Mechanisms involved in resistance to carbapenems among *Acinetobacter baumannii* isolates recovered in Brazil: A systematic review and meta-analysis

Adrielle Pieve de Castro¹, William Gustavo Lima¹⁶, Cristina Sanches¹⁶, Magna Cristina de Paiva¹²*¹

¹ Laboratório de Diagnóstico Laboratorial e Microbiologia Clínica. Universidade Federal de São João del-Rei, Campus Centro-Oeste Dona Lindu, Divinópolis, Minas Gerais, Brasil.

² Laboratório de Diagnóstico Microbiológico, Campus Centro Oeste Dona Lindu/ Universidade Federal de São João Del-Rei. 400 Sebastião Gonçalves Coelho street, Chanadour, Divinópolis, Minas Gerais, Brazil, CEP: 35501-293.

*Corresponding author E-mail address: magnacpaiva@ufsj.edu.br

ORCID IDs: ¹0000-0002-1260-5619, ¹⁶0000-0001-8946-9363, ¹⁰0000-0002-8562-1337, ¹⁰0000-0001-9375-7261

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**Summary**

**Introduction:** Infections caused by carbapenem-resistant *Acinetobacter baumannii* (CRAB) is a health problem due to the limited therapeutic options available. This study was carried out to evaluate the main mechanisms of resistance of carbapenems in CRAB in the last 10 years in Brazil and to describe the susceptibility profile to tigecycline and polymyxins in these isolates. **Material and methods:** A systematic review was carried out according to Prisma in PUBMED/MEDLINE, Scopus, SciELO, Biblioteca Virtual de Saúde (BVS) and Cochrane Library. Data regarding enzyme resistance to carbapenems were evaluated by meta-analysis according to the random effect. **Results:** 21 articles were selected according to inclusion and exclusion criteria that evaluated 1096 CRAB. Most of the studies were carried out in the southern (33.3 %) and southeast (23.8 %) regions of Brazil (33.3 %) and in 2016 and 2018. According to the meta-analyses, OXA-type carbapenemase was the main mechanism involved in the low susceptibility to carbapenems in CRAB (98%; 95% CI: 0.91, 0.99; I² = 95%), with *bla*$_{OXA-23-24}$ (91 %; 95 % CI: 0.76; 0.97; I² = 97 %).
or bla\text{OXA-51-like} / \text{ISAba1} (84 %; 95 % CI: 0.15, 0.99; I^2 = 98 %) genes, followed by metallo-β-lactamas (MBL) (12 %, 95 % CI: 0.09, 0.15, I^2 = 99 %) and \textit{Klebsiella pneumoniae} carbapenemase (KPC) (6 %, 95 % CI: 0.04; 0.08; I^2 = 87 %). **Conclusion:** The included studies showed that susceptibility to colistin (99 %) and tigecycline (93 %) remains high and was not affected by carbapenem resistance.

**Keywords:** \textit{Acinetobacter baumannii}, Brazil, resistance, carbapenem, tigecycline, colistin.

**RESUMEN**

Mecanismos involucrados en la resistencia a carbapenémicos entre aislamientos de \textit{Acinetobacter baumannii} recuperados en Brasil: una revisión sistemática y metanálisis

**Introducción:** las infecciones por \textit{Acinetobacter baumannii} resistente a carbapenémicos (CRAB) es un problema de salud debido a las limitadas opciones terapéuticas disponibles. Este estudio se realizó para evaluar los principales mecanismos de resistencia de los carbapenémicos en CRAB en los últimos 10 años en Brasil y describir el perfil de susceptibilidad a tigeciclina y polimixinas en estos aislados.

**Material y métodos:** se realizó una revisión sistemática de acuerdo con Prisma en PUBMED/MEDLINE, Scopus, SciELO, Biblioteca Virtual de Saúde (BVS) y Cochrane Library. Los datos referentes a resistencia enzimática a los carbapenémicos se evaluaron mediante metaanálisis según el efecto aleatorio. **Resultados:** se seleccionaron 21 artículos según criterios de inclusión y exclusión que evaluaron 1.096 CRAB. La mayoría de los estudios se llevaron a cabo en las regiones sur (33,3%) y sureste (23,8 %) de Brasil (33,3 %) y en los años 2016 y 2018. Según los metaanálisis, la carbapenemasa tipo OXA fue el principal mecanismo implicado en la baja susceptibilidad a los carbapenémicos en CRAB (98 %; IC 95 %: 0.91; 0.99; I^2 = 95 %), con bla\text{OXA-23-like} (91 %; 95 % CI: 0.76; 0.97; I^2 = 97 %) o bla\text{OXA-51-like} / \text{ISAba1} (84 %; 95 % CI: 0.15; 0.99 ; I^2 = 98 %) genes, seguida de metalo-β-lactamasas (MBL) (12 %; IC95 %: 0.09; 0.15; I^2 = 99 %) y \textit{Klebsiella pneumoniae} carbapenemase (KPC) (6 %; IC95 %: 0.04; 0.08; I^2 = 87 %). **Conclusión:** los estudios incluidos mostraron que la susceptibilidad a la colistina (99 %) y tigeciclina (93 %) sigue siendo alta y no se ve afectada por la resistencia a los carbapenémicos.

