*Technological research article / https://doi.org/10.15446/rcciquifa.v52n2.110730*

# Development and characterization of doxycycline gelatin nanoparticles

Perla García-Guzmán<sup>1</sup>', Luis Ocampo<sup>1</sup>, Lilia Gutierrez<sup>1</sup>, Hector Sumano<sup>1</sup>, María Josefa Bernad<sup>2</sup>

1 Facultad de Medicina Veterinaria y Zootecnia, Universidad Nacional Autónoma de México (UNAM), Ciudad de México, México.

2 Facultad de Química, Universidad Nacional Autónoma de México (UNAM), Ciudad de México, México.

Corresponding author E-mail: pgarcí[a@correo.xoc.uam.mx](mailto:lortegaa@correo.xoc.uam.mx)

Received: December 6, 2022 Corrected: March 3, 2023 Accepted: March 8, 2023

#### **SUMMARY**

Introduction: Doxycycline (DOXI) is a wide-spectrum antimicrobial drug used for urinary, intestinal, respiratory, ocular, dental, dermatological, and sexually transmitted infections. The development of drug delivery systems based on carriers like nanoparticles (NP) allow to overcome limitations like instability in biological environment, bacterial resistance, and poor cellular penetration. Also, cationic nanocarriers may allow interaction with bacterial membranes or mucus layers to improve the bactericidal action. Aim: To develop and to characterize a drug delivery system for DOXI based on cationic gelatin nanoparticles, intended to mucus delivery for antibacterial therapy. Materials and methods: Gelatin nanoparticles were prepared using the desolvation method, where the effect of stirring speed, concentration of Pluronic F-68, gelatin and volume of crosslinking agent were investigated. The system was characterized by particle size, zeta potential, FT-IR, SEM, Entrapment efficiency and mathematical modeling of *in vitro* profile release. Results and discussion: A stable nanoparticle dispersion  $(\sim 200 \text{ nm}, \text{PDI} > 0.3)$  was obtained with high entrapment efficiency  $({\sim}60\%)$  and cationic surface propperties, whit prolonged release for 8 h. **Conclusions:** The process factors and formulation studied successfully lead a doxycycline loaded cationic gelatin nanoparticles with desirable characteristics. The NP showed prolonged release for 8 hours with an anomalous transport as main mechanism of drug delivery. The system prepared own cationic surface properties to be a possible mucoadhesive system.

Keywords: Doxycycline, gelatin, nanoparticles, modified release.

## **RESUMEN**

### Desarrollo y caracterización de nanopartículas de gelatina de doxiciclina

Introducción: la doxiciclina (DOXI) es un fármaco antimicrobiano de amplio espectro utilizado para infecciones urinarias, intestinales, respiratorias, oculares, dentales, dermatológicas y de transmisión sexual. El desarrollo de sistemas de administración de fármacos basados en acarreadores como las nanopartículas (NP), permiten superar limitaciones como la inestabilidad en el entorno biológico, la resistencia bacteriana y la mala penetración celular. Además, los nanoacarreadores catiónicos pueden permitir la interacción con membranas bacterianas o mucosas para mejorar la acción bactericida. Objetivo: desarrollar y caracterizar un sistema de administración de fármacos para DOXI basado en nanopartículas catiónicas de gelatina, destinado a la administración en mucosas para terapia antibacteriana. Materiales y métodos: se prepararon nanopartículas de gelatina utilizando el método de desolvatación, donde se investigó el efecto de la velocidad de agitación, la concentración de Pluronic F-68, la gelatina y el volumen del agente de entrecruzamiento. El sistema se caracterizó por el tamaño de partícula, potencial zeta, FT-IR, SEM, eficiencia de entrampe y modelado matemático del perfile de liberación *in vitro*. Resultados y discusión: se obtuvo una dispersión estable de nanopartículas ( $\sim$ 200 nm, PDI>0,3) con alta eficiencia de entrampe  $(\sim 60\%)$ , propiedades superficiales catiónicas, y liberación prolongada durante 8 h. Conclusiones: los factores de proceso y la formulación estudiados permitieron obtener nanopartículas catiónicas de gelatina cargadas con DOXI con características deseables. Las NP mostraron liberación prolongada durante 8 horas con un transporte anómalo como principal mecanismo de liberación de fármaco. El sistema preparado presentó propiedades superficiales catiónicas adecuadas para ser un posible sistema mucoadhesivo.

Palabras clave: Doxiciclina, gelatina, nanopartículas, liberación modificada.

### Resumo

## Desenvolvimento e caracterização de nanopartículas de gelatina de doxiciclina

Introdução: a doxiciclina (DOXI) é um antimicrobiano de amplo espectro utilizado para infecções urinárias, intestinais, respiratórias, oculares, dentárias, dermatológicas

e sexualmente transmissíveis. O desenvolvimento de sistemas de liberação de fármacos baseados em carreadores como as nanopartículas (NPs) permite superar limitações como instabilidade no ambiente biológico, resistência bacteriana e baixa penetração celular. Além disso, os nanocarreadores catiônicos podem permitir a interação com membranas bacterianas ou mucosas para aumentar a ação bactericida. Objetivo: desenvolver e caracterizar de um sistema de liberação de fármacos para DOXI baseado em nanopartículas catiônicas de gelatina, destinado à liberação mucosa para terapia antibacteriana. Materiais e métodos: as nanopartículas de gelatina foram preparadas usando o método de dessolvatação, onde o efeito da velocidade de agitação, concentração de Pluronic F-68, gelatina e volume do agente de reticulação foram investigados. O sistema foi caracterizado por tamanho de partícula, potencial zeta, FT-IR, SEM, eficiência de captura e modelagem matemática do perfil de liberação *in vitro*. Resultados e discussão: obteve-se uma dispersão estável de nanopartículas (~200 nm, PDI>0,3) com alta eficiência de captura (~60%), propriedades de superfície catiônica e liberação prolongada por 8 h. Conclusões: os fatores de processo e formulação estudados permitiram a obtenção de nanopartículas de gelatina catiônica carregada com DOXI com características desejáveis. As NPs apresentaram liberação prolongada por 8 horas com transporte anormal como principal mecanismo de liberação do fármaco. O sistema preparado apresentou propriedades de superfície catiônica adequadas para ser um possível sistema mucoadesivo.

Palavras-chave: Doxiciclina, gelatina, nanopartículas, liberação modificada.

#### **INTRODUCTION**

Doxycycline (DOXI) is a semisynthetic wide-spectrum antimicrobial drug tetracycline derivative. Currently, therapeutic applications for DOXI include urinary, intestinal, respiratory, ocular, dental, dermatological, and sexually transmitted infections, but recently DOXI have demosntrated potential application on anti-inflammatory and anticancer treat- ments. DOXI can exist in hydrochloride and hyclate salts forms with high solubility and high permeability according to the biopharmaceutical classification (BCS) [1, 2]. Some limitations that can influence the effectiveness of DOXI include instability in biological environment, bacterial resistance, and poor cellular penetration, which hinder drug action in intracellular pathogens [1]. The development of drug delivery systems based on carriers like micro and nanoparticles (NP) allow to overcome those limitations, since tetracyclines are easily oxidized and degraded the NP provides greater stability. In addition, nanocarriers with positively charged surfaces, for instance, improve electrostatic interaction with the negatively charged bacterial wall

and consequent bactericidal action of antibiotics [1]. Besides, the administration of drugs on mucous membranes offers several advantages, including retention at the site of administration, as well as local administration in order to reduce the systemic dose, reducing adverse effects or conversely promoting the absorption of drugs according to the desired effect [3]. Mucoadhesive carriers adhere to the mucosa increasing the residence time at the administration site, while in mucus penetration, micro or nanoparticles can spread over the mucosa and penetrate deep regions so that their residence time increases considerably, reaching the epithelium of absorption [4]. It has been reported that certain surface properties of nanoparticles influence their mucoadhesive or mucopermeation behavior, especially hydrophilic surfaces such as biopolymers. Among the variety of polymers used for the preparation of carriers is gelatin. Gelatin (GEL) is a biopolymer derived from collagen, which has a positive charge at a pH below its isoelectric point (IP) due to the protonation of amino groups of lysine and arginine residues. It is biocompatible and biodegradable, has high availability and low cost. In addition, the release kinetics can be modified by varying the molecular weight of gelatin, as well as the amount of crosslinking agent during preparation. Gelatin carriers have demonstrated to be suitable for mucus administration, for antibacterial drugs like DOXI. Thus, this works aim to develop and characterize a drug delivery system for Doxycycline based on gelatin, intended to mucus delivery for antibacterial therapy.

## Materials And Methods

#### Materials

Gelatin (type A, gel strength ~225 g Bloom), Pluronic F-68 and Doxycycline Hyclate were purchased from Sigma-Aldrich. Glutaraldehyde (GA) grade I (25% solution in water) was obtained from Merck. Regenerate cellulose dialysis bag (MWCS 12- 14 kDa) was obtained from Spectra/Por, USA. Ultrapure water was obtained with a Barnstead Nanopure diamond system. All buffer solutions were prepared consistent with the USP30-NF25.

#### Nanoparticle formulation

Gelatin nanoparticles were prepared using the desolvation method previously reported in the literature with some modifications [5]. Type A gelatin (GEL) was dissolved in deionized water at 40.0±1.0 °C with constant magnetic stirring at 250 rpm. Subsequently, the gelatin solution was adjusted to a pH of  $3.0\pm0.1$  with HCl 2.0 N and Doxycycline Hyclate (DOXI) was added. On the other hand, Pluronic F-68  $(PF68)$  was dissolved in acetone at a concentration of 3.0% (w/v) at room temperature and constant magnetic stirring. The solution of Pluronic F-68 was added drop by drop to the gelatin solution maintaining stirring with an Ultra Turrax® T18 (IKA). Finally, an aqueous solution of glutaraldehyde (2.0% v/v) was added as a crosslinking agent and kept in constant magnetic stirring for 12 h.

