urea (LU) plus microbial catalyst (MC). *Mean biomasses of cuts 1st and 4th in kg ha⁻¹.

depletes soil K content. Overall, there were no significant effects of treatments on the percentage of OC, OM, P, N, Ca, Mg and CEC. However, there was an increase in N and a decrease in P, as observed in the LU treatment, opposed to the soil conditions before the experiment (Table 6).

Discussion

Several studies have reported the positive effect of N fertilization on forage DMY including Mombasa at 28 to 40-days harvests (Barros *et al.*, 2018; de Oliveira *et al.*, 2020; Escarela *et al.*, 2017; Galindo *et al.*, 2017). The present study showed an increase of 100 % in the biomass with LU compared to the control after the first 35-d harvest (Table 2). Barros *et al.* (2018) reported positive effects of the rate of N (100, 200 and 400 kg ha⁻¹) with urea and coated urea. As observed in this study, similar results with 168 kg N ha⁻¹ of LU showed high DMY (Table 2). Regarding the N source, similar performance was found by de Oliveira *et al.* (2020), when they applied 150 kg N ha⁻¹ as ammonium sulfate. It must be noted that Galindo *et al.* (2017) studied N-sources such as urea and ammonium nitrate at different doses by cutting it during the dry season, and found no influence of it among sources and doses on biomass production.

The results of cumulative DMY in the present study (Table 2) agree with those reported by Galindo *et al.* (2017, 2018), where cumulative yields with N fertilization during the dry season reached 10,518 kg DM ha⁻¹ applying 300 kg N ha⁻¹. For example,

Table 5. Effects of fertilizer treatments on Mombasa root dry matter yield and leaf root ratio after six 35-d harvests at the Agricultural Experiment Station Isabela, Puerto Rico

Variable	¹ FT			
	² Control	³ Microbial catalyst	⁴ LU	⁵ MC+LU
Root biomass (kg DM ha ⁻¹)	560	416	646	836
⁶ DM %	49	53	50	48
⁷ LRR	66:33 (2.0)	72:27 (2.7)	78:21(3.7)	73:27 (2.7)

¹Fertilizer treatment (FT), ²Control group, ³Microbial catalyst (MC), ⁴Liquid urea (LU), ⁵Microbial catalysts (MC) plus liquid urea (LU), ⁶dry matter percentage and ⁷Leaf root ratio. No significant means among treatments were found ($p \leq 0.05$) based on a one-way analysis of variance (ANOVA) and Tukey's test was used as a post hoc test.

Table 6. Soil chemical properties on Mombasa plots (study initiated on March, 2019) at the Agricultural Experiment Station Isabela, Puerto Rico

*Attributes	Units	Value
pH in water		5.4
pH in CaCl ₂		4.73
Phosphorus	mg kg ⁻¹	18.4
Nitrogen	mg kg ⁻¹	28
Potassium	meq 100g ⁻¹	0.31
Calcium	meq 100g ⁻¹	2.85
Magnesium	meq 100g ⁻¹	0.92
Organic carbon	%	1.25
Organic matter	%	2.16
CEC	meq 100g ⁻¹	4.08

*No calibrated rating

mean yields were more than 1700 kg DM ha⁻¹, which is adequate for animal consumption (Galindo *et al.*, 2018). Galindo *et al.* (2017) also observed that guinea grass presented the maximum response with increasing N-rates (range of 300 to 1200 kg N ha⁻¹ yr⁻¹). Caldas *et al.* (2020) found no significant differences between granular urea and LU with micronutrients on biomass production, however, sole granular urea DMY (3767 kg ha⁻¹) was slightly higher than LU (2871 kg ha⁻¹). Overall, these results were similar to our findings among treatments with LU and LU+MC (Table 2).