**Palabras clave:** \textit{Acinetobacter baumannii}, Brasil, resistencia, carbapenem, tigeciclina, colistina.
Mechanisms in resistance to carbapenems among A. baumannii isolates

Resumo

Mecanismos envolvidos na resistência a carbapenêmicos entre isolados de Acinetobacter baumannii recuperados no Brasil: Uma revisão sistemática e metanálise

Introdução: as infecções causadas por Acinetobacter baumannii resistente aos carbapenêmicos (CRAB) são um problema de saúde devido às limitadas opções terapêuticas disponíveis. Este estudo foi realizado para avaliar os principais mecanismos de resistência aos carbapenêmicos em CRAB nos últimos 10 anos no Brasil e descrever o perfil de susceptibilidade à tigeciclina e às polimixinas nesses isolados.

Material e métodos: foi conduzida uma revisão sistemática segundo o Prisma nas bases de dados PUBMED/MEDLINE, Scopus, SciELO, Biblioteca Virtual de Saúde (BVS) e Biblioteca Cochrane. Os dados relativos à resistência enzimática aos carbapenêmicos foram avaliados por meta-análises de acordo com o efeito aleatório.

Resultados: foram selecionados 21 artigos de acordo com os critérios de inclusão e exclusão que avaliaram 1.096 CRAB. A maioria dos estudos foi realizada nas regiões Sul (33,3 %) e Sudeste (23,8 %) do Brasil e nos anos de 2016 e 2018. De acordo com as metanálises, a carbapenemase do tipo OXA foi o principal mecanismo envolvido na baixa susceptibilidade aos carbapenêmicos em CRAB (98 %; 95% IC: 0,91, 0,99; I² = 95 %), com blaOXA-23-like (91 %; 95 %; IC: 0,76; 0,97; I² = 97 %) ou blaOXA-51-like / ISAba1 genes, seguidos por metalo-β-lactamases (MBL) (12 %, 95 % IC: 0,09, 0,15, I² = 99 %) e Klebsiella pneumoniae carbapenemase (KPC) (6 %, IC 95 %: 0,04; 0,08; I² = 87 %).

Conclusão: os estudos incluídos mostraram que a susceptibilidade à colistina (99 %) e tigeciclina (93 %) permanece alta e não foi afetada pela resistência aos carbapenêmicos.

Palavras-chave: Acinetobacter baumannii, Brasil, resistência, carbapenem, tigeciclina, colistin.

Introduction

Acinetobacter baumannii is a ubiquitous, non-glucose-fermenting, Gram-negative, oxidase negative, aerobic, pleomorphic, and non-motile coccobacillus (typically 1.0–1.5 µm by 1.5–2.5 µm in size) responsible for a significant proportion of healthcare-associated infections (HAIs) worldwide [1, 2]. The HAIs caused by A. baumannii mainly affect immunocompromised individuals, and are especially frequent in the Intensive Care Unit (ICU) acquired infections [1-4]. This pathogen has been associated with
several types of infections, including pneumonia (mostly mechanical ventilator-associated), urinary tract infections, bacteremia, osteomyelitis, skin and soft tissue infections, and meningitis [5, 6]. Generally, these infections are caused by strains of *A. baumannii* that are resistant to the various antibiotics of choice, making pharmacological therapy a major challenge for healthcare professionals worldwide [6]. The presence of multi-drug resistance (MDR) is common among isolates of *A. baumannii* from hospitals, limiting treatment options and leading to increased morbidity and mortality of HAIs caused by this pathogen [3, 6].

Carbapenem antibiotics such as imipenem, meropenem, doripenem, but not ertapenem, are considered an important treatment option for MDR-*A. baumannii* infection. However, a considerable increase in the number of CRAB strains has been documented worldwide, inclusive in Brazil [7], and the extensive-drug resistance (XDR) phenotype stands out among the isolates of the *A. baumannii*. This critical panorama has stimulated the World Health Organization (WHO) to classify CRAB as a top priority organism for research and development of new antibiotics, with the aim to control the rapid and worrying advance of this bacterium in hospital environments [8, 9].

Resistance to carbapenems can occur through the combination of different mechanisms, such as decreased permeability of external membranes, alteration of the affinity of penicillin-binding proteins (PBPs), overexpression of efflux pumps, and production of carbapenem-hydrolyzing β-lactamases (carbapenemases) [10]. The main mechanism of resistance to carbapenems between *A. baumannii* types is the overexpression of carbapenemases. Three types of carbapenemases have been recognized in this Gram-negative bacterium, namely Ambler class A β-lactamases (i.e., GES-14 and KPC), Ambler class B metallo-β-lactamases (e.g., IMP, VIM, SIM-1, and NDM) and Ambler class D oxacillinas (OXAs) [10]. Among the different carbapenemase types, the OXAs play an important role in the resistance of *A. baumannii* isolates to carbapenems, and has been reported in several countries as the main determinant of resistance [11]. It has been described in different families in this species: intrinsic, chromosomally located and also found in plasmid OXA-51-likes (Neves *et al.*, 2016); and the acquired OXA-23-like, OXA-40-like (formally OXA-24-like), OXA-58-like, OXA-143-like, and OXA-235-like [12, 13]. The presence of insertion sequences (IS) immediately upstream of *bla*OXA genes is known to induce the overexpression of OXA-51, OXA-23, or OXA-58, and to generate carbapenem resistance at a high level in *A. baumannii*.

The treatment of HAIs caused by CRAB is limited to the use of antimicrobials such as of polymixins or tigecycline [14]; however, it is known that the infusion of these drugs in monotherapy can favor the selection of strains with intermediate or complete resistance, especially to tigecycline [15, 16]. Associated with this, the emergence of a deter-
Mechanisms in resistance to carbapenems among A. baumannii isolates

The emergence of resistance to carbapenems in A. baumannii is highly significant in emerging countries such as Brazil. Since the first description, the incidence and prevalence of CRAB have increased in this country, especially in the last decade [18, 19]. However, despite the large number of reports of these infections, there are still no studies aiming to define the overall profile on infection rates and resistance of these isolates in Brazil, making it difficult to design national infection control measures to combat the spread of CRAB. Thus, this study aimed to evaluate, through a systematic review of the biomedical literature and meta-analysis of recovered data, the susceptibility profile to tigecycline and polymyxins of CRAB recovered over the last 10 years in Brazil. In addition, this study also describes the main mechanism of resistance to carbapenems amongst Brazilian CRAB isolates, and determines the frequency of each type of carbapenemase, identified through the analysis of subgroups.

Materials and methods

A systematic review and meta-analysis was conducted according to the Cochrane Handbook guidelines [20]. In order to conduct the review, the steps of searching, selecting, extracting the data of interest, and analyzing results were performed according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (Prisma) statement [21].

Search strategies

A systematic search was conducted using PUBMED/MEDLINE, Scopus, Scientific Electronic Library Online (SciELO), Web of Science, Cochrane library, LILACS, and Virtual Health Library (VHL) databases, for articles published in English or Portuguese, using the following Medical Subject Heading (MeSH) terms and key words: (“Acinetobacter” OR “Acinetobacter baumannii” OR “Acinetobacter Infections” OR “Infections, Acinetobacter” OR “Acinetobacter Infection*”) AND “Brazil”. All details of the search strategy are showed in the supplementary file. Additionally, we screened the reference lists of all included studies and relevant systematic reviews, in order to
identify additional eligible studies. The search was conducted up to October 18, 2019, and no date limits were established.

Inclusion and exclusion criteria

Articles reporting CRAB infections among hospitalized patients in Brazil were included using the PEOS strategy, as follows: “Population”, bacterial isolates from patients of both sexes and all age ranges, recovered from patients hospitalized in Brazil; “Exposition”, infection by CRAB; “Outcomes”, resistance against polymyxins (i.e., polymyxin B and colistin), and tigecycline and carbapenemases type identified; “Study design”, observational studies and hospital epidemiologic surveillance programs.

Studies were included on the conditions that: (i) clinical species such as *A. baumannii* were correctly isolated and characterized; (ii) the studies have evaluated a susceptibility pattern to polymyxins, using standard broth macro- or micro-dilution methods, according to the Clinical Laboratory Standard Institute guidelines (CLSI) or European Committee on Antimicrobial Susceptibility Testing (EUCAST); (iii) the article showed original material and was written in English or Portuguese; and (iv) it was published from 2009 onwards. Review articles, e-mails and editorials were excluded, as well as: (i) articles conducted in Brazil, but which used isolates from other countries; (ii) studies that did not show the identification method used for *A. baumannii* or the resistance mechanism; and (iii) articles that did not identify the genus/species of the isolates. In the event that the article complied with the inclusion criteria but the full text was not available, the corresponding author was contacted by e-mail up to three times (with intervals of 14 days), and the study was included if it was sent before the final contact.

Study selection

The titles, abstracts and keywords were first analyzed by two independent authors (A.P.C. and W.G.L.), in accordance with the inclusion criteria. Next, the pre-selected articles were subjected to a full-text evaluation, in order to decide whether to include or exclude the study. If there was disagreement between authors, a third researcher (M.C.P.) was consulted by consensus, and the kappa coefficient (performed with 95% confidence interval) was used to analyze the degree of agreement between evaluators [22].

Data analyses

Articles that filled all inclusion criteria were submitted to an analytical full-text reading, and the following data were extracted: (i) reference (authorship and year); (ii) period and place (city and country) of data collection; (iii) type, origin and character-
Mechanisms in resistance to carbapenems among *A. baumannii* isolates

**Results and discussion**

*A. baumannii* is a pathogen particularly important as a causative agent of HAIs. The emergence of resistance to carbapenems is notable in this bacterium, and the frequency of carbