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ARTÍCULO DE INVESTIGACIÓN / RESEARCH ARTICLE

**CONTRIBUTION TO THE KNOWLEDGE OF DESMIDS
(ZYGNEMATOPHYCEAE, STREPTOPHYTA) IN A TROPICAL LAKE
(COLOMBIA)**

Contribución al conocimiento de las Desmidias (Zygnematophyceae, Streptophyta) en un lago tropical (Colombia)

Running head: Desmids species in a tropical lake, Colombia

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ABSTRACT

I here describe 24 taxa of desmids from the Colombian Oriental Plains (Orinoquía). *Docidium undulatum* var. *dilatatum* (Cleve) West & G.S. West; *Micrasterias arcuata* var. *robusta* O. Borg; *Micrasterias radiata* var. *gracillima*. G.M. Smith; *Micrasterias torreyi* var. *curvata*. Willi Krieger; *Pleurotaenium constrictum* var. *laeve* Irénée-Marie; *Pseudomicrasterias arcuata* (Bailey) C.B. Araujo, C.E.M. Bicudo, Stastny & Skaloud; *Spondylosium pulchrum* (Bailey) W. Archer 1861; *Staurodesmus wandae* var. *longissimus* (Borge) Teiling, and *Staurastrum subindentatum* var. *brasiliense* Borge 1918, are reported for the first time in Colombia. The richness and coexistence of desmid species found are remarkable because they are bioindicators of oligotrophic conditions, geographic isolation, and well-developed littoral zone properties in the lakes of the Orinoquía. Desmids are early warning indicators of the significance of this environment for biodiversity and conservation.

Keywords: Bioindicators, Carimagua, Orinoco, Species, Tropical Savanna

RESUMEN

Describo 24 taxones de desmidias de un lago de los Llanos Orientales de Colombia (Orinoquía). *Docidium undulatum* var. *dilatatum* (Cleve) West & G.S. West; *Micrasterias arcuata* var. *robusta* O. Borg; *Micrasterias radiata* var. *gracillima*. G.M. Smith; *Micrasterias torreyi* var. *curvata*. Willi Krieger; *Pleurotaenium constrictum* var. *laeve* Irénée-Marie; *Pseudomicrasterias arcuata* (Bailey) C.B. Araujo, C.E.M. Bicudo, Stastny & Skaloud; *Spondylosium pulchrum* (Bailey) W. Archer; *Staurodesmus wandae* var. *longissimus* (Borge) Teiling, and *Staurastrum subindentatum* var. *brasiliense* Borge 1918, se registran por primera vez para Colombia. La riqueza y coexistencia de especies de

desmicias es notable porque son bioindicadores de condiciones oligotróficas, aislamiento geográfico y propiedades de la zona litoral bien desarrollada en los lagos de la Orinoquía. Las desmicias son indicadores del valor de la biodiversidad y la conservación de estos ambientes acuáticos.

Palabras claves: Bioindicadores, Carimagua, Orinoco, Especies, Sabana Tropical

INTRODUCTION

The littoral zones of freshwater aquatic ecosystems are key habitats for biodiversity (Domozych & Domozych, 2008). These habitats host benthic-attached microbial communities and tychoplanktonic communities, which together contribute to primary productivity, substratum stability, and aquatic biodiversity. Desmids are loosely associated with macrophytes of the littoral zone and the bottom sediment and are indicative of oligotrophic and poorly mineralized environmental conditions (Gonzalez et al., 2019). In these environments, desmids may respond to environmental stresses and disturbances and then contribute to the community's resilience (Lyons, 2001).

The great majority of desmids taxa are of incomparable beauty and have wide geographic distributions, especially in tropical and subtropical areas. They inhabit almost exclusively freshwater aquatic environments and are characteristic of oligotrophic aquatic bodies that are well-oxygenated, of optimal ecological conditions, and with a slightly acidic pH, which makes these organisms excellent ecological indicators of water quality (Coesel, 1996).

Desmids do not contribute substantially to the phytoplankton diversity (Coesel, 1982) but

are associated with aquatic plants and lake sediments (Pals et al., 2016) and represent a diverse and abundant component especially in ombrotrophic peatlands (Neustupa et al., 2012).

Several studies already show that Colombia, together with Brazil, is a hot spot for desmids diversity. More than 500 species have been registered and described for the country (West & West, 1896; West, 1914; Taylor, 1935; Coesel et al., 1986; Coesel et al., 1987; Coesel et al., 1988; Coesel, 1992; Coesel, 1997). Although high mountain lakes have a particular richness and diversity of desmids species (Taylor, 1935; Donato et al., 1987; González & Mora-Osejo, 1996), the aquatic ecosystems of the Orinoquia and Amazonia contain uncommon, and interesting species (Duque & Donato, 1993; 1995a; 1995b; 1996a; 1996b; Coesel, 1992; Nuñez-Avellaneda & Duque, 2000).

The aquatic ecosystems located in the tropical savannas of the Colombian Orinoquia are characterized by a high diversity of algae, particularly desmids and diatoms. Neotropical savannas conform to an ecological-physiognomic type of natural ecosystem that is found exclusively in warm and humid tropical areas (the low-altitude wet tropics) (Sarmiento, 1984). They occupy approximately 20 % of the earth's surface and 45 % corresponds to those located in South America (Sarmiento, 1984).

The savannas are mainly located in Brazil, Venezuela, and Colombia, and dominate the landscape in the plains that surround the Orinoco River, the so-called *llanos* (eastern plains). The eastern plains between Colombia and Venezuela cover a surface of approximately half a million square kilometers, constituting the largest uninterrupted surface of neotropical savanna north of the Equator (Sarmiento & Pinillos, 2001).

Within this complex of the middle basin of the Orinoco River, two types of aquatic ecosystems can be differentiated: i) lakes or lagoons located in the alluvial plains, known as overflow lagoons or floodplains; they are of a temporary nature and are completely dependent on the contribution direct from the main riverbed (Correa et al., 2005); and ii) wetlands located in the alluvial terraces that are fed either by rains (seasonal) or by some streams and aquifers (Correa et al., 2005). These wetlands are permanent elements in the Orinoco landscape. Among them, we find estuaries, *morchales* (*Mauritia flexuosa*), lakes, or small artificial reservoirs.

