

Acaricidal activity and repellency of commercial essential oils on *Tetranychus urticae* in vitro and protected cultivation

Actividad acaricida y repelencia de aceites esenciales comerciales sobre *Tetranychus urticae* in vitro y cultivo protegido

Julielson Oliveira Ataide^{1*}, Francieli Destefani Deolindo¹, Filipe Garcia Holtz¹,
Andressa Huver¹, Hugo Bolsoni Zago¹, and Luciano Menini²

ABSTRACT

This study analyzed the toxicity by residual contact and the repellency effect of the essential oils of *Rosmarinus officinalis*, *Mentha piperita*, *Melaleuca alternifolia*, and *Commiphora myrrha* and their mixtures on adult females of *Tetranychus urticae* in laboratory and protected cultivation. The essential oil of *C. myrrha* exhibited a LC_{50} of 0.55 ml L^{-1} , and the mixtures *R. officinalis* + *M. piperita* + *M. alternifolia* + *C. myrrha*, *C. myrrha* + *M. piperita*, and *C. myrrha* + *M. alternifolia* showed 68%, 60%, and 36% mortality, respectively. The essential oils of *C. myrrha* and *M. alternifolia* showed 95 and 70% of repellency, respectively. Mixtures of *C. myrrha* + *M. alternifolia*, *C. myrrha* + *M. piperita*, and *C. myrrha* + *R. officinalis* provided repellency of 85, 74 and 73%, respectively. Toxicity by residual contact of the essential oil of *C. myrrha* in protected cultivation exhibited 93% mortality, while the acaricide fenpyroximate showed 80%. The constituents of essential oils were eucalyptol (49.66%), *M. piperita* menthol (48.53%), *M. alternifolia* terpinen-4-ol (48.93%), and *C. myrrha* benzyl benzoate (97.71%). The essential oil of *C. myrrha* and the mixtures *R. officinalis* + *M. piperita* + *M. alternifolia* + *C. myrrha* and *C. myrrha* + *M. piperita* showed significant mortality. However, further studies are needed to assess the cost/benefit ratio and the effects on non-target organisms.

Key words: spider mite, mixtures, toxicity.

RESUMEN

El presente estudio analizó la toxicidad por contacto residual y el efecto repelente de los aceites esenciales de *Rosmarinus officinalis*, *Mentha piperita*, *Melaleuca alternifolia* y *Commiphora myrrha* y sus mezclas en hembras adultas de *Tetranychus urticae* en laboratorio y cultivo protegido. El aceite esencial de *C. myrrha* exhibió una Cl_{50} de 0.55 ml L^{-1} y las mezclas *R. officinalis* + *M. piperita* + *M. alternifolia* + *C. myrrha*, *C. myrrha* + *M. piperita*, y *C. myrrha* + *M. alternifolia* presentaron 68%, 60%, y 36% de mortalidad, respectivamente. Los aceites esenciales de *C. myrrha* y *M. alternifolia* mostraron 95 y 70% de repelencia, respectivamente. Las mezclas de *C. myrrha* + *M. alternifolia*, *C. myrrha* + *M. piperita* y *C. myrrha* + *R. officinalis* proporcionaron repelencia de 85, 74 y 73%, respectivamente. La toxicidad por contacto residual del aceite esencial de *C. myrrha* en cultivo protegido presentó una mortalidad del 93%, mientras que el acaricida fenpiroximato presentó un 80%. Los componentes de los aceites esenciales fueron: eucaliptol (49.66%), *M. piperita* mentol (48.53%), *M. alternifolia* terpinen-4-ol (48.93%), y *C. myrrha* bencil benzoato (97.71%). El aceite esencial de *C. myrrha* y las mezclas *R. officinalis* + *M. piperita* + *M. alternifolia* + *C. myrrha* y *C. myrrha* + *M. piperita* mostraron una mortalidad significativa. Sin embargo, se necesitan más estudios para evaluar la relación costo/beneficio y los efectos en organismos no objetivo.

Palabras clave: ácaro araña, mezclas, toxicidad.

Introduction

The two-spotted spider mite, *Tetranychus urticae* Koch (Acari: Tetranychidae), is a polyphagous cosmopolitan pest responsible for serious damage to crops of economic interest in irrigated and protected cultivation systems worldwide (Migeon *et al.*, 2019). The considerable negative effect of this pest on agriculture is related to its short biological cycle, high fertility rate and ability to cause injuries (Araújo *et al.*, 2020). The main form of control is

through the application of synthetic acaricides, widely used throughout Brazil (Araújo *et al.*, 2020). The indiscriminate use of these synthetic products has favored the selection of resistant populations (Monteiro *et al.*, 2015). *T. urticae* is currently resistant to 95 active ingredients with different modes of action, with records of more than 500 cases of resistant populations in various regions of the world and Brazil (Mota-Sanchez & Wise, 2021). Additionally, these acaricides can cause other undesirable effects, such as biological imbalance due to the elimination of beneficial organisms (Efrom *et al.*, 2012). Thus, there is a need to

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¹ Universidade Federal do Espírito Santo, Departamento de Entomologia, Campus Alegre (Brazil).

² Instituto Federal de Educação, Ciência e Tecnologia do Espírito Santo, Campus Alegre (Brazil).

* Corresponding author: julielsonoliveira@hotmail.com



look for control alternatives that can minimize such problems. To this end, researchers have been investigating new acaricides based on essential oils extracted from plants. The literature reports the effects of essential oils and their constituents on several arthropods, including *T. urticae* (Mar *et al.*, 2018). However, few studies have addressed the acaricidal properties of essential oils and their mixtures for the management of *T. urticae* in protected cultivation (Born *et al.*, 2018).