The nanoparticles formulation was optimized in one factor at a time, to obtain a stable dispersion. In this regard, various parameters were investigated for their effects on particle size and PDI. In order to optimize the methodology for obtaining the nanoparticles, the effect of some process parameters were investigated, such as: effect of the stirring speed (1000, 5000, 8000 and 10000 rpm), concentration of Pluronic F-68 (0.5, 1, 2, 3 and 5% w/w), gelatin concentration (10 mg/mL and 20 mg/mL) and volume of the crosslinking agent Glutaraldehyde 2.0 % v/v (100, 300, 500, 1000 and 1500 µL).

#### Nanoparticle characterization

#### *Particle size and Z potential (ξ)*

The hydrodynamic diameter and polydispersity index (PDI) were determined by dynamic light scattering (DLS) technique using a Zetasizer Zen3600 (Malvern Instruments) at 25.0 $\pm$ 1.0 °C with a detection angle of 173° (n=3). The Z potential (ξ) was determined with the same equipment by electrophoretic mobility in deionized water  $(n=3)$ .

#### *Entrapment Efficiency (EE%)*

The entrapment efficiency (%) was determined by centrifugation method or indirect method, by UV-VIS spectrophotometric quantification of doxycycline hyclate contained in the clear supernatant after centrifugation  $(n=3)$  [6]. An aliquot of 1.5 mL of the nanoparticles was placed and centrifuged (Biofuge Primo R) at 14000 rpm for 1.0 h. The absorbance of the supernatant at the maximum absorption wavelength  $(\lambda_{\text{max}}=343.43 \text{ nm})$  was determined with an S2000 spectrophotometer using a DT1000 deuterium light source, with a SAD500 interface (Ocean Optics, Inc.), using a 10 mm long quartz cell (Prolab). A calibration curve of doxycycline hyclate was prepared in water in a range of 0.5 to 20  $\mu$ g/mL from a stock solution (n=3) and the linearity of the results was evaluated by an analysis of variance (ANOVA) using the Statgraphics Centurion XVIII° software. The entrapment efficiency was calculated as follows (equation 1):

$$
Entrapment efficiency = \frac{\text{Total amount of DOX - Free DOX}}{\text{Total amount of DOX}} \times 100 \qquad (\text{Eq. 1})
$$

#### Scanning electron microscopy (SEM)

The morphology of nanoparticles was determined by scanning electron microscopy (SEM) using a JSM-35 CF ( JEOL) microscope at 20.0 kV with gold coating. A drop of freshly prepared NP dispersion was deposited and air-dried at room conditions and coated with gold, finally, the sample was analyzed in vacuum.

### Infrared spectroscopy (FT-IR)

The FT-IR spectra were recorded with a Spectrum 400 FTIR/FIR spectrophotometer (Perkin-Elmer). The infrared spectra of the samples were measured in a wavelength range of 4000-400 cm<sup>-1</sup>.

## *In vitro profile release*

The release profile of the prepared systems was evaluated using the dialysis bag method. An amount of NP dispersion was placed in the regenerated cellulose dialysis bag (MWCS 12-14 kDa, Spectra/Por) previously hydrated in deionized water at room temperature for 12.0 h. The dialysis bag was suspended in the release medium (100.0 mL) of phosphate buffer solution pH 7.0 $\pm$ 0.1 at 37.0 $\pm$ 1 °C in a closed beaker with constant magnetic stirring (250.0 rpm). At specified time intervals, a volume was extracted from the release medium and analyzed using a spectrophotometric method. The medium was replaced by adding an equal amount of fresh medium, which was preheated to  $37.0\pm1.0$  °C, after each sampling to maintain sink conditions. An UV-Vis spectrophotometric quantification was performed at the wavelength of maximum absorbance ( $\lambda_{\text{max}}$ =343.43 nm). The analysis was performed on an S2000 spectrometer using a DT1000 deuterium light source, SAD500 port (Ocean Optics, Inc.) with a 10 mm long quartz cell (Prolab).

## *Mathematical modeling of profile release*

There are many theories or kinetic models to describe drug release from controlled or immediate release forms, where the amount of drug released is a function of time. The release data was fitted to two of the most renowned models to describe drug release kinetics and mechanism.