The littoral zones of these lakes contain a vast diversity of microorganisms associated with aquatic plants and are the habitats that mainly contribute to the richness, diversity, and productivity of oligotrophic systems (Frankovich et al., 2006). Several hypotheses explain the growth of epiphytic communities and the supporting role of aquatic plants in aquatic environments. The neutral substrate hypothesis (Blindow, 1987) postulates that the epiphytic structures do not interact biologically or chemically with the host aquatic plant. However, Carignan & Kalff, (1982), Mutinová et al., (2016), and Eminson & Moss (1980), highlight that macrophytes may transfer inorganic nutrients to epiphytic algae. In contrast, aquatic plants may also hurt epiphytic communities through the production of allelopathic substances or by facilitating the action of predators (Hilt, 2006).

Identification of species is extremely important to properly define the structure and dynamics of biological communities (Santos et al., 2018). The current research aimed to describe and report poorly common species of desmids, the majority of exclusive tropical distribution, found in a lowland lake located in the tropical savanna of the Colombian

Orinoquía. I suggest that expanding the current knowledge of these taxa contributes to the identification of areas for the protection of biodiversity and the conservation of reference ecosystems.

MATERIALS AND METHODS

Study Area

The eastern tropical savanna plains of Colombia are characterized by three landscape units: foothills of the Eastern Cordillera, flood plains, and high plains. The *llanos orientales* of Colombia correspond to the low relief region located east of the foot of the eastern mountain range and comprise approximately 53 % of the area of Colombia. Its formation began in the Andean orogeny (IGAC, 1991).

This study was performed in the Carimagua Lake (Department of Meta) which lies in the *llanos orientales* and is located 4°34'36,3"N 71°20'18,9"W at an altitude of 172 m a.s.l. It is surrounded by open savanna vegetation and only a few patches of *Mauritia* – a dominating palm forest reaching to the shore -.

Carimagua Lake consists of about 480 ha and its name comes from a community of Indigenous Guahiba and signifies "*source of water*". It is a tropical freshwater lake with high transparency, oligotrophic, and a littoral zone that borders the lake with aquatic plants such as *Utricularia*, *Eleocharis*, *Cabomba*, *Nymphaea*, *Limnobium*, *Eichornia*, *Eriocaulon*, *Mayaca*, *Cyperus*, among other.

The climate is characterized by a monomodal regime with a dry period from November to March and a rainy season from April to November. According to the Köppen- Geiger classification *Aw* type or *savanna climate* predominates: a slightly rainy and humid tropical climate. Based on the records of the Carimagua Research Station the local mean annual precipitation is 1860 mm. The mean annual temperature is 26 – 27°C with less than 3°C variation between monthly means (Berrio et al., 2000).

The predominant vegetation is savanna (85 %) and forest (15 %). In the savannas *Paspalum* sp., *Andropogon* sp., *Panicum* sp., *Axonopus* sp., are the predominant grasses; while the shrubs *Curatella americana*, *Byrsonima crassifolia* and *Palicourea rigida* are present. The soils are iron-based armor formations (Latosols). The soils have moderately high organic matter and very poor drainage capacity (Goosen, 1971). The predominant plintite stones are characterized by having iron in the oxidized state.

FIELDWORK

An intensive sampling was performed during the dry season (March). Data on the physical and chemical characteristics of the water and on the desmids, and communities were collected. A total of 12 samples were obtained from the littoral zone and limnetic zone of the lake.

Samples of aquatic plants (*Utricularia*, *Cabomba*, *Nymphaea*, *Limnobium*, *Eichornnia*, *Mayaca*), allowing the development of rich varied tycho plankton assemblages were collected separately for study in the desmid's community. Samples from the limnetic zone

were obtained with a 26 µm plankton mesh. Samples after squeezing and planktonic were preserved in Transeau solution (1:1).

Water samples were collected in the field in triplicate (open water and littoral zone) for immediate measurement of pH, conductivity, temperature, alkalinity, and dissolved oxygen. A Hach Seinson 176 probe was used in the field. The concentrations NO_3^- and NO_2^- were determined separately, following standard APHA (1998) methods. Soluble reactive phosphorus (SRP) was measured in GF/F filtered samples, using the molybdenum-blue method (details also in APHA, 1998).

LABORATORY WORK

Regional keys were used for taxonomic identification, such as Förster (1964, 1969, 1974), Teiling (1967), Prescott (1966), Prescott et al., (1975, 1977, 1982), Coesel & Meesters (2007). The validity of the taxa was verified using Algaebase (Guiry and Guiry, 2018). Scanning electron microscopy (SEM) observations were performed on samples placed on acetone-washed glass coverslips (10 or 12 mm in diameter), then they were dried using a poly-L-Lysine solution. After fixation, it was transferred to an acetone series. Finally, the cells were dried to a critical – point with liquid CO_2 , and subsequently sputter-coated with gold and examined using a Transmission Electron Microscope ZEISS EM910.

RESULTS

GENERAL CHARACTERISTICS OF THE LAKE

Carimagua is a shallow lake (139 cm) of high temperatures (30.1 °C), very low conductivity ($\geq 5.9 \leq 8.61 \mu\text{S cm}^{-1}$), acidic pH ($\geq 4.67 \leq 5.41 [\text{H}^+]$), alkalinity ($\geq 12.0 \leq 15.0 \text{ mg l}^{-1}$), relatively high concentrations of dissolved oxygen ($\geq 6.72 \leq 7.57 \text{ mg l}^{-1}$), percentage of oxygen ($\geq 95.5 \leq 104.6 \text{ mg l}^{-1}$) and low nutrient content, especially PRS ($\geq 0.09 \leq 0.12 \text{ mg l}^{-1}$), NO_3^- ($\geq 0.12 \leq 0.15 \text{ mg l}^{-1}$) and NO_2^- ($\geq 0.08 \leq 0.03 \text{ mg l}^{-1}$).