This study evaluated the residual contact toxicity and the repelling effect of the essential oils of *Rosmarinus officinalis* (Lamiaceae), *Mentha piperita* (Lamiaceae), *Melaleuca alternifolia* (Myrtaceae) and *Commiphora myrrha* (Burseraceae) and their mixtures on adult females of *T. urticae* under laboratory conditions and protected cultivation.

Materials and methods

Biological material

The adult females of *T. urticae* were bred on jack bean (*Canavalia ensiformis* L. (Fabaceae) plants in air-conditioned rooms with $25 \pm 1^\circ\text{C}$, $65 \pm 3\%$ relative humidity and 12 h photoperiod in the Núcleo de Desenvolvimento Científico e Tecnológico em Manejo de Pragas e Doenças (NUDE-MAFI) at the Centro de Ciências Agrárias e Engenharia at Universidade Federal do Espírito Santo - Campus de Alegre, Espírito Santo, Brazil (CCAUE-UFES).

Obtaining the essential oils

Essential oils with 100% purity were purchased from the company Casa do Saboeiro[®] Ltda, located at Rio Grande do Sul, São Paulo, Brazil.

Chemical characterization of essential oils

The essential oils were analyzed by gas chromatography with flame ionization detector (GC/FID) (GC-2010 Plus, Shimadzu, Tokyo, Japan) and gas chromatography coupled to mass spectrometry (GC/MS) (QP2010 Plus, Shimadzu, Tokyo, Japan) adapting the methodology of Souza *et al.* (2017). The following chromatographic conditions were used in both analyses: capillary column of fused silica (30 m x 0.25 mm) with stationary phase Rtx[®]-5MS (0.25 μm of film thickness); N_2 (in GC/FID analysis) and He (in GC/MS analysis) as carrier gas with a flow rate of 3.0 ml/min; the oven temperature followed a schedule in which it remained at an initial temperature of 40°C for 3 min and then gradually increased by $3^\circ\text{C}/\text{min}$ until it reached 240°C , maintaining this temperature for 5 min, with an injector temperature of 250°C , detector temperature of 280°C and split ratio of 1:30. GC/MS analysis was performed on

equipment GCMS-QP2010 Plus with detector and AOC-5000 sample injection system (Shimadzu, Tokyo, Japan) operated by electronic impact with impact energy of 70 eV, scan speed of 1,000, scanning interval of 0.50 fragments sec^{-1} , and detected fragments from 29 to 400 m/z.

The identification of the chemical components of the oils was carried out by comparing their mass spectra with those available in the Wiley 7, NIST 05, NIST 05s peak spectrum database with the co-injection of standards and by the retention indexes (RI). To calculate the RI, a mixture of linear n-alkanes (C7 to C40) was used as standard. The calculated RI for each compound was compared with values reported in the literature (Adams, 2007).

Preparation of mixtures of essential oils

The blends of the essential oils of *R. officinalis*, *M. piperita*, *M. alternifolia* and *C. myrrha* were made in the following proportion (Tab. 1), according to the methodology proposed by Pavela (2015).

TABLE 1. Proportions of mixtures of essential oils (v/v).

| Mixtures | Proportions of mixtures | Essential oils |
|----------|-------------------------|------------------------------------------------------------------------------------|
| 1 | 1:1:1:1 | <i>R.officinalis</i> + <i>M.piperita</i> + <i>M.alternifolia</i> + <i>C.myrrha</i> |
| 2 | 1:1 | <i>C.myrrha</i> + <i>M.piperita</i> |
| 3 | 1:1 | <i>C.myrrha</i> + <i>M.alternifolia</i> |
| 4 | 1:1 | <i>C.myrrha</i> + <i>R.officinalis</i> |
| 5 | 1:1 | <i>M.alternifolia</i> + <i>R.officinalis</i> |
| 6 | 1:1 | <i>M.piperita</i> + <i>R.officinalis</i> |
| 7 | 1:1 | <i>M.piperita</i> + <i>M.alternifolia</i> |

Residual contact toxicity test of essential oils and mixtures

The residual contact toxicity of essential oils was evaluated on adult females of *T. urticae* using the methodology adapted from Paes *et al.* (2015). Jack bean leaf discs with a 4.5 cm diameter were placed on a 6.5 cm diameter acrylic box with a 5 mm deep layer of a 1% ml L^{-1} agar-water solution; Tween 80 was diluted to a concentration of 0.05% (ml L^{-1}). To spray the treatments (control (distilled water + Tween 80), essential oil + distilled water + Tween 80, and mixtures of essential oils at a 1:1 proportion + distilled water + Tween 80 at a concentration of 2% (ml L^{-1})) an airbrush calibrated to a pressure of 15 N m^{-2} was used, at a distance of 30 cm. Each leaf disc was infested with 20 adult female mites aged 5 d, placed on the underside of the leaf. For each essential oil, five replicates were performed, totaling 100 mites per treatment. The concentration of treatments was in accordance with Ataide *et al.* (2020).

The test was conducted in an air-conditioned chamber (temperature of $25\pm 1^{\circ}\text{C}$, relative humidity of $70\pm 10\%$ and photoperiod of 12 h). After 24 h, adult female mortality was assessed. To confirm mortality, the mites were lightly touched with a fine bristle brush (number 00) on the dorsal area. Immobile mites were considered dead.

Lethal concentration estimate

The lethal concentration (LC) was estimated with essential oils that reached a mortality rate above 80%, calculated by PROBIT analysis (Finney, 1971). Therefore, the lethal concentration of myrrh essential oil was estimated on adult females of *T. urticae*. For this, seven concentrations were used: 0.0, 0.3, 0.5, 0.8, 1.2, 1.5, and 2.0% (ml L^{-1}) of myrrh essential oil, with the lower limit (concentration that causes the death of about 10% of *T. urticae*) and higher (concentration that causes the death of about 90% of *T. urticae*) determined by preliminary tests. In the control, distilled water + Tween 80 was used; for the treatments, essential oil + distilled water + Tween 80 was used. This step of the test was performed according to the procedures adopted in the toxicity test. Mortality was assessed after 24 h.