Higuchi model: Higuchi is related to particles of active dispersed in homogeneous matrices submitted to a diffusing medium. The simplified Higuchi model describes drug release as a diffusion process based in the Fick´s law, square root time dependent (equation 2). This model can be used to describe the drug dissolution from several types of modified release pharmaceutical dosage forms, and is the most widely used model to describe drug release from pharmaceutical matrices. Where  $M_t/M_\infty$  is a fraction of drug releases at time *t*, *k* is the release rate constant. The extend model is based on the hypothesis that (*i*) initial concentration of drug in the system is much higher than the drug solubility, (*ii*) the edge effect is negligible, which means that diffusion is unidirectional, (*iii*) drug molecules are much smaller than system thickness, (*iv*) swelling or dissolution of system is negligible, (*v*) drug diffusivity is constant, and (*vi*) in the release environment perfect sink conditions are maintained [7].

$$
\frac{M_t}{M_\infty} = kt^{\nu^2}
$$
 (Eq. 2)

Korsmeyer-Peppas model (Power Law): This model is a semi-empirical equation developed by Korsmeyer and Peppas to describe drug release from polymeric systems, it can be seen as a generalization of the observation of the superposition of two apparently independent mechanisms of drug transport, relaxation, and diffusion, usually applied when the release mechanism is not known or when more than one type of phenomenon of drug release is involved (equation 3) [7]. Where  $M_t/M_\infty$  is a fraction of drug releases at time *t*, *k* is the release rate constant and *n* is the release exponent. Depending on the value of n that better adjusts to the release profile, concluding for values of *n*= 0.5 the drug release is governed by Fick diffusion, values of *n* between 0.5 and 1.0 corresponds to a non-Fickian transport or anomalous transport, and the mechanism of drug release is governed by diffusion and swelling, and *n*=1.0 corresponds to Case II transport the mechanism driving the drug release is the swelling or relaxation of polymeric chains [7].

$$
\frac{M_{t}}{M_{\infty}} = kt^{n} \tag{Eq. 3}
$$

All fitting models were performed by non-linear direct fitting and analyzed by using DDSolver program as analysis tool. The examination of the goodness of fit statistics was made by comparison of the coefficient of determination or R-square  $(R^2)$ , the adjusted coefficient of determination  $(\mathrm{R}^\mathbf{2}_\mathit{adjusted})$  due to is more meaningful when comparing models with different numbers of parameters.

#### Results and discussion

In recent years, gelatin nanoparticles have proven to be carriers with advantages for modified drug release. Gelatin nanoparticles have been developed by various methods such as coacervation, solvent emulsion-evaporation, nanoprecipitation and desolvation [8]. NP were prepared using the desolvation method, which is a widely used method due to its good reproducibility and ease of preparation. In addition, it allows to modify the characteristics of interest in the NP such as size, *Z* potential, entrapment efficiency, etc. through the manufacturing conditions [9]. It was prepared a solution of gelatin which was adjusted to pH of 3.0±0.1, because in acidic conditions, gelatin

type A (Isoelectric point= 7.0-9.0), presents a positive charge by the protonation of the amino groups  $(\text{-NH}_3{}^+)$  of lysine residues. During synthesis, the positive charge avoids the uncontrolled agglomeration of gelatin chains, allowing to obtain nanoparticles with better characteristics of size and PDI [10]. The desolvation agent (acetone) was used to dehydrate gelatin chains and induce conformational changes in their structure and subsequently induce precipitation of NP [8]. Finally, GA was used as a crosslinking agent to stabilize the structure of the NPs in order to prevent their rapid dissolution in aqueous media and a rapid release of the drug at body temperature. GA is a crosslinking agent that has been extensively studied with proteins, as it reacts easily at room temperature with the simultaneous formation of aldimine bonds (-CH=N-) between the amino groups of proteins [11].

Figure 1 shows the results of formulation studies to obtain NP. The effect of various conditions on manufacturing were investigated, studying one parameter at a time: effect of gelatin concentration (10 mg/mL and 20 mg/mL) (Figure 1a), stirring speed (1000, 5000, 8000 and 10000 rpm) (Figure 1b), Pluronic F-68 concentration (0.5, 1, 2 and 3 w/v %) (Figure 1c), and volume of the crosslinking agent Glutaraldehyde (GA) at 2.0 % v/v (100, 300, 500, 1000 and 1500 µL) (Figure 1d).

Figure 1a showed the results of the concentration of gelatin effect. It was observed that the preparation of NP with a lower concentration of gelatin resulted in a decrease in particle size from 735.3 nm to 341.6 nm, along with a decrease in PDI from 0.224 to 0.208. The low concentration of gelatin allowed the formation of smaller particles with a more homogeneous size distribution. This is due to the lower availability of the biopolymer in the particle formation medium. The gelatin concentration of 10 mg/mL was established and the effect of stirring rate on particle size and PDI was evaluated.

In Figure 1b stirring speed effect was evaluated in particle size (nm) and PDI, in order to obtain a final formulation with particle size  $\leq$  250 nm and polydispersity index  $PDI \leq 0.3$  which indicates a narrow size distribution. The use of homogenizers in the development of nanoparticles by desolvation method is rarely reported, however, Jia Xue and Qixin Zhong reported the use of high-speed homogenization for the preparation of thymol hybrid nano dispersions based on gelatin and lecithin [12]. It was observed that as the speed of agitation increased, the size decreased from 511.4 nm to 219.8 nm and PDI values were observed from 0.343 to 0.088. The results are similar to other results reported in the literature in that it was observed that increasing the speed of agitation favors the arrangement of chains in small droplets that precipitate allowing the formation of smaller particles. The speed of 10,000 rpm was selected in a high-cut homogenizer (Ultra Turrax®).

Subsequently, in Figure 1c the effect of the surfactant Pluronic F-68 was studied, which allows to stabilize the dispersion of particles and reduce the size according to previous studies reported in the literature [5]. At the concentration of Pluronic F-68 from 0.5 and 1%, the formation of NP was not observed but the appearance of large agglomerates which may be due that at low concentrations, there is not enough surfactant to stabilize the formation of NP and there is agglomeration. From 2 to 3% it was observed the NP formation, however, the smaller particles were obtained at the concentration of 3%, since Pluronic F-68 decreases the surface tension between organic and aqueous phase and leads to the formation of smaller solvent droplets [5]. However, at higher concentrations of Pluronic F-68, an increase in particle size was observed, which may be due an excess that may interact with gelatin significantly increasing the particle size.

It is known that in gelatin carriers swelling behavior, mechanical and thermal properties depend on the crosslinking degree [13]. Glutaraldehyde (GA) is a well-known crosslinking agent, it have an aldehyde group (-CHO) that reacts with the amino group of the lysine residues of proteins leading an aldimine linkage (-CH=N-) generating inter and intra covalent bonds [14]. To study the effect of glutaraldehyde as a crosslinking agent with 100, 300, 500, 1000 and 1500 μL aliquots of a 2% v/v aqueous glutaraldehyde solution were added to nanoparticles. After crosslinking reaction, the NP were obtained from  $\sim$ 200 to  $\sim$ 600 nm with PDI from  $\sim$ 0.2 to  $\sim$ 0.3 as shown in Figure 1d. The addition of 100 and 300 μL was studied, however, the formation of a stable dispersion of NP was not observed. It was possible to obtain a stable dispersed system from 500 to 1500 μL. The addition of the crosslinking agent allowed to obtain a stable system with adequate size with a volume of 1500 μL because at lower volumes of Glutaraldehyde the particle size increased which may be due to the agglomeration of the particles.

A decrease in particle size was observed as glutaraldehyde volume increased from 500 to 1500 µL may be related to the intra-particle crosslinking effect. Finally, Glutaraldehyde is consumed during the cross-linking process and with an adequate purification, there is no evidence of *in vivo* toxicity nor adverse effects have been reported, however minimum concentration of crosslinker was utilized [15, 16].

## *Particle size and Z potential (ξ)*

The final formulation presented an average particle size of 214.6 nm with a PDI of 0.090. Figure 2 presents the particle size distribution with the final conditions of the previous formulation studies, where a population with a modal distribution was observed.



Figure 1. Gelatin NP formulation studies

Zeta potential is another important physicochemical parameter that influences the stability of colloidal dispersions. High positive or negative zeta potential values  $(\xi \geq 30 \text{ mV})$  cause strong repellent forces, whereas repulsion between particles with similar electric charge prevents aggregation of the particles and thus ensures easy redispersion [17]. The NP Z potential was  $22\pm1.51$  mV, which is similar to other reports. The nanoparticles were positively charged, this result may indicate the presence of gelatin on the particle surface, according to other reports. The zeta potential was relatively low ( $\xi \le 30$  mV), however, their surface has a sufficient zeta potential to prevent further agglomeration of the particles observed in good size and PDI measurements [10,18]. Also, cationic nanoparticles have demonstrated to improve oral bioavailability of drugs by mucoadhesion [19]. Gastrointestinal tract is an attractive place for mucoadhesive delivery systems to modified the release of drugs. The mucosal surface is negatively charged, a positive charge on the polymer like gelatin NP might facilitate the mucoadhesive process. A positively charged polymer may be attracted to the biologic surface by electrostatic attraction, followed by mechanical interlocking of the polymer chains by

van der Waals force, hydrogen bond, and other forces [19, 20]. Also, positively charged colloidal drug carriers may increase the permeability and potential uptake of slightly soluble drugs [19].



Figure 2. NP Particle size distribution

#### *Entrapment Efficiency (EE%)*

Once the manufacturing conditions for size, PDI and Z potential were determined, the entrapment efficiency of the system was determined with a calibration curve of Doxycycline hyclate from a stock solution, which was linear  $(R^2=0.999)$   $(n=3)$  in a concentration range of 0.5 to 20 μg/mL. A nanoparticulated formulation is preferred to be with high drug loading capacity to reduce the quantity of the carrier required for administration. The amount of bound drug and the nature of interaction between drug and nanoparticles depend on the chemical structure of the drug as well as the polymer and the conditions of drug loading [13]. It is known that hydrophilic drugs may be loaded into gelatin carriers by incubating the drug with aqueous gelatin solution prior to nanoparticle formation. The entrapment efficiency was determined by an indirect method, and the NPs showed a  $59.83\pm1.45\%$  of EE (n=3) which is similar to other reports where DOXI loaded gelatin microspheres were prepared with %EE from 49 to 88% [21].

#### *Scanning electron microscopy (SEM)*

The shape, size and surface morphology of the NPs were determined by SEM. Figure 3 shows the NPs that were obtained with spherical morphology and smooth surface, without cracks or heterogeneities, this morphology is similar to that obtained by other authors for gelatin nanoparticles used in the release of drugs. The NP observed by SEM presented particle sizes similar to the results obtained by the DLS technique (Figure 3a), which is suitable for administration in mucous membranes orally or nasally.





### *Infrared spectroscopy (FT-IR)*

The system obtained together with the individual components (gelatin, Pluronic F68, Doxycycline hyclate) were characterized by FT-IR in the range of 4000 to 400 cm<sup>-1</sup>, with which the characteristic bands of the functional groups of each component were observed. Figure 4 shows the spectra obtained by FT-IR. Pure Doxycycline (DOXI) has characteristic bands in 3331.85 cm<sup>-1</sup> corresponding to the vibrations of the bonds  $(O-H)$  and  $(N-H)$ , in 1663.34 cm<sup>-1</sup> corresponding to the primary amine  $(N-H)$ , 1611.09 cm<sup>-1</sup> of the carbonyl group (C=O), 1458.05 cm<sup>-1</sup> due to the doubling of the group (CH<sub>2</sub>), 1329.80 vibrations (C-H), 1216.67 and 1170.63 cm<sup>-1</sup> indicative of the vibrations of the link (C-N). In the spectrum of gelatin (GEL) the characteristic bands of 3282.08 cm<sup>-1</sup> corresponding to amide A (N-H), 1637.22 cm<sup>-1</sup> of amide I (C=O), 1528.97 cm-1 of amide II (N-H) are observed. In the spectrum of Pluronic F68 (PF68) the characteristic bands in 3505.65, 2880.76 and 1099.18 cm-1 corresponding to the vibrations of the bonds (O-H), (C-H) and (C-O) were observed. In the spectrum of NPs loaded with DOXI (DOXI/NP), bands are observed in 3306.22 cm-1 corresponding to amide A (N-H), 1652.15 cm<sup>-1</sup> of amide I (C=O), indicative of the presence of gelatin, however there is also the appearance of a new band in 1450.58 cm-1 corresponding to the aldimine bond  $(R-CH=N-R')$  which confirms the crosslinking of the amino groups of lysine residues in gelatin with glutaraldehyde (-CHO), the bands indicative of the presence of Pluronic F68 were also observed in 2881.44 and 1102.80  $cm^{-1}$  corresponding to the bonds (C-H) and (C-O) [8, 22]



Figure 4. FT-IR spectroscopy of the NP and individual components

### *In vitro release profile*

*In vitro* release studies were performed using the dialysis bag method. Regenerated cellulose bags of pore size of 12-14 kDa (Spectra/Por®) previously hydrated for 12 h were used. Figure 5 presents the release study, which was maintained for 6 hours at 7.0 $\pm$ 0.1 at 37.0 $\pm$ 1°C it was observed a biphasic patter, since during the first 2 hours the 83.11 $\pm$ 1.78 % (n=3) was released. Subsequently, the system was maintained for 5 hours in which up to  $91.46 \pm 1.47$  % (n=3) was released. The system presented a profile with a quick release during the first 2 h and then a slow release until 6 h. The profile found may be suitable to achieve effective plasma concentrations quickly and continue with a slow release until the second administration. It has been reported that the profile of this type of system can be modified, by increasing the crosslinking agent. In addition, since the drug has a hydrophilic character, it can be deposited on the surface of the system, which favors a rapid release at the beginning of the study named Burst effect, as other authors have already reported.

An adjustment of the data obtained was made to the mathematical models of Higuchi and Korsmeyer-Peppas, which is useful to explain the speed and mechanism of release. It was observed that, of the selected models, the particulate system showed a better fit to the Korsmeyer-Peppas model (60%), with respect to the Higuchi model, according to the highest values of R<sup>2</sup> and R<sup>2</sup> <sub>adj</sub> (Table 1). The Korsmeyer-Peppas model allows to determine the parameter *n*, which is related to different types of release mechanisms, the prepared system presented a value of n of 0.593 which is related to an

anomalous transport mechanism (0.5>n>1.0), which is indicative of a combination of mechanisms of Fickian diffusion and swelling. The drug release kinetics from gelatin nanoparticles depends on the rate of water uptake, drug dissolution/diffusion rate and the polymer glass-rubbery transition including matrix erosion/degradation rate [23]. Gelatin is a biopolymer capable of providing this type of mechanism since when in contact with water the highly soluble drug can dissolve and diffuse through the channels of the particle, as well as present a certain degree of swelling. Due to its properties like particle size, zeta potential and prolonged release, the prepared system could be a good candidate for administration on mucus membranes. Since its high isoelectric point (IP=7-9), in different types of mucus such as buccal (pH 6.5-7.5), nasal (pH 5.5- 6.5), vaginal (pH 3.5-4), etc. the system would remain positive allowing interaction by electrostatic attraction with mucus [4].



Figure 5. NP profile release (experimental data (•)), Higuchi model (blue line), Korsmeyer-Peppas model (red line)





# **CONCLUSIONS**

The process factors and formulation studied successfully lead a doxycycline loaded gelatin nanoparticles with desirable characteristics as low particle size, narrow particle distribution and high drug entrapment efficiency (>50 %). The system was characterized using various instrumental techniques and *in vitro* tests to evaluate its effectiveness. The NP showed prolonged release for 8 hours with an anomalous transport as main mechanism of drug delivery. The system prepared own cationic surface properties to be a possible mucoadhesive system.

### **ACKNOWLEDGMENTS**

The authors would like to acknowledge to posdoctoral program of DGAPA and UNAM for the funding and support of the study.

#### Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

#### **REFERENCES**

- 1. R.C. Feitosa, E.S.A. Ishikawa, M.F.A. da Silva, A.A. da Silva-Júnior, L. Oliveira-Nascimento, Five decades of doxycycline: Does nanotechnology improve its properties?, *Int. J. Pharm.*, 618, 121655 (2022). Doi: [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.ijpharm.2022.121655) [ijpharm.2022.121655](https://doi.org/10.1016/j.ijpharm.2022.121655)
- 2. R. Alshaman, A. Alattar, R.M. El-Sayed, A.R. Gardouh, R.E. Elshaer, A.Y. Elkazaz, M.A. Eladl, M. El-Sherbiny, N.E. Farag, A.M. Hamdan, S.A. Zaitone, Formulation and characterization of doxycycline-loaded polymeric nanoparticles for testing antitumor/antiangiogenic action in experimental colon cancer in mice, *Nanomaterials* (Basel), 12(5), 857 (2022). Doi: [https://doi.org/10.3390/](https://doi.org/10.3390/nano12050857) [nano12050857](https://doi.org/10.3390/nano12050857)
- 3. S.K. Lai, Y.Y. Wang, J. Hanes, Mucus-penetrating nanoparticles for drug and gene delivery to mucosal tissues, *Adv. Drug Deliv. Rev.*, 61, 158–171 (2009). Doi: https://doi.org/10.1016/j.addr.2008.11.002.
- 4. K. Netsomboon, A. Bernkop-Schnürch, Mucoadhesive vs. mucopenetrating particulate drug delivery, *Eur. J. Pharm. Biopharm.*, 98, 76–89 (2016). Doi: <https://doi.org/10.1016/j.ejpb.2015.11.003>
- 5. A.A. Kharia, A.K. Singhai, Formulation and evaluation of gastroretentive drug delivery system of acyclovir as mucoadhesive nanoparticles, *Int. J. PharmTech Res.*, 5, 1538–1545 (2013).
- 6. V. Pokharkar, V. Patil, L. Mandpe, Engineering of polymer-surfactant nanoparticles of doxycycline hydrochloride for ocular drug delivery, *Drug Deliv.*, 22, 955–968 (2015).<https://doi.org/10.3109/10717544.2014.893381>
- 7. M.L. Bruschi, Mathematical models of drug release, in: *Strategies to Modify the Drug Release from Pharmaceutical Systems*, Woodhead Publishing, 2015. Doi: <https://doi.org/10.1016/b978-0-08-100092-2.00005-9>
- 8. A.O. Elzoghby, Gelatin-based nanoparticles as drug and gene delivery systems: Reviewing three decades of research, *J. Control. Release*, 172, 1075–1091 (2013). Doi: <http://dx.doi.org/10.1016/j.jconrel.2013.09.019>
- 9. S.M. Ahsan, C.M. Rao, The role of surface charge in the desolvation process of gelatin: Implications in nanoparticle synthesis and modulation of drug release, *Int. J. Nanomed.*, 12, 795–808 (2017). Doi: https://doi.org/10.2147/IJN. S124938.
- 10. S. Jain, P.U. Valvi, N.K. Swarnakar, K. Thanki, Gelatin coated hybrid lipid nanoparticles for oral delivery of Amphotericin B, *Mol. Pharm.*, 9, 2542–2553 (2012). Doi: <https://doi.org/10.1021/mp300320d>
- 11. S. Farris, J. Song, Q. Huang, Alternative reaction mechanism for the cross-linking of gelatin with glutaraldehyde, *J. Agric. Food Chem.*, 58, 998–1003 (2010). Doi: <https://doi.org/10.1021/jf9031603>
- 12. J. Xue, Q. Zhong, Blending lecithin and gelatin improves the formation of thymol nanodispersions, *J. Agric. Food Chem.*, 62, 2956–2962 (2014). Doi: [https://](https://doi.org/10.1021/jf405828s) [doi.org/10.1021/jf405828s](https://doi.org/10.1021/jf405828s)