ALGAL FLORA

The dominant algal group was Diatoms, followed by Cyanobacteria and Desmidiaceae family. The most abundant genera of diatoms were *Navicula*, *Gomphonema*, *Tabellaria*, *Frustulia*, and *Pinnularia*. The most common Cyanobacteria were *Anabaena* and *Aphanocapsa*. The dominant Desmidiaceae genera were *Staurastrum* sp and *Cosmarium* sp.

IDENTIFICATION AND DESCRIPTION OF DESMIDS TAXA

We taxonomically identified 24 species of taxa belonging to 14 genera and 2 families. The majority of the species were part of the genera, *Pseudomicrasterias* (1), *Micrasterias* (5), *Staurastrum* (3), *Cosmarium* (2), *Euastrum* (2), and *Closterium* (1) while the other genera are 10 of the total amounts of the species.

Bambusina borreri (Ralfs) Cleve 1864

Articulated filiform filament (Fig. 1a). Cell longer (78-90 μm) than wide (32 - 38 μm). Semi - barrel-shaped cells (amphora) are slightly wavy in the middle part and thickened in the apical part with a truncated termination (Fig. 1b). Cell wall with signs of longitudinal striae (Fig. 1b) and with remains of mucilage. Isthmus (18-20 μm) slightly domed. Strongly welded junctions between cells, as wide as they are long (Fig. 1b). According to Coesel & Meesters (2007) cell wall, particularly in the apical part of the semicell, delicately striata (rows of pores).

Closterium porrectum Nordstedt 1870

Strongly curved (lunate) cell of medium size, cell longer than wide (Fig. 1c). Semi-cells with visible streaks or ribs (up to 8 in the frontal view) that reach the isthmus (Fig. 1d). Slightly rounded apices. L: 360-370 μm ; W: 26-30 μm .

Cosmarium connatum Brébisson ex Ralfs 1848

Large cells, longer (45-50 μm) than wide (35-40 μm). Elliptical semicells and rounded apex (Fig. 1e). Cell wall perforated by pores without ornamentation but arranged radially and symmetrically. Isthmus (30- 32 μm) closed by a circular structure (Fig. 1f).

Cosmarium polymorphum Nordstedt 1870

Cells longer (30 - 35 μm) than wide (24-26 μm). Semicells truncated at the polar apex and at the base with limits of the isthmus (Fig. 1g). Wall perforated by finely ornamented pores and numerous protrusions (more than 10 per half-cell) (Fig. 1h). Isthmus (10 - 12 μm), narrow and open to the outside (Fig. 1i).

Docidium undulatum var. *dilatatum* (Cleve) West & G.S. West 1904

Cylindrical cells longer (250-300 μm) than wide (10-14 μm); semi-cells with 7-8 undulations along each margin; apices dilated, truncated, with rounded angles (Fig. 2a). Junction at the base of the isthmus showing a very prominent ring of 9 to 10 folds (Fig. 2b). Smooth cell wall. First report for Colombia.

Cosmarium crenatum Ralfs ex Ralfs 1841

Medium-sized cell, longer than wide. Sinus closed, open at the end (Fig. 2c). Lobed semicells. Lower lateral lobes rounded by a flat cleft. Truncated apex with a narrow central incision (Fig. 2c). Truncated polar lobes with rounded angles. The bumps are sculpted by pores. L: 65-70 μm . W: 25-30 μm . I: 9-11 μm .

Euastrum evolutum (Nordstedt) West & G.S. West 1896

Medium-sized cell, longer than wide (Fig. 2d). Narrow sinus, dilated to the outside. The trapezoidal. Basal corners with spines. Spiny upper and lower lateral lobes, by a separate incision and with evident and deep holes (Fig. 2e). Protruding apex, truncated, with deep

central incision. Polar lobes with long terminal spine and short spines (Fig. 2e). On the sides of the polar lobes there are 2 spiny appendages. Structure in the middle part magnified and triangular with 3 protrusions, one of them the prominent central one above 2 central pores separated by another protrusion that supports the base. L: 48-50 μm . Width: 32-35 μm . I: 10-12 μm .

Hyalotheca dissiliens Brébisson ex Ralfs 1848

Cylindrical cells, longer than wide, with a cleft between the half cells that converges in a wide and evident constriction (Fig. 2f). Medium breast. Long cylindrical filaments with more than 10 cells (Fig. 2g). Mucilaginous sheath remains present.

L: 25-28 μm . W: 20-22 μm . I: 14 -16 μm .

Mateola curvata (Nordstedt) Coesel 1997

Quadrangular cells with 4 vigorous projections ending in a simple spine (Fig. 2h). Cells wider (including spines) than long (Fig. 2i). Tubules that firmly bind longer than wide filaments. Twisted filaments between 12 to 15 cells. Profuse mucilaginous sheath.

L: 23-25 μm . W: 20-23 μm .

Pseudomicrasterias arcuata (Bailey) C.B. Araujo, C.E.M. Bicudo, Stastny & Skaloud 2022

Quadrangular cells. Smooth cell wall. Isthmus closed and prominent open to the outside (Fig. 3a). Semi-cells with slender and elongated basal and polar lobes. The basal ones have an ascending curve that brings their tips closer to the tips of the polar lobes (Fig. 3a). Pantropical distribution. L: 60-80 μm . W: 60-90 μm . I: 10-12 μm . First report for Colombia.

Micrasterias arcuata var. *gracilis* West & G.S.West 1896

Cells symmetrically arranged, wider than long. Basal lobes longer and so thin than the polar ones. Basal lobes, parallel to the polar lobe, exceed it in length (Fig. 3b).

L: 60 - 80 μm . W: 80 - 100 μm . I: 10m -12 μm .

Micrasterias arcuata var. *robusta* O. Borge 1899

Ends of the basal lobes and the polar lobes directed upwards sharply (Fig. 3c), the pointed polar cuneate lobes. L: 55–60 μm . W: 40–42 μm . I: 7-8 μm . First report for Colombia.

Micrasterias borgei var. *aequalis* Willi Krieger 1939

Flat cell, longer than wide (Fig. 3d). Hemisoma with 5 lobes. Deep sinus. Prominent polar lobe, both in it and in the basal lobes with robust spines arranged laterally, symmetrically (Fig. 3e) and directed upwards. Forked lobes ending in sharp forked tips. Ornate cell wall. (Fig. 3f). L: 140-160 μm . W: 130-140 μm . I: 20 - 25 μm . First report for Colombia.