Repellency test of essential oils and mixtures

The double choice chance method adapted from Aslan *et al.* (2004) was carried out using Petri dishes of 10 cm in diameter containing culture medium at 1% ml L^{-1} (water + agar). Two leaf discs of *C. ensiformis* (2.5 cm in diameter, treated and untreated) were placed in each Petri dish, joined by a glass coverslip (18 mm). The experiment was carried out in triplicate, with six replicates with 20 *T. urticae*, totaling 120 mites per treatment. Each treated leaf disc was immersed according to the proposed treatments: distilled water + essential oil + Tween 80 and the mixtures of essential oils at a 1:1 proportion + distilled water + Tween 80 at a concentration of 2% (ml L^{-1}). The untreated discs were immersed in distilled water + Tween 80. After immersion, the leaf discs were placed to dry for 20 min at room temperature. Subsequently, 20 *T. urticae* adult females of 5 d of age were placed on the underside of the leaf in the center of the coverslip with the aid of a fine-bristle brush (number 00). The treatments were placed in an air-conditioned room (temperature of $25\pm 1^{\circ}\text{C}$, relative humidity of $70\pm 10\%$ and photoperiod of 12 h). After 72 h, the number of mites present in the treated and untreated discs was counted. The mites present on the untreated discs were considered to be repelled by the essential oil.

Toxicity test by residual contact in protected cultivation

The experiment was carried out in a protected cultivation at the Centro de Ciências Agrárias e Engenharia at

Universidade Federal do Espírito Santo (CCAUE-UFES) located in Alegre, ES, Brazil. The upper part of the greenhouse of the protected cultivation was covered with 150 μm plastic film and the side was covered with Optinet 50 mesh anti-aphid screen.

Jack beans (*Canavalia ensiformis* var. Coriacea (Biblioth. Robot) (Fabaceae) were sown manually and deposited in disposable cups (100 ml) containing Provaso[®] (Agrosolo, Conceição da Barra, ES, Brazil) organic substrate. Irrigation was performed daily, with each plant receiving the same amount of water for irrigation. When the *C. ensiformis* plants reached the age of 14 d, uniform plants were selected. After 10 d, the inoculation of 5,000 adult females of *T. urticae* was carried out. Ten d after inoculation, spraying was performed with an airbrush calibrated at a pressure of 15 N m^{-2} at a distance of approximately 30 cm from *C. ensiformis* plants, with a volume of 5 ml of the syrup of the aforementioned treatments.

Approximately 1 ml was sprayed on the upper and lower sides of each leaf of *C. ensiformis*; the treatments were as follows: distilled water + Tween 80 (control), *C. myrrha* essential oil + distilled water + Tween 80 (treatment) at LC_{95} and the acaricide fenpyroximate from the chemical group pyrazole + distilled water (positive treatment) at the commercial concentration.

The experimental design was in randomized blocks, with three treatments, five replicates and five blocks. Each block consisted of 15 plants, with five plants from each treatment arranged at random. After 24 h of exposure to treatments, the total number of live and dead mites was counted. To confirm mortality, the mites were lightly touched with a fine bristle brush (number 00); immobile mites were considered dead. The temperature of the protected cultivation varied between 25°C and 38°C , and the humidity between 40% and 85%.

Data analysis

For the acute toxicity test, an ANOVA was applied and then the means were compared by the Scott-Knott test ($P\leq 0.05$). For the pair test, the Pearson's chi-square test was applied. The concentration-response curves as well as the lethal concentration (LC_{50}) of the essential oil of myrrh were subjected to PROBIT analysis (Finney, 1971). In the test in protected cultivation, a randomized block design was used, with means compared by the Tukey's test ($P\leq 0.05$). All analyzes were performed using R software (R Development Core Team, 2010).

Results

Chemical characterization of essential oils

In the gas chromatography analysis with flame ionization detector (GC/FID), eight constituents were identified in the essential oil of *R. officinalis*, in which eucalyptol (49.66%), α -Pinene (17.08%) and camphor (14.79%) were the main ones. Fourteen constituents were identified in the essential oil of *M. alternifolia*, with terpinen-4-ol (48.93%) and γ -Terpinene (20.88%) being the main ones. Nine constituents were found in the essential oil of *M. piperita*, in which menthol (48.53%) and menthone (24.25%) were the main ones. Two constituents were identified in the essential

oil of *C. myrrha*, with benzyl benzoate (97.71%) being the main one (Tab. 2).

Residual contact toxicity test of essential oils and mixtures

The residual contact toxicity of essential oils and their mixtures against adult females of *T. urticae* is shown in Figure 1. The essential oil *C. myrrha* exhibited 95% mortality ($F_{11,48} = 119.12$; $P < 0.001$), and the mixtures *R. officinalis* + *M. piperita* + *M. alternifolia* + *C. myrrha*, *C. myrrha* + *M. piperita*, and *C. myrrha* + *M. alternifolia* showed 68%, 60% and 36% ($F_{11,48} = 119.12$; $P < 0.001$).