An improvement in Mombasa grass CP content using high levels of N input was proven by several studies when harvesting at 24 to 45-d in optimal conditions (Caldas *et al.*, 2020; de Oliveira *et al.*, 2020; Escarela *et al.*, 2017). The mean values of the CP percentage observed in this study at 35-d were 10.2 % for LU and 7.4 % for MC (Table 3). For example, Galindo *et al.* (2018) reported CP values of 12 to 14 %, more than the control with 10 %, using ammonium nitrate or granular urea, respectively. Caldas *et al.* (2020) found crude protein content ranged from 12 to 13 %

when harvested at 24 to 30-d with similar doses of N. With other grasses such as Marandu grass, Delevatti *et al.* (2019) forage accumulation rate (FAR reported values close to 16 % CP. It must be noted that high N application rates promote a substantial increase in biomass and can impact the CP level (Escarela *et al.*, 2017). However, our results differed; this study showed ≤ 10 % of CP, corresponding to limited CP requirement by livestock (Abbasi *et al.*, 2018).

In this study, both ADF and NDF values were high due to the mature growth of Mombasa (35-d regrowth) (Table 4). Mean percentage values for ADF were 39.9 % for MC, while those treated with MC+LU and LU averaged 42.2 % (two percentage points over the MC). These values were similar to corresponding values reported by Castagnara *et al.* (2011) for ADF (42 %) at 42-d harvests. However, lower values of ADF were found by Delevatti *et al.* (2019) forage accumulation rate (FAR with 27.8 % for 180 kg N ha⁻¹, de Oliveira *et al.* (2020) reported 35.6 % and Galindo *et al.* (2018) 34.8 % using 50 kg N ha⁻¹ when Mombasa was harvested at 28-d regrowth. Our NDF values were similar to those reported by Castagnara *et al.* (2011), that reported 73.5 % of NDF, and de Oliveira *et al.* (2020) with 70.5 %. However, it is possible to reach lower levels as shown by Galindo *et al.* (2018) (around 67 % using 50 to 100 kg N ha⁻¹) and Delevatti *et al.* (2019) forage accumulation rate (FAR (56.4 % using 180 kg N ha⁻¹).

Several authors such as de Oliveira *et al.* (2020) and Delevatti *et al.* (2019) of *Cenchrus ciliaris* and *Panicum maximum* grown under irrigation at Gode, Somali region. The study was executed using 2 x 3 factorial arrangements in randomized complete block design with three replications. The treatments were three level of fertilizer application (0, 50, 100 kg ha⁻¹ of urea reported that NDF and ADF increased with increasing levels of N. This study contrasts with previous studies, for example, with the low values of NDF at high N doses reported by Caldas *et al.* (2020) and Galindo *et al.* (2018). High NDF levels reduce voluntary feed intake and digestibility while high ADF levels reduce forage digestibility. These results could be explained because high N rates promote the growth of new structures and forage that induces lignin synthesis to facilitate a better mechanical self-assembly of tissues and the dilution effect in the cell wall, possibly responsible for the decrease of NDF (de Oliveira *et al.*, 2020). However, this mechanism was not observed in this study. It could be argued that the present results are due to the high capacity of accumulating structural carbohydrates (e. g., Galindo *et al.*, 2018).

Warm-season grass yield has a negative response to N fertilization, indicating that, for these species, any level of N input resulted in the allocation of proportionally less biomass to the roots in favor of the allocation to the shoots. However, in the present study, there was evidence of a large difference when

Table 7. Effects of fertilizer treatments on soil chemical properties after six 35-d harvests of Mombasa at the Agricultural Experiment Station Isabela, Puerto Rico

¹ FT	pH		⁹ OC	¹⁰ OM	¹¹ P	¹² TN	K ⁺	Ca ²⁺	Mg ²⁺	¹³ CEC
	CaCl ₂		-----%-----		----mg kg ⁻¹ ----		-----meq 100 g ⁻¹ -----			
² Control	5.33	4.55	1.27	2.18	16.3	54.3	0.37b	2.13	0.86	3.35
³ MC	5.40	4.56	1.26	2.17	15.1	54.3	0.22a	2.23	0.86	3.29
⁴ LU	5.21	4.46	1.15	1.98	14.4	61.3	0.14a	1.82	0.68	2.63
⁵ MC+LU	5.19	4.28	1.19	2.05	12.9	70	0.14a	1.71	0.65	2.49
⁶ p-value	ns	ns	ns	ns	ns	ns	*	ns	ns	ns
⁷ LSD	0.39	0.38	0.33	0.58	8.8	24.8	0.10	0.79	0.26	0.9162
⁸ CV %	3.58	4.12	13.2	13.2	28.7	19.7	16.1	19.6	15.0	14.78