Micrasterias radiata var. *gracillima*. G.M. Smith 1922

Cells with a globose shape, especially at the border with the isthmus (Fig. 3g). Smooth cell wall. Semi-cells bulging at the limit of the isthmus. Isthmus closed but expanded outward. The basal lobes of the semicells thin, symmetrically tapered and forked. Slightly convergent polar lobe. L: 160 (with lobules); 66.2 (no lobules) μm ; W: 103.3 (with lobules); 43.1 (no lobules) μm . I: 30 μm . First report for Colombia.

Micrasterias torreyi var. *curvata*. Willi Krieger 1939

Relatively large cells with a highly constricted linear isthmus then open outward (Fig. 3h). Semicircular cells with five lobes; the basal lobe divided into two lobes; the cuneate polar lobe. Lobes bifurcated, with a lateral spine shorter than the prominent one (Fig. 3i). L: 210-220 μm . W: 220-226 μm . I: 35-38 μm . Exclusive South American or tropical distribution. First report for Colombia.

Pleurotaenium constrictum var. *laeve* Irénée-Marie 1954

Slightly curved cylindrical cells (Fig. 4a), longer than wide, with an inflated base and ornamented by a membrane that protrudes from the base (Fig. 4b). Semi cells with dotted ornamentation (Fig. 4b), wavy wall; apex with 2 tubercles, one at each angle. L: 310-320

μm . W: 25-30 μm . I: 20-22 μm . Exclusive South American or tropical distribution. First report for Colombia.

Spondylosium pulchrum (Bailey) W. Archer 1861

Cells flattened; apices truncated. Open isthmus (Fig. 4c). Well-marked breast. Cells united by the apposition of the apices in filaments. Medium deep constriction (Fig. 4d). Tubes connecting prominent filament cells as long as wide. First report for Colombia.

L: 35-38 μm . W: 45-48. I: 15-16 μm .

Staurodesmus calyxoides (Wolle) Croasdale 1957

Cells longer (40-43 μm) than wide (26-30 μm) (not including spines). Isthmus 18-20 μm . Semi-cells with 6 long spines (hexaradial symmetry) and robust in various directions (Fig. 4e). Ventral margin of the half-cells domed or slightly curved. Uniformly dotted cell wall. Isthmus completely closed. Exclusive South American or tropical distribution.

Staurodesmus wandae var. *longissimus* (Borge) Teiling 1966

Cells longer (76 - 78 μm not including spines; 96 - 98 μm with spines) than wide (50 – 60 μm with spines) (Fig. 4f). Symmetrically dotted cuneate semicells. Isthmus broad (15 -16 μm), and open. Simple and smooth pores without ornamentation (Fig. 4g). Slightly curved spines emerge at the angles on bumps (Fig. 4h). Length of cells including spines, width of

cells including spines. Exclusive South American or tropical distribution. First report for Colombia.

Staurastrum leptacanthum var. *borgei* Kurt Förster 1969

Longer than wide triangular cell (without processes). The circular apical view. Long processes forming a medium ring with numerous processes (nine). Processes with two spines (bifurcated) at the top (Fig. 4i). L: 45 -50 μm without processes. W: 35-40 μm . I: 20-23 μm . Wide geographical distribution.

Staurastrum radians West & G.S.West 1898

Cells almost twice as wide as long and with processes (Fig. 5a). Smooth cell wall. Semicells with eight arms (processes) arranged symmetrically and radially (Fig. 5b). Tridentate processes at the apex highly ornamented with spines and other protrusions. Isthmus closed slightly open outwards. L: 34- 35 μm . W: 82-84. I: 16-18.

Staurastrum subindentatum var. *brasiliense* Borge 1918

Cells longer (100-110 μm including processes) than wide (32 – 35 μm). Cell wall with smooth, finely distributed pores (Fig. 5c). In each semicell, a pair of long denticulate and arcuate processes arise that end in two spines. A pair of bumps on the lamina of each half-cell (Fig. 5d). In the upper straight plane of the semicell two spines at the ends. Narrow

sinus with dilations at the end. Diffuse gelatinous sheath. Isthmus 8- 10 μm . Exclusive South American or tropical distribution. There is only one report for Brazil. First report for Colombia.

Triploceras gracile Bailey 1851

Cells longer (38-40 μm) than wide (2.5-3.5 μm) (Fig. 5e). Semi-cells that taper towards the apex and end in three smooth spines of equal size (Fig. 5f). Smooth cell wall. Dispersed mucilaginous sheath.

Xanthidium regulare Nordstedt 1870

Cells longer (100-110 μm with process) than wide (60 - 65 μm with process). Hexagonal elliptical semicells (front view) with simple spines at each angle which is projected (Fig. 5g). In the equatorial plane and in the center of the semicell it is ornamented (dotted) and a spine starts in line from the same axis. Deep median constriction. Isthmus closed. Exclusive South American or tropical distribution.

DISCUSSION

The Carimagua lake, hydrologically isolated, oligotrophic, and located in the tropical savanna of the Colombian Orinoquia, is habitat of many of them (*Docidium undulatum* var. *dilatatum*; *Euastrum evolutum*; *Mateola curvata*, *Micraterias arcuata* var. *robusta* and

Micrasterias arcuata var. *gracilis*) and interesting as *Micrasterias torreyi* var. *curvata*, *Pleurotaenium constrictum* var. *laeve*, *Staurodesmus wandae* var. *longissimus*, *Pseudomicrasterias arcuate*, *Staurodesmus calyxoides* and *Staurastrum subindentatum* var. *brasiliense*.

Carimagua lake offers the perfect location for the development of Zygnematophyceae species. High temperatures, oligotrophy, moderate size, and shallow depth facilitate the development of the littoral zone that functions as a patch of high species richness (Scheffer et al., 2006). It is remarkable that this relatively small, shallow, and isolated tropical lakes develop abundant macrophyte vegetation, with the increase in the number of associated species, particularly desmids.