TABLE 2. Chromatographic analysis of the essential oils of *R. officinalis*, *M. alternifolia*, *M. piperita*, and *C. myrrha*.

| Name of compound | CRI | TRI | Relative area (%) | | | |
|-----------------------|------|------|-----------------------|------------------------|--------------------|------------------|
| | | | <i>R. officinalis</i> | <i>M. alternifolia</i> | <i>M. piperita</i> | <i>C. myrrha</i> |
| α -Pinene | 930 | 932 | 17.08 | 2.41 | | |
| Camphene | 943 | 946 | 4.63 | | | |
| β -Pinene | 972 | 974 | 4.76 | 0.63 | | |
| β -Myrcene | 991 | 988 | | 0.54 | | |
| α -Terpinene | 1014 | 1014 | | 8.24 | | |
| α -Cymene | 1022 | 1022 | | 4.11 | | |
| β -Phellandrene | 1026 | 1025 | | 1.58 | | |
| Eucalyptol | 1028 | 1026 | 49.66 | 2.23 | | |
| γ -Terpinene | 1057 | 1054 | | 20.88 | | |
| Terpinolene | 1086 | 1086 | | 3.01 | | |
| Linalool | 1100 | 1095 | 2.35 | | | |
| Camphor | 1141 | 1141 | 14.79 | | | |
| Isopulegol | 1143 | 1145 | | | 1.73 | |
| Menthone | 1153 | 1148 | | | 24.25 | |
| Isoborneol | 1154 | 1155 | 0.72 | | | |
| Neomenthol | 1163 | 1161 | | | 16.61 | |
| Menthol | 1172 | 1167 | | | 48.53 | |
| Isomenthol | 1182 | 1179 | | | 1.07 | |
| Terpinen-4-ol | 1178 | 1178 | | 48.93 | | |
| α -Terpineol | 1189 | 1186 | 3.94 | 3.51 | 0.78 | |
| Piperitone | 1252 | 1249 | | | 1.46 | |
| Menthyl acetate | 1293 | 1294 | | | 4.78 | |
| trans-Caryophyllene | 1414 | 1417 | | | 0.79 | |
| Aromadendrene | 1434 | 1439 | | 1.24 | | |
| Viridiflorene | 1491 | 1496 | | 1.42 | | |
| δ -Cadinene | 1520 | 1522 | | 1.27 | | |
| Cubenol | 1655 | 1645 | | | | 2.29 |
| Benzyl benzoate | 1759 | 1759 | | | | 97.71 |

The compounds were identified by LTPRI index (GC/FID) and mass spectrometry (GC/MS) using an Rtx®-5MS column. CRI - retention index calculated from data obtained by sampling of saturated n-alkanes (C7-C40). TRI - tabulated retention index (Adams, 2007). Compounds with relative areas >0.5% were identified.

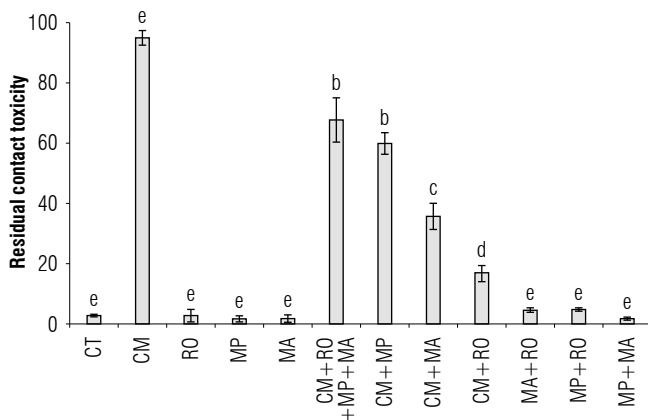


FIGURE 1. Means (\pm standard error) of mortality of *T. urticae* females at a temperature of $25\pm 2^\circ\text{C}$, relative humidity of $70\pm 10\%$, and a photoperiod of 12 h by essential oil mixtures; CT - control, CM - *C. myrrha*, RO - *R. officinalis*, MP - *M. piperita*, MA - *M. alternifolia*. Equal letters do not indicate statistically significant differences from each other according to the Scott-Knott test at 5% probability.

Lethal concentration

The lethal concentration of the essential oil of *C. myrrha* showed an LC_{95} of 1.6 ml L^{-1} and an LC_{50} of 0.55 ml L^{-1} (Tab. 3).

Repellency test of essential oils and mixtures

The repellency effect of essential oils and their mixtures in adult females of *T. urticae* is shown in Figure 2. Among the essential oils and their mixtures, the essential oils of *C. myrrha* and *M. alternifolia* showed 95 and 70% of repellency, respectively ($F_{10, 89} = 4.020$; $P < 0.001$). Mixtures of *C. myrrha* + *M. alternifolia*, *C. myrrha* + *M. piperita*, and *C. myrrha* + *R. officinalis* provided repellency of 85, 74 and 73%, respectively ($F_{10, 89} = 4.020$; $P < 0.001$).

Toxicity test by residual contact in protected cultivation

The essential oil of *C. myrrha* showed 93% toxicity against adult females of *T. urticae* in protected cultivation, whereas

TABLE 3. Lethal concentration of *C. myrrha* oil on adult females of *T. urticae*.

| Essential oil | N | DF | Slope \pm SE | χ^2 | P | Lethal concentration (ml L^{-1}) (LC 95%) | |
|------------------|-----|----|----------------|----------|------|------------------------------------------------------|----------------------|
| | | | | | | LC_{50} | LC_{95} |
| <i>C. myrrha</i> | 300 | 3 | 3.54 ± 0.36 | 2.02 | 0.56 | $0.55 (0.48\pm 0.61)$ | $1.6 (1.34\pm 2.07)$ |

N - Number of individuals; DF - degrees of freedom; Slope - slope of the line; X - Chi-square; P - P-value; SE - confidence interval.