¹Fertilizer treatment (FT), ²Control group, ³Microbial catalyst (MC), ⁴Liquid urea (LU), ⁵Microbial catalysts (MC) plus liquid urea (LU), ⁶p-values results significant ($p \leq 0.05$) and no significant results (ns) ($p \geq 0.05$). Means within a column followed by the same letter are not significantly different among treatments based on a one-way analysis of variance (ANOVA). ⁷Least Square Difference (LSD) at $p < 0.05$ was used as a post hoc test. ⁸C.V. % is the coefficient of variation. ⁹Organic carbon, ¹⁰organic matter, ¹¹phosphorus, ¹²total nitrogen, ¹³cation exchange capacity.

high N was put in with MC, versus the control (Table 5). While N had a significant effect on aboveground biomass of Mombasa, it had little or no effect on root biomass. In this study, there was 100 % more root biomass in fertilizer plots using a high N rate than on unfertilized plots. This experiment considered a possible trend that root mass was higher in the LU treatment, but insufficient evidence was found. Root biomass and rhizodeposition decreased after applying an available N source because plants use C reserves for new regrowth (Zang *et al.*, 2016) especially in the rhizosphere (C excess and N limitation. These processes restrict organic matter stabilizing, for example, urea fertilization depletes soil K content (Table 7). According to Reimer *et al.* (2020), a higher N input represents mining nutrients from the soil, specially P, K, and Mg. Maltas *et al.* (2018) carbon (C reported a non-significant but positive effect of N fertilization on OC. The effect observed (Table 7) agrees with previous studies that reported that urea fertilization decreased OC decomposition (Li *et al.*, 2017) but this effect is still very uncertain and depends on living plants. We investigated the effects of mineral N (N_{min}, and that the percentage of OC remained constant.

Our results underscore the significance of improving N fertilization, NUE and harvest frequencies and its effects on biomass production and chemical soil properties. Moreover, these results offer a comprehensive view of management practices to take advantage of LU and most productive grass varieties like Mombasa. Further research could be conducted to find an economic way to supply N and promote the adoption of better nutrient management practices.

Conclusion

LU provided an excellent source of N, especially in the NH₄⁺ form required by Mombasa. This research showed the LU application rate and MC as an alternative to enhance Mombasa production in Puerto Rico through synchronized 35-d harvests. The CP concentration with LU suggests that split applications of LU might be profitable in the intensive management of this grass under cutting. Finally, detergent fiber, NDF and ADF were high for 35-d harvests; shorter harvest frequencies could be required to improve the fiber quality of Mombasa.

Acknowledgments

We thank the Tropical Agriculture Research Station (TARS) at Mayagüez, Puerto Rico, for helping with the soil analysis. This work was supported in part by the U.S. Department of Agriculture (USDA), the National Institute of Food and Agriculture, and the Hatch project H94G. The authors declare that they have no conflict of interests.

References

- Abbasi, I. H. R., Abbasi, F., Abd El-Hack, M. E., Abdel-Latif, M. A., Soomro, R. N., Hayat, K., Mohamed, M. A. E., Bodinga, B. M., Yao, J. and Cao, Y. (2018). Critical analysis of excessive utilization of crude protein in ruminants ration: Impact on environmental ecosystem and opportunities of supplementation of limiting amino acids—a review. *Environmental Science and Pollution Research*, 25(1), 181-190. <https://doi.org/10/gcvt3h>
- Barros da Silva, A., Brandão Carvalho, C. A., de Almeida Pires, C., de Carvalho Almeida, J. C. and de Deus Nepomuceno,