The richness and coexistence desmid species are striking; many of them are picoplanktonic. These species, despite their low abundance, contribute to maintaining the properties (interactions) and functioning of the ecosystem and provide a buffer against environmental disturbances, increasing the resilience of the ecosystem (Walker et al., 1999). Likewise, these rare species are indicators of habitats with a limited supply of nutrients, low conductivity, and acidic pH (Coesel, 1982).

Finally, to counteract the threats to aquatic biodiversity, especially rare species and bioindicators, the management and handling strategies of oligotrophic lakes of moderate size, shallow and with aquatic vegetation must be prioritized. In conclusion, this dataset of floristic record including several relatively little known desmid taxa (*Mateola curvata*, *Euastrum evolutum*, *Micrasterias borgei* var. *aequalis*, *Micrasterias radiata* var.

gracillima. G.M. Smith; *Staurodesmus calyxoides*, *Std. wandae* var. *longissimus*, *Pseudomicrasterias arcuata*) with widely distributed species (e.g. *Bambusina borneri*, *Cosmarium connatum*, *Hyalotheca dissiliens*, *Xanthidium regulare*).

These species were recorded with low abundances and can be considered locally rare species and can be used as conservation indicators (Gonzalez et al., 2019) of the oligotrophic and threatened lakes of the Colombian Orinoquía.

CONCLUSIONS

The study in Lake Carimagua, in the Eastern Plains of Colombia, has revealed a remarkable diversity of desmids, with the registration of 24 taxa, of which 10 are unprecedented reports for the country. These species, indicators of oligotrophic conditions and high-water quality, underline these aquatic ecosystems' ecological importance and fragility in the region. The richness and coexistence of desmids in the lake and the presence of varied aquatic vegetation demonstrate the potential of these habitats to sustain a unique and significant biodiversity. Furthermore, the identification of these species contributes to a deeper understanding of the biodiversity of tropical savannas and reinforces the need for adequate conservation measures to protect these vulnerable ecosystems. The information obtained is crucial for future research and for the formulation of sustainable management strategies that seek to conserve the biological wealth of the Orinoquía. The study highlights the relevance of desmids as bioindicators and their role in assessing the health of aquatic ecosystems, which is vital for the conservation and management of biodiversity in this region.

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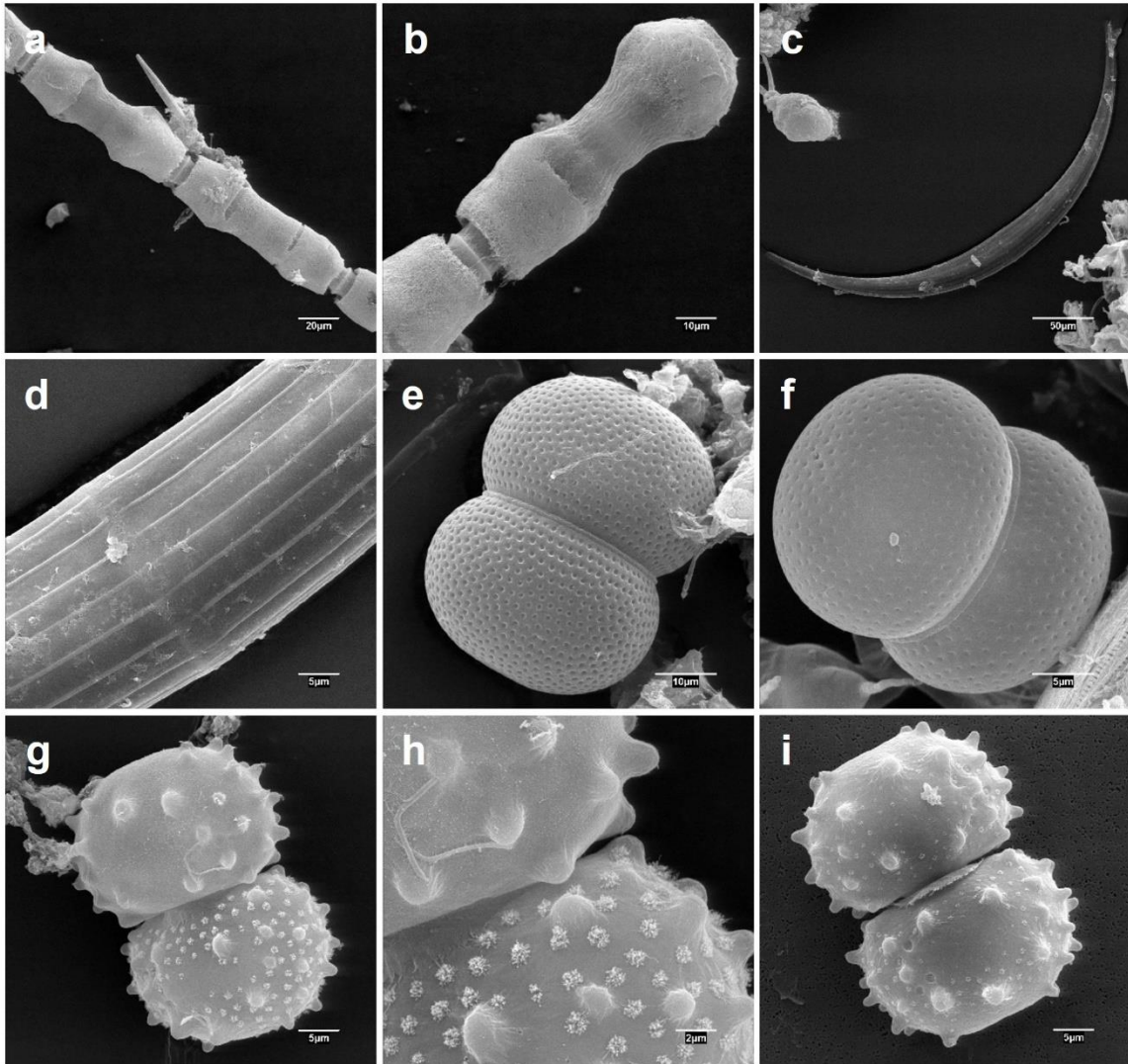


Fig. 1. *a-b. Bambusina borneri*. *a*. Detail filament. *b*. View of the semi-cells junctions. *c-d. Closterium porrectum*. *c*. General detail. *d*. Stria or ribs and the isthmus. *e-f. Cosmarium connatum*. *e*. Rounded apex of the semi-cell. *f*. Detail of the ornamentation of the cell wall and the isthmus. *g-h-i. Cosmarium polymorphum*. *g*. General view. *h-i*. Detail of the bumps and pores in the wall of the semi-cells.

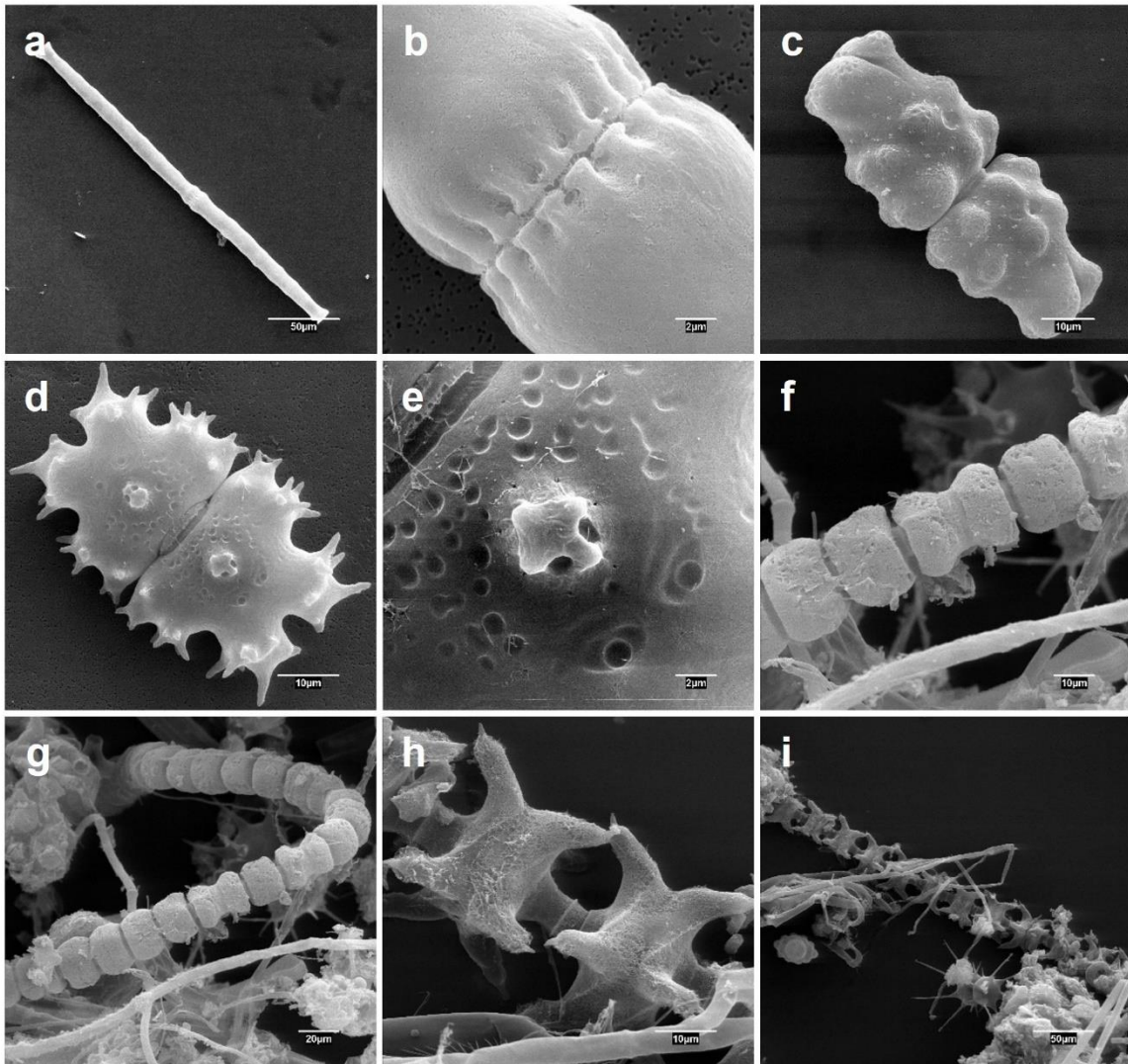


Fig. 2. *a-b. Docidium undulatum* var. *dilatatum*. a. General view. b. Detail of the folds. c. *Cosmarium crenatum*. c. Lateral view. The protrusions located on the wall of the semicells are evident. d-e. *Euastrum evolutum*. d. Front view. e. detail of the triangular structure with the protrusions and pores. f-g. *Hyalotheca dissiliens*. f. Details of semi-cells. g. filament. h-i. *Mateola curvata*. h. Detail of the cells. i. General view of the filament.