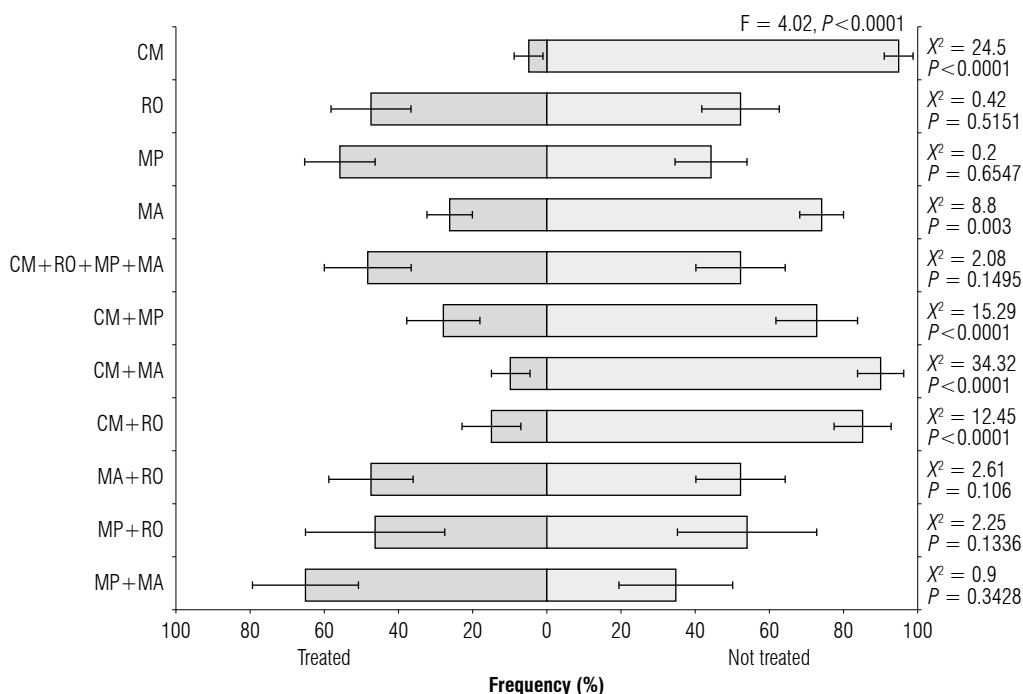


FIGURE 2. Frequency (%) (\pm standard error) of *T. urticae* adult females repelled at a temperature of $25\pm 2^\circ\text{C}$, a relative humidity of $70\pm 10\%$ and a photoperiod of 12 h on the application of essential oils and their mixtures; CM - *C. myrrha*, RO - *R. officinalis*, MP - *M. piperita*, MA - *M. alternifolia*.

the fenpyroximate showed 80% toxicity ($F_{2,8} = 545.24$; $P < 0.001$) (Fig. 3).

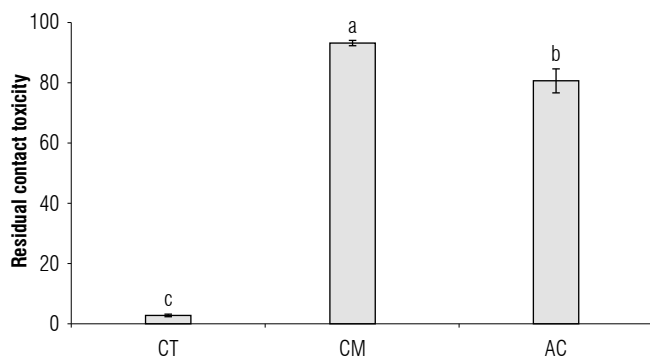


FIGURE 3. Means (\pm standard error) of mortality of females *T. urticae* in protected cultivation by the application of essential oil and acaricide; CT - control, CM - *C. myrrha* and AC - acaricide (fenpyroximate); equal letters indicate treatments that do not differ statistically from each other according to the Tukey's test at 5% probability.

Discussion

The major constituent identified in the essential oil of *C. myrrha* was benzyl benzoate (97.71%) which differed from the major constituents identified by Mohamed *et al.* (2014). The major constituents in the essential oil of *M. piperita* were menthol (48.53%) and menthone (24.25%), which were the same major constituents found by Kizil *et al.* (2010) and Moghaddam *et al.* (2013). The major constituents found in *M. alternifolia* were terpinen-4-ol (48.93%) and γ -Terpinene (20.88%), with the same major constituents found by Zhang *et al.* (2018) and Silva *et al.* (2019). In the essential oil of *R. officinalis*, the major constituents found were eucalyptol (49.66%), α -Pinene (17.08%) and camphor (14.79%); similar results were found by Borges *et al.* (2018) and Jardak *et al.* (2017) for constituents α -Pinene and camphor.

This study found that the essential oil of *C. myrrha* showed residual contact toxicity against adult females of *T. urticae*, exhibiting an LC_{50} of 0.55 ml L^{-1} . Araújo *et al.* (2012) found that the essential oil of *Piper aduncum*, Piperaceae showed an LC_{50} of $7.17 \mu\text{l ml}^{-1}$ on adults of *T. urticae*. Ribeiro *et al.* (2019) evaluated four different *Citrus* sp. essential oils on *T. urticae*. Of these, the essential oil of *Citrus limon*, Rutaceae showed an LC_{50} of $25.18 \mu\text{l ml}^{-1}$ L. Essential oils that show efficiency in the control of pest arthropods by residual contact can penetrate the tegument layers (Enan, 2001). Consequently, since essential oils are made up of several substances, they can act in more than one place exhibiting a neurotoxic action (Isman, 2006).