- D. (2018). Effects of nitrogen dosage and urea source on morphological composition and forage accumulation in massai grass. *Semina: Ciências Agrárias*, 39(4), 1407-1416. <https://doi.org/10.5433/1679-0359.2018v39n4p1407>
- Beinroth, F. (2000). *Land Resources for Forage Production in the Tropics*. 1st ed., CRC Press.
- Bittencourt Caldas, M., Pereira Diniz, J., da Silva, A. J., da Silva Baio, S. P., Rezende Zuffo Borges, M. C., Batista Nogueira, K., Garcia Roque, C. and Teodoro, P. E. (2020). Forms of nitrogen fertilizer application in Panicum maximum. *Bioscience Journal*, 36(1), 23-29. <https://doi.org/10.14393/BJ-v36n1a2020-39714>
- Castagnara, D. D., Mesquita, E. E., Neres, M. A., Oliveira, P., Deminici, B. B. and Bamberg, R. (2011). Nutritional value and structural characteristics of tropical grasses under nitrogen fertilization. *Archivos de Zootecnia*, 60(232), 931-942. <https://dx.doi.org/10.4321/S0004-05922011000400010>
- de Oliveira da Silva, R., Rocha Chaves Miotto, F., Neuman Miranda Neiva, J., Monteiro da Silva, L. F., Barros de Freitas, I., Araújo, V. L. and Restle, J. (2020). Effects of increasing nitrogen levels in Mombasa grass on pasture characteristics, chemical composition, and beef cattle performance in the humid tropics of the Amazon. *Tropical Animal Health and Production*, 52(6), 3293-3300. <https://doi.org/10.1007/s11250-020-02360-0>
- Delevatti, L. M., Cardoso, A. S., Barbero, R. P., Leite, R. G., Romanzini, E. P., Ruggieri, A. C. and Reis, R. A. (2019). Effect of nitrogen application rate on yield, forage quality, and animal performance in a tropical pasture. *Scientific Reports*, 9(1), 1-9. <https://doi.org/10.1038/s41598-019-44138-x>
- Dobermann, A. R. (2005). *Nitrogen Use Efficiency - State of the Art*. Agronomy & Horticulture - Faculty Publications. 6, 1-16. <https://digitalcommons.unl.edu/agronomyfacpub/316>
- Escarela, C. M., Pietroski, M., de Mello Prado, R., Silva Campos, C. N. and Caione, G. (2017). Effect of nitrogen fertilization on productivity and quality of Mombasa forage (*Megathyrus maximum* cv. Mombasa). *Acta Agronômica*, 66(1), 42-48. <https://doi.org/10.15446/acag.v66n1.53420>
- Estrada Álvarez, J., Villa Duque, N. and Henao Uribe, F. J. (2015). Digestibility of a sugarcane silage with pig manure, and its evaluation in a double-purpose cattle production system. *Pastos y Forrajes*, 38(4), 425-443. http://scielo.sld.cu/scielo.php?script=sci_arttext&pid=S0864-03942015000400006&lng=es&tlng=es
- Galindo, F., Buzetti, S., Teixeira Filho, M.C., Dupas, E. and Ziolkowski Ludkiewicz, M. G. (2017). Application of different nitrogen doses to increase nitrogen efficiency in Mombasa guinegrass (*Panicum maximum* cv. mombasa) at dry and rainy seasons. *Australian Journal of Crop Science*, 11(12), 1657-1664. <https://doi.org/10.21475/ajcs.17.11.12.pne907>
- Galindo, F. S., Beloni, T., Buzetti, S., Teixeira Filho, M. C., Dupas, E. and Ziolkowski Ludkiewicz, M. G. (2018). Technical and economic viability and nutritional quality of mombasa guinea grass silage production. *Acta Scientiarum. Agronomy*, 40(1), e36395. <https://doi.org/10.4025/actasciagron.v40i1.36395>
- Hassan, A., Zewdu T., Urge, M. and Fikru, S. (2015). Effect of Nitrogen Fertilizer Application on Nutritive Value of *Cenchrus ciliaris* and *Panicum Maximum* Grown under Irrigation at Gode, Somali Region. *Journal of Nutrition & Food Sciences*, s11, s11005. <https://doi.org/10.4172/2155-9600.s11-005>
- Hodge, A. (2005). Nitrogen in Soils: Plant Uptake. *Encyclopedia of Soils in the Environment*, 39-46. <https://doi.org/10.1016/B0-12-348530-4/00159-4>