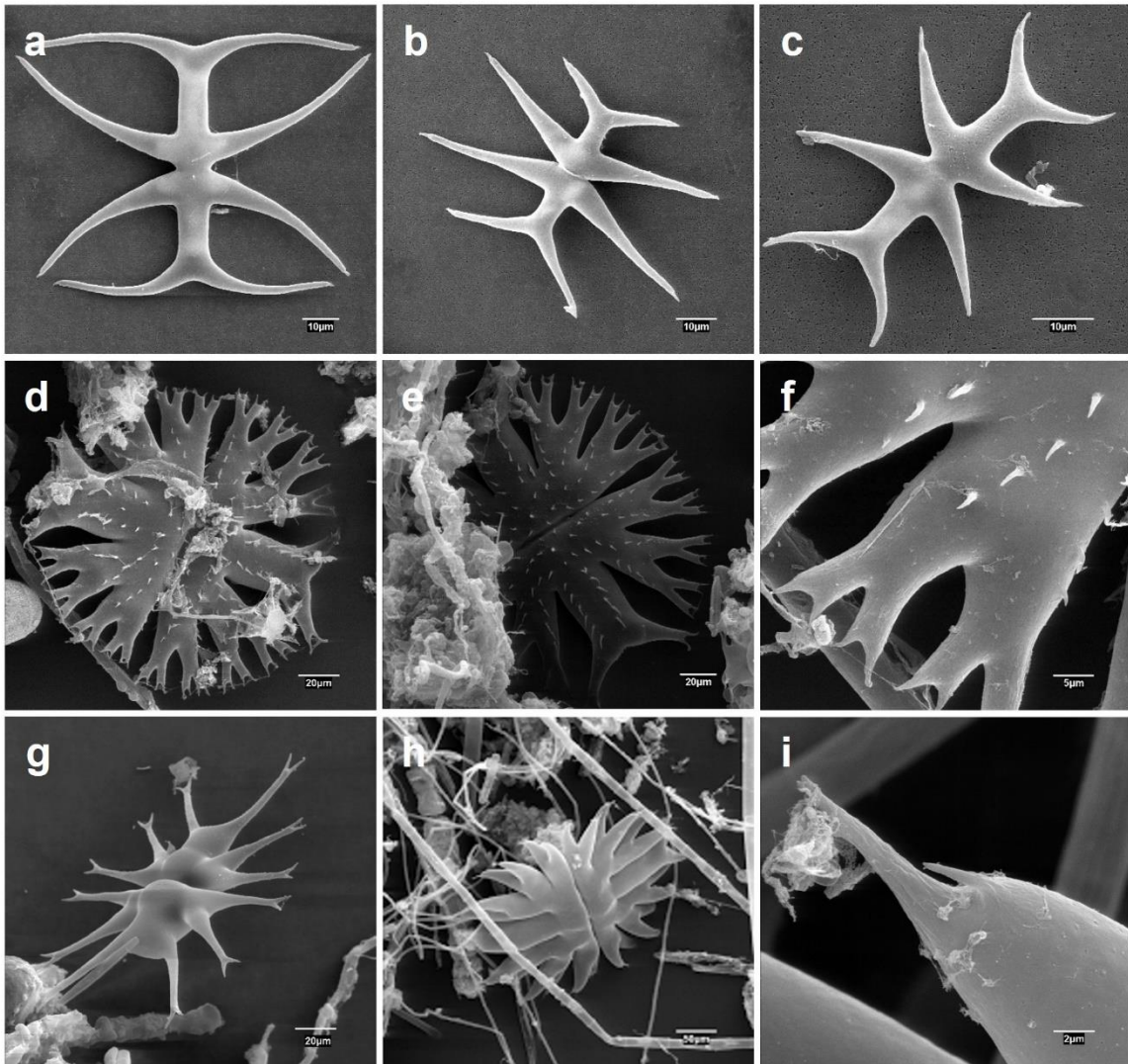


Fig. 3. a. *Pseudomicrasterias arcuata*. a. Front view. Note the ascending direction of the basal lobes. b. *Micrasterias arcuata* var. *gracilis*. b. Cells symmetrically arranged. c. *Micrasterias arcuata* var. *robusta*. c. Polar and basal lobes directed upwards. d-e-f. *Micrasterias borgei* var. *aequalis*. d. Front view. e. Detail of the arrangement of the spines. f. bifurcated lateral lobe. g-h-i. *Micrasterias radiata* var. *gracillima*. g. Smooth cell wall. h-i. *Micrasterias torreyi* var. *curvata*. h. General view. i. Lateral spine on the lamina of the lobe.