- 13. A. Kumari, S.K. Yadav, S.C. Yadav, Biodegradable polymeric nanoparticles based drug delivery systems, *Colloids and Surfaces B: Biointerfaces*, 75(1), 1–18 (2010). Doi: <https://doi.org/10.1016/j.colsurfb.2009.09.001>
- 14. J.B. Science, T. Nguyen, B. Lee, Fabrication and characterization of cross-linked gelatin, *Journal of Biomedical Science and Engineering*, 3, 1117–1124 (2010). Doi: <https://doi.org/10.4236/jbise.2010.312145>
- 15. P. Lai, W. Daear, R. Löbenberg, E.J. Prenner, Overview of the preparation of organic polymeric nanoparticles for drug delivery based on gelatine, chitosan, poly(d,l-lactide-co-glycolic acid) and polyalkylcyanoacrylate, *Colloids and Surfaces B: Biointerfaces*, 118, 154–163 (2014). Doi: [https://doi.org/10.1016/j.col](https://doi.org/10.1016/j.colsurfb.2014.03.017)[surfb.2014.03.017](https://doi.org/10.1016/j.colsurfb.2014.03.017)
- 16. K.J. Geh, M. Hubert, G. Winter, Optimization of one-step desolvation and scale-up of gelatin nanoparticle production, *J. Microencapsul.*, 33(7), 595–604 (2016). Doi:<https://doi.org/10.1080/02652048.2016.1228706>
- 17. J.A. Carvalho, A.S. Abreu, V.T.P. Ferreira, E.P. Gonçalves, A.C. Tedesco, J.G. Pinto, J. Ferreira-Strixino, M. Beltrame Junior, A.R. Simioni, Preparation of gelatin nanoparticles by two step desolvation method for application in photodynamic therapy, *J. Biomater. Sci. Polym. Ed.*, 29, 1287–1301 (2018). Doi: [https://](https://doi.org/10.1080/09205063.2018.1456027) [doi.org/10.1080/09205063.2018.1456027](https://doi.org/10.1080/09205063.2018.1456027)
- 18. S. Azarmi, Y. Huang, H. Chen, S. McQuarrie, D. Abrams, W. Roa, W.H. Finlay, G.G. Miller, R. Löbenberg, Optimization of a two-step desolvation method for preparing gelatin nanoparticles and cell uptake studies in 143B osteosarcoma cancer cells, *J. Pharm. Pharm. Sci.*, 9, 124–132 (2006).
- 19. M.H. El-Shabouri, Positively charged nanoparticles for improving the oral bioavailability of cyclosporin-A, *Int. J. Pharm.*, 249, 101–108 (2002). Doi: [https://](https://doi.org/10.1016/S0378-5173(02)00461-1) [doi.org/10.1016/S0378-5173\(02\)00461-1](https://doi.org/10.1016/S0378-5173(02)00461-1)
- 20. J. Wang, Y. Tauchi, Y. Deguchi, K. Morimoto, Y. Tabata, Y. Ikada, Positively charged gelatin microspheres as gastric mucoadhesive drug delivery system for eradication of *H. pylori*, *Drug Deliv.*, 7, 237–243 (2000). Doi: [https://doi.](https://doi.org/10.1080/107175400455173) [org/10.1080/107175400455173](https://doi.org/10.1080/107175400455173)
- 21. D.V. Way, M. Nele, J.C. Pinto, Production of doxycycline-loaded gelatin microspheres through thermal treatment in inverse suspensions, *Polym. Eng. Sci.*, 58, 802–809 (2018). Doi: <https://doi.org/10.1002/pen.24628>
- 22. G.M. El Maghraby, A.H. Alomrani, Synergistic enhancement of itraconazole dissolution by ternary system formation with pluronic F68 and hydroxypropylmethylcellulose, *Sci. Pharm.*, 77(2), 401–417 (2009). Doi: [https://doi.](https://doi.org/10.3797/scipharm.0901-08) [org/10.3797/scipharm.0901-08](https://doi.org/10.3797/scipharm.0901-08)

23. A.K. Gupta, M. Gupta, S.J. Yarwood, A.S.G. Curtis, Effect of cellular uptake of gelatin nanoparticles on adhesion, morphology and cytoskeleton organisation of human fibroblasts, *J. Control. Release*, 95, 197–207 (2004). [https://doi.](https://doi.org/10.1016/j.jconrel.2003.11.006) [org/10.1016/j.jconrel.2003.11.006](https://doi.org/10.1016/j.jconrel.2003.11.006)

# How to cite this article

P. García-Guzmán, L. Ocampo, L. Gutierrez, H. Sumano, M.J. Bernad, Development and characterization of doxycycline gelatin nanoparticles, *Rev. Colomb. Cienc. Quim. Farm.*, 52(2), 640-657 (2023). https://doi.org/10.15446/rcciquifa.v52n2.110730