The mixtures of *R. officinalis* + *M. piperita* + *M. alternifolia* + *C. myrrha* and *C. myrrha* + *M. piperita* at a 1:1 ratio

showed promising results in terms of the toxic effect from residual contact on adult females of *T. urticae* with intermediate toxicity, which resulted in an additive interaction. Although we did not find synergistic effects, this type of interaction has been described by Mwaiko (1992) for mixtures of bark essential oils from two species of *Citrus* (*C. limon*, *C. aurantium*, Rutaceae) against *Culex pipiens* L. (Diptera: Culicidae) larvae. Benelli *et al.* (2017) evaluated the acute toxicity of binary mixtures of essential oils of the Apiaceae family on the larvae of *C. quinquefasciatus*, an important vector of filariasis, and found that *Trachyspermum ammi* Apiaceae + *Pimpinella anisum*, Apiaceae (1:2 ratio) and *S. olusatrum* + *P. anisum* (1:1 ratio) were the most toxic to the pest.

The repellent effect of essential oils in integrated pest management is important; as these substances keep pests away from the crop, damage is minimized (Da Camara *et al.*, 2015). The results of the repellency tests suggest that the essential oils of *C. myrrha* and *M. alternifolia* and the mixtures of *C. myrrha* + *M. piperita*, *C. myrrha* + *M. alternifolia* and *C. myrrha* + *R. officinalis* showed a repellent effect on females of *T. urticae*. In previous studies, Araújo Júnior *et al.* (2010) found that the essential oils of *C. aurantium* and *C. sinensis* var. Mimo showed repellency of *T. urticae* at a concentration below 2.5%, while the essential oil of *C. sinensis* var. Pear had a neutral effect. Sararit and Auamcharoen (2020) observed that the essential oils of *Anethum graveolens* and *Allium sativum* exhibited repellency of adult females of *T. urticae* at concentrations of 15 to 20%. Farahani *et al.* (2020) noted that essential oils of *Thymus daenensis* (Lamiaceae), *Satureja khuzestanica* (Lamiaceae), and *Satureja bakhtiarica* (Lamiaceae) showed repellency against adult *T. urticae*. The repelling action of essential oils triggers an escape behavior of mites, detected by the olfactory sensilla present in the legs (Missbach *et al.*, 2014; Oliveira *et al.*, 2018).

The tests in protected cultivation confirmed the laboratory observations that the essential oil of *C. myrrha* was toxic to adult females of *T. urticae*. In a study carried out to avoid the spread of mites in a greenhouse, Da Camara *et al.* (2015) observed that the essential oils of *C. sinensis* and *C. aurantium* were repellent to *T. urticae*. The essential oil of *C. aurantium*, in particular, prevented the dispersion of the pest for a period of 1 week. In a greenhouse trial, Potenza *et al.* (2006) observed toxicity by contact of the aqueous extract of *Allamanda cathartica* (Apocynaceae), *Dieffenbachia brasiliensis* (Araceae), *Cenchrus purpureus* (Schumach.) Morrone (Poaceae), *Annona squamosa* (Annonaceae), *Ruta graveolens* (Rutaceae), *Sonchus oleraceus*

(Asteraceae), *Spondias purpurea* (Anacardiaceae), *Lytechinus variegatus* (Euphorbiaceae), *Impatiens walleriana* (Balsaminaceae), *Stryphnodendron adstringens* (Fabaceae), *Solanum melongena* (Solanaceae), *Campsiandra angustifolia* var. *angustifolia* (Fabaceae), and *Allium* (Amaryllidaceae) on *T. urticae*. However, only plants with *D. brasiliensis*, *R. graveolens*, *A. squamosa*, *S. oleraceus*, *I. walleriana*, *A. angustifolia*, *S. adstringens*, and *S. melongena* promoted a significant reduction in the population of *T. urticae* of between 60 and 86%. In another study, EcoTrol (containing 10% rosemary oil), a pesticide based on the essential oil of rosemary, proved to be efficient in the control of *T. urticae* in a greenhouse (Miresmailli & Isman, 2006).

Conclusions

The essential oil of *C. myrrha* and the binary and quaternary mixtures *C. myrrha* + *M. piperita* and *R. officinalis* + *M. piperita* + *M. alternifolia* + *C. myrrha*, showed significant mortality in this study. The essential oil of *C. myrrha*, in turn, showed results in a protected cultivation similar to those obtained with the application of commercial acaricides for the control of *T. urticae* in orchards and in a protected system in Brazil. This study shows the feasibility of using a botanical acaricide with *C. myrrha* essential oil as an active ingredient for the management of *T. urticae*. However, for large-scale use, studies to reduce essential oil production costs and the effects on non-target organisms are needed.

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Conflict of interest statement

The authors declare that there is no conflict of interests regarding the publication of this article.

Author's contributions

HBZ and LM carried out the experiments. JOA, FDD, FGH, and AH carried out the laboratory and protected cultivation experiment and collected the data. JOA and FDD carried out the data analysis and writing of the manuscript. All the authors reviewed the manuscript.

Literature cited

- Adams, R. P. (2007). *Identification of essential oil components by gas chromatography/mass spectrometry* (4th ed.). Allured Publishing Corporation.
- Araújo Júnior, C. P., Camara, C. A. G., Neves, I. A., Ribeiro, N. C., Gomes, C. A., Moraes, M. M., & Botelho, P. S. (2010). Acaricidal

activity against *Tetranychus urticae* and chemical composition of peel essential oils of three *Citrus* species cultivated in NE Brazil. *Natural Product Communications*, 5(3), 471–476. <https://doi.org/10.1177/1934578X1000500326>