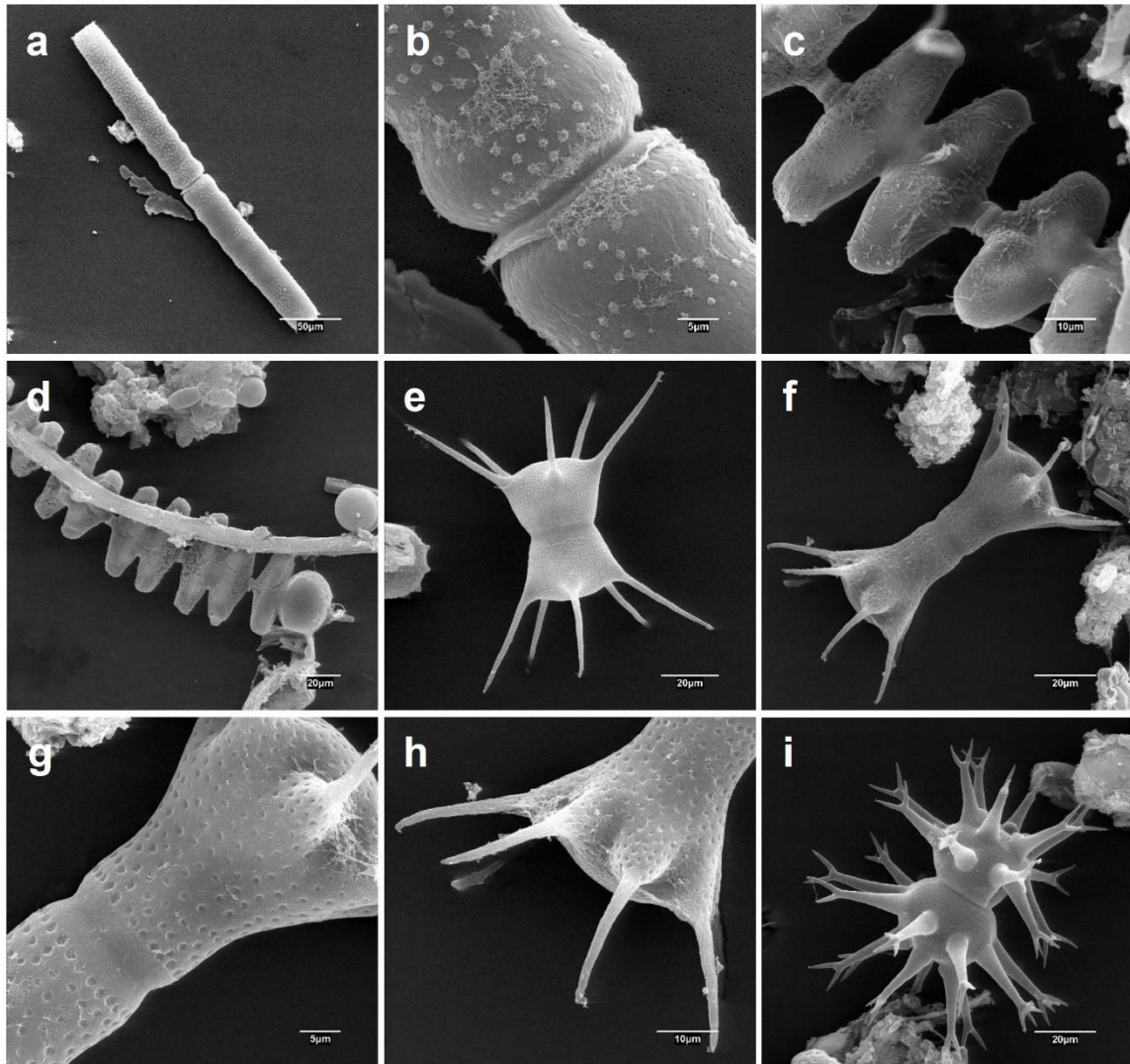


Fig. 4. a-b *Pleurotaenium constrictum* var. *laeve*. a. General view. b. Detail of the isthmus and ornate pits in the lamina of the semicells. c-d. *Spondylosium pulchrum*. c. Cells flattened. d. Filaments with mucilaginous sheath remainder. e. *Staurodesmus calyxoides*. e. View of hexarradial symmetry. f-g.h. *Staurodesmus wandae* var. *longissimus*. f. Front view. g. detail of the isthmus. h. Arrangement of the spines in the basal part of the semicell. i. *Staurastrum leptacanthum* var. *borgei*. i. Processes with two spines.

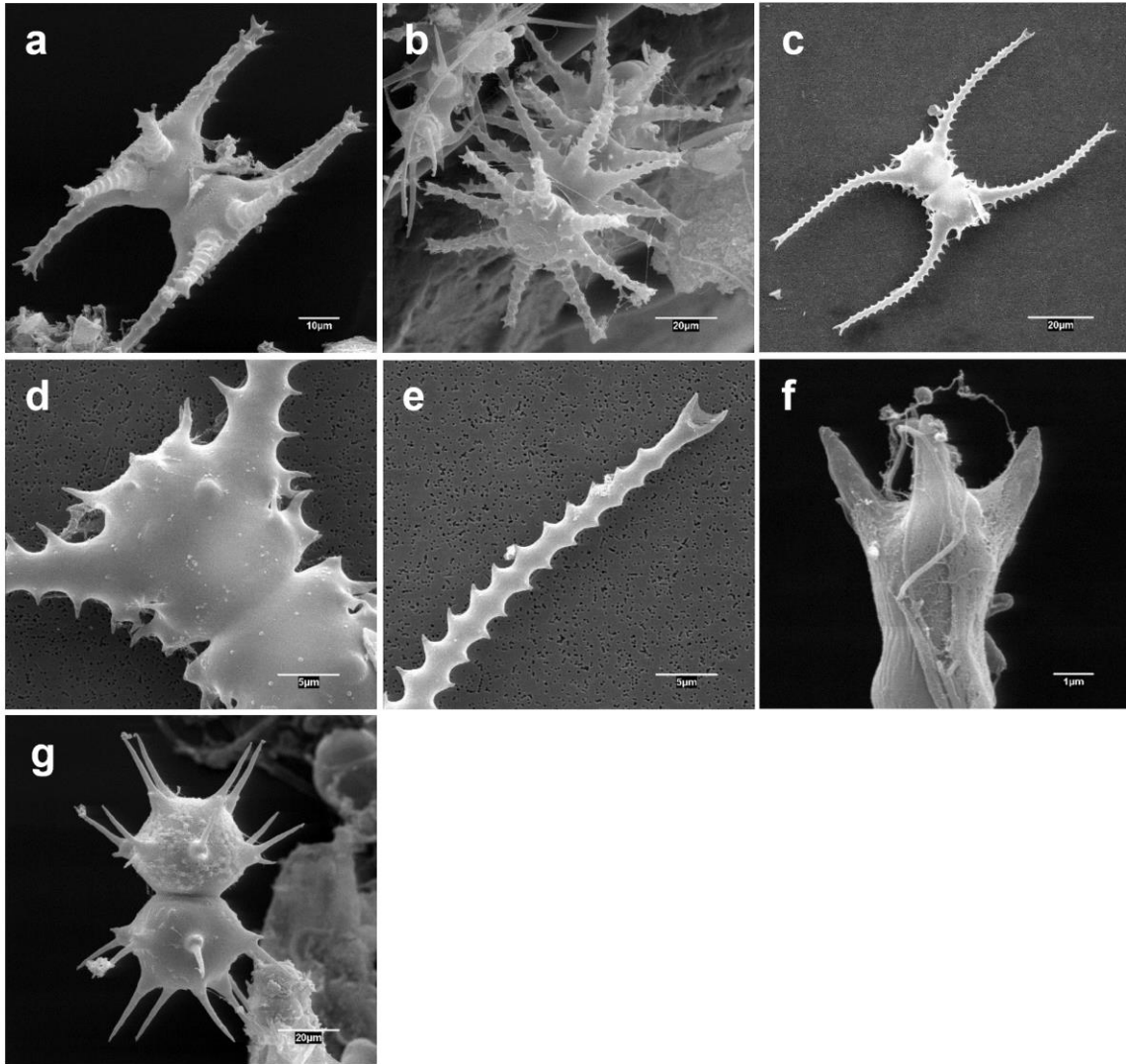


Fig. 5. a-b. *Staurastrum radians*. a. Semicells. b. Symmetrical arrangement of the ornate arms of each half cell. c-d. *Staurastrum subindentatum* var. *brasiliense*. c. Layout of the general plan. d. Detail of the spines and protrusions in the half-cell. e-f. *Triploceras gracile*. e. General view of the semicell. f. Detail of the apex showing the spines. g. *Xanthidium regulare*. g. Hexagonal elliptical semicells (front view).