- Hare, M. D., Phengphet, S., Songsiri, T. and Sutin, N. (2015). Effect of nitrogen on yield and quality of *Panicum maximum* cv. Mombasa and Tanzania in Northeast Thailand. *Tropical Grasslands - Forrajes Tropicales*, 3(1), 27-33. [https://doi.org/10.17138/tgft\(3\)27-33](https://doi.org/10.17138/tgft(3)27-33)
- Li, X. G., Jia, B., Lv, J., Ma, Q., Kuzyakov, Y. and Li, F. (2017). Nitrogen fertilization decreases the decomposition of soil organic matter and plant residues in planted soils. *Soil Biology and Biochemistry*, 112, 47-55. <https://doi.org/10.1016/j.soilbio.2017.04.018>
- Maltas, A., Kebli, H., Oberholzer, H. R., Weisskopf, P. and Sinaj, S. (2018). The effects of organic and mineral fertilizers on carbon sequestration, soil properties, and crop yields from a long-term field experiment under a Swiss conventional farming system. *Land Degradation and Development*, 29(4), 926-938. <https://doi.org/10.1002/ldr.2913>
- Mariano, E., de Sant Ana Filho, C. R., Bortoletto-Santos, R., Bendassolli, J. A. and Trivelin, P. C. O. (2019). Ammonia losses following surface application of enhanced-efficiency nitrogen fertilizers and urea. *Atmospheric Environment*, 203, 242-251. <https://doi.org/10.1016/j.atmosenv.2019.02.003>
- Martínez Dalmau, J., Berbel, J. and Ordóñez Fernández, R. (2021). Nitrogen Fertilization. A Review of the Risks Associated with the Inefficiency of Its Use and Policy Responses. *Sustainability*, 13(10), 5625. <https://doi.org/10.3390/su13105625>
- Muñoz, M. A., Lugo, W. I., Santiago, C., Matos, M., Ríos, S. and Lugo, J. (2018). Taxonomic classification of the soils of Puerto Rico, 2017. Agricultural Experiment Station, Bulletin 313. https://www.uprm.edu/tamuk/wp-content/uploads/sites/299/2019/06/Taxonomic_classification_soils_PR_2018_reduced.pdf
- Nelson, D.W. and Sommers, L.E. (1996). Total Carbon, Organic Carbon, and Organic Matter. *Methods of Soil Analysis: Part 3, Chemical Methods*, 5.3. <https://doi.org/10.2136/sssabookser5.3.c34>
- Norman, H.C., Humphries, A.W., Hulm, E., Young, P., Hughes, S.J., Rowe, T., Peck, D.M. and Vercoe, P.E. (2021). Productivity and nutritional value of 20 species of perennial legumes in a low-rainfall Mediterranean-type environment in southern Australia. *Grass and Forage Science*, 76(1), 134-158. <https://doi.org/10.1111/gfs.12527>
- Norton, B.E., Barnes, M. and Teague, R. (2013). Grazing Management Can Improve Livestock Distribution: Increasing accessible forage and effective grazing capacity. *Rangelands*, 35(5), 45-51. <https://doi.org/10.2111/RANGELANDS-D-13-00016.1>
- Olivastro Teixeira, S., Olivastro Teixeira, R., Bezerra dos Santos, V., de Carvalho, M. A. C. and Mitsuo Yamashita, O. (2018). Doses de fósforo e nitrogênio na produção de *Brachiaria* híbrida cv. Mulato II. *Revista Ceres*, 65(1), 28-34. <https://doi.org/10.1590/0034737X201865010005>
- Perera, R.S., Cullen, B.R. and Eckard, R.J. (2019). Growth and Physiological Responses of Temperate Pasture Species to Consecutive Heat and Drought Stresses. *Plants* 8(7), 227. <https://doi.org/10.3390/plants8070227>
- Prosser, J.I. (2005). Nitrogen in Soils: Nitrification. *Encyclopedia of Soils in the Environment*, 31-39. <https://doi.org/10.1016/B0-12-348530-4/00512-9>
- Reimer, M., Hartmann, T. E., Oelofse, M., Magid, J., Bünenmann, E. K. and Möller, K. (2020). Reliance on Biological Nitrogen Fixation Depletes Soil Phosphorus and Potassium Reserves. *Nutrient Cycling in Agroecosystems*, 118(3), 273-291. <https://doi.org/10.1007/s10705-020-10101-w>
- Sakiroglu, M., Dong, C., Hall, M. B., Jungers, J. and Picasso, V. (2020). How does nitrogen and forage harvest affect

- belowground biomass and nonstructural carbohydrates in dual-use Kernza intermediate wheatgrass? *Crop Science*, 60(5), 2562-2573. <https://doi.org/10.1002/csc2.20239>
- SAS Institute Inc. (2021). *SAS/IML® 14.1 User's Guide*. Cary, NC: SAS Institute INC. <http://support.sas.com/documentation/cdl/en/imlsug/68152/HTML/default/viewer.htm#titlepage.htm>
- Salinas, J. G. and Valencia G., C. A., 1983. *Oxisoles y ultisoles en America Latina tropical*. Cali, Colombia. Centro Internacional de Agricultura Tropical (CIAT). <https://cgspace.cgiar.org/handle/10568/54508>
- Sparks, D. L., Page A. L., Helmke, P.A., Loeppert R. H., Soltanpour, P. N., Tabatabai, M. A., Johnston, C. T. and Summer M. E. (1996). *Methods of Soil Analysis: Part 3 Chemical methods*, 5.3. Soil Science Society of America, Inc., American Society of Agronomy, Inc. <https://doi.org/10.2136/sssabookser5.3>
- Sumiyoshi, Y., Crow, S. E., Litton, C. M., Deenik, J. L., Taylor, A. D., Turano, B. and Ogoshi, R. (2016). Belowground impacts of perennial grass cultivation for sustainable biofuel feedstock production in the tropics. *GCB Bioenergy*, 9(4), 694-709. <https://doi.org/10.1111/gcbb.12379>
- Sun, X., Longhurst, B., Luo, J. and Luo, N. (2008). Fertiliser Nitrogen and Factors Affecting Pasture Responses. *The Open Agriculture Journal*, 2(1), 35-42. <https://doi.org/10.2174/1874331500802010035>
- USDA, 2014. *Informe Anual de la Oficina para la Reglamentación de la Industria Lechera*. Instituto de Estadísticas de Puerto Rico. <https://estadisticas.pr/en/inventario-de-estadisticas/informe-anual-de-la-oficina-para-la-reglamentacion-de-la-industria-lechera>
- Teague, R., Provenza, F., Kreuter, U., Steffens, T. and Barnes, M. (2013). Multi-paddock grazing on rangelands: Why the perceptual dichotomy between research results and rancher experience? *Journal of Environmental Management*, 128, 699-717. <https://doi.org/10.1016/j.jenvman.2013.05.064>
- Van Soest, P. J. (1994). *Nutritional ecology of the ruminant*. 2^a Ed. Cornell University Press.
- Vicente-Chandler, J. (1964). *The Intensive Management of Tropical Forages in Puerto Rico*. Agricultural Experiment Station. 187, 1-152. <https://books.google.cl/books?id=Xql6HAAACAAJ>
- Zang, H., Wang, J. and Kuzyakov, Y. (2016). N fertilization decreases soil organic matter decomposition in the rhizosphere. *Applied Soil Ecology*, 108, 47-53. <https://doi.org/10.1016/j.apsoil.2016.07.021>
- Zhang, S., Fang, Y., Luo, Y., Li, Y., Ge, T., Wang, Y., Wang, H., Yu, B., Song, X., Chen, J., Zhou, J., Li, Y. and Chang, S. X. (2021). Linking soil carbon availability, microbial community composition and enzyme activities to organic carbon mineralization of a bamboo forest soil amended with pyrogenic and fresh organic matter. *Science of the Total Environment*, 801, 149717. <https://doi.org/10/gpb2km>