- Araújo, M. J. C., Câmara, C. A. G., Born, F. S., & Moraes, M. M. (2020). Acaricidal activity of binary blends of essential oils and selected constituents against *Tetranychus urticae* in laboratory/greenhouse experiments and the impact on *Neoseiulus californicus*. *Experimental and Applied Acarology*, 80, 423–444. <https://doi.org/10.1007/s10493-020-00464-8>
- Araújo, M. J. C., Câmara, C. A. G., Born, F. S., Moraes, M. M., & Badji, C. A. (2012). Acaricidal activity and repellency of essential oil from *Piper aduncum* and its components against *Tetranychus urticae*. *Experimental and Applied Acarology*, 57, 139–155. <https://doi.org/10.1007/s10493-012-9545-x>
- Aslan, I., Özbek, H., Çalmaşur, Ö., & Şahin, F. (2004). Toxicity of essential oil vapours to two greenhouse pests, *Tetranychus urticae* Koch and *Bemisia tabaci* Genn. *Industrial Crops and Products*, 19(2), 167–173. <https://doi.org/10.1016/j.indcrop.2003.09.003>
- Ataide, J. O., Pratisoli, D., Fragoso, D. F. M., & Pinheiro, P. F. (2020). Caracterização e atividade inseticida do óleo essencial de *Zingiber officinale* Roscoe - Zingiberaceae sobre *Duponchelia fovealis* Zeller, 1847 (Lepidoptera: Crambidae). *Revista em Agronegócio e Meio Ambiente*, 13(2), 693–705. <https://doi.org/10.17765/2176-9168.2020v13n2p693-705>
- Benelli, G., Pavela, R., Iannarelli, R., Petrelli, R., Cappellacci, L., Cianfaglione, K., Afshar, F. H., Nicoletti, M., Canale, A., & Maggi, F. (2017). Synergized mixtures of Apiaceae essential oils and related plant-borne compounds: larvicidal effectiveness on the filariasis vector *Culex quinquefasciatus* Say. *Industrial Crops and Products*, 96, 186–195. <https://doi.org/10.1016/j.indcrop.2016.11.059>
- Borges, R. S., Keita, H., Ortiz, B. L. S., Sampaio, T. I. S., Ferreira, I. M., Lima, E. S., Silva, M. J. A., Fernandes, C. P., Oliveira, A. E. M. F. M., Conceição, E. C., Rodrigues, A. B. L., Pereira Filho, A. C. M., Castro, A. N., & Carvalho, J. C. T. (2018). Antiinflammatory activity of nanoemulsions of essential oil from *Rosmarinus officinalis* L.: in vitro and in zebrafish studies. *Inflammopharmacology*, 26, 1057–1080. <https://doi.org/10.1007/s10787-017-0438-9>
- Born, F. S., Camara, C. A. G., Melo, J. P. R., & Moraes, M. M. (2018). Acaricidal property of the essential oil from *Lippia gracilis* against *Tetranychus urticae* and a natural enemy, *Neoseiulus californicus*, under greenhouse conditions. *Experimental and Applied Acarology*, 75, 491–502. <https://doi.org/10.1007/s10493-018-0286-3>
- Da Camara, C. A. G., Akhtar, Y., Isman, M. B., Seffrin, R. C., & Born, F. S. (2015). Repellent activity of essential oils from two species of *Citrus* against *Tetranychus urticae* in the laboratory and greenhouse. *Crop Protection*, 74, 110–115. <https://doi.org/10.1016/j.cropro.2015.04.014>
- Efrom, C. F. S., Redaelli, L. R., Meirelles, R. N., & Ourique, C. B. (2012). Side-effects of pesticides used in the organic system of production on *Apis mellifera* Linnaeus, 1758. *Brazilian Archives of Biology and Technology*, 55(1), 47–53. <https://doi.org/10.1590/S1516-89132012000100005>
- Enan, E. (2001). Insecticidal activity of essential oils: octopaminergic sites of action. *Comparative Biochemistry and Physiology*, 130(3), 325–337. [https://doi.org/10.1016/S1532-0456\(01\)00255-1](https://doi.org/10.1016/S1532-0456(01)00255-1)

- Farahani, S., Bandani, A. R., & Amiri, A. (2020). Toxicity and repellency effects of three essential oils on two populations of *Tetranychus urticae* (Acari: Tetranychidae). *Persian Journal of Acarology*, 9(1), 67–81.
- Finney, D. J. (1971). *Probit analysis*. Cambridge University Press.
- Isman, M. B. (2006). Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Annual Review of Entomology*, 51, 45–66. <https://doi.org/10.1146/annurev.ento.51.110104.151146>
- Jardak, M., Elloumi-Msaddi, J., Aifa, S., & Mnif, S. (2017). Chemical composition, anti-biofilm activity and potential cytotoxic effect on cancer cells of *Rosmarinus officinalis* L. essential oil from Tunisia. *Lipids in Health and Disease*, 16, Article 190. <https://doi.org/10.1186/s12944-017-0580-9>
- Kizil, S., Haşimi, N., Tolan, V., Kilinç, E., & Yüksel, U. (2010). Mineral content, essential oil components and biological activity of two mentha species (*M. piperita* L., *M. spicata* L.). *Turkish Journal of Field Crops*, 15(2), 148–153.
- Mar, J. M., Silva, L. S., Azevedo, S. G., França, L. P., Goes, A. F. F., Santos, A. L., Bezerra, J. A., Nunomura, R. C. S., Machado, M. B., & Sanches, E. A. (2018). *Lippia origanoides* essential oil: an efficient alternative to control *Aedes aegypti*, *Tetranychus urticae* and *Cerataphis lataniae*. *Industrial Crops and Products*, 111, 292–297. <https://doi.org/10.1016/j.indcrop.2017.10.033>
- Migeon, A., Tixier, M. S., Navajas, M., Litskas, V. D., & Stavrinides, M. C. (2019). A predator-prey system: *Phytoseiulus persimilis* (Acari: Phytoseiidae) and *Tetranychus urticae* (Acari: Tetranychidae): worldwide occurrence datasets. *Acarologia*, 59(3), 301–307. <https://doi.org/10.24349/acarologia/20194322>
- Miresmailli, S., & Isman, M. B. (2006). Efficacy and persistence of rosemary oil as an acaricide against twospotted spider mite (Acari: Tetranychidae) on greenhouse tomato. *Journal of Economic Entomology*, 99(6), 2015–2023. <https://doi.org/10.1093/jee/99.6.2015>
- Missbach, C., Dweck, H. K. M., Vogel, H., Vilcinskis, A., Stensmyr, M. C., Hansson, B. S., & Grosse-Wilde, E. (2014). Evolution of insect olfactory receptors. *eLife*, 3, Article e02115. <https://doi.org/10.7554/eLife.02115>
- Moghaddam, M., Pourbaige, M., Tabar, H. K., Farhadi, N., & Hosseini, S. M. A. (2013). Composition and antifungal activity of peppermint (*Mentha piperita*) essential oil from Iran. *Journal of Essential Oil Bearing Plants*, 16(4), 506–512. <https://doi.org/10.1080/0972060X.2013.813265>
- Mohamed, A. A., Ali, S. I., EL-Baz, F. K., Hegazy, A. K., & Kord, M. A. (2014). Chemical composition of essential oil and *in vitro* antioxidant and antimicrobial activities of crude extracts of *Commiphora myrrha* resin. *Industrial Crops and Products*, 57, 10–16. <https://doi.org/10.1016/j.indcrop.2014.03.017>
- Monteiro, V. B., Gondim Jr., M. G. C., Oliveira, J. E. M., Siqueira, H. A. A., & Sousa, J. M. (2015). Monitoring *Tetranychus urticae* Koch (Acari: Tetranychidae) resistance to abamectin in vineyards in the Lower Middle São Francisco Valley. *Crop Protection*, 69, 90–96. <https://doi.org/10.1016/j.cropro.2014.12.012>
- Mota-Sanchez, D., & Wise, J. C. (2021). *The arthropod pesticide resistance database*. Michigan State University. <http://www.pesticideresistance.org>
- Mwaiko, G. L. (1992). Citrus peel oil extracts as mosquito larvae insecticides. *East African Medical Journal*, 69(4), 223–226.
- Oliveira, A. P., Santos, A. A., Santana, A. S., Lima, A. P. S., Melo, C. R., Santana, E. D. R., Sampaio, T. S., Blank, A. F., Araújo, A. P. A., Cristaldo, P. F., & Bacci, L. (2018). Essential oil of *Lippia sidoides* and its major compound thymol: toxicity and walking response of populations of *Sitophilus zeamais* (Coleoptera: Curculionidae). *Crop Protection*, 112, 33–38. <https://doi.org/10.1016/j.cropro.2018.05.011>
- Paes, J. P. P., Rondelli, V. M., Costa, A. V., Vianna, U. R., & Queiroz, V. T. (2015). Caracterização química e efeito do óleo essencial de Erva-de-Santa-Maria sobre o ácaro-rajado de morango. *Revista Brasileira de Fruticultura*, 37(2), 346–354. <https://doi.org/10.1590/0100-2945-057/14>
- Pavela, R. (2015). Acute toxicity and synergistic and antagonistic effects of the aromatic compounds of some essential oils against *Culex quinquefasciatus* Say larvae. *Parasitology Research*, 114, 3835–3853. <https://doi.org/10.1007/s00436-015-4614-9>
- Potenza, M. R., Gomes, R. C. O., Jocy, T., Takematsu, A. P., & Ramos, A. C. O. (2006). Avaliação de produtos naturais para o controle do ácaro rajado *Tetranychus Urticae* (Koch, 1836) (Acari: Tetranychidae) em casa de vegetação. *Arquivos do Instituto Biológico*, 73(4), 455–459.
- R Development Core Team. (2010). *R: a language and environment for statistical computing, reference index version 2.12.1*. R Foundation for Statistical Computing.
- Ribeiro, N. C., Camara, C. A. G., Melo, J. P. R., & Moraes, M. M. (2019). Acaricidal properties of essential oils from agro-industrial waste products from citric fruit against *Tetranychus urticae*. *Journal of Applied Entomology*, 143, 731–743. <https://doi.org/10.1111/jen.12642>
- Sararit, P., & Auamcharoen, W. (2020). Biological activities of essential oils from *Anethum graveolens* L. and *Allium sativum* L. for controlling *Tetranychus truncatus* Ehara and *Tetranychus urticae* Koch. *Journal of Biopesticides*, 13(1), 1–12.
- Silva, C. S., Figueiredo, H. M., Stamford, T. L. M., & Silva, L. H. M. (2019). Inhibition of *Listeria monocytogenes* by *Melaleuca alternifolia* (tea tree) essential oil in ground beef. *International Journal of Food Microbiology*, 293, 79–86. <https://doi.org/10.1016/j.ijfoodmicro.2019.01.004>
- Souza, T. S., Ferreira, M. F. S., Menini, L., Souza, J. R. C. L., Parreira, L. A., Cecon, P. R., & Ferreira, A. (2017). Essential oil of *Psidium guajava*: influence of genotypes and environment. *Scientia Horticulturae*, 216, 38–44. <https://doi.org/10.1016/j.scienta.2016.12.026>
- Zhang, X., Guo, Y., Guo, L., Jiang, H., & Ji, Q. (2018). *In vitro* evaluation of antioxidant and antimicrobial activities of *Melaleuca alternifolia* essential oil. *BioMed Research International*, 2018, Article 2396109. <https://doi.org/10.1155/2018/2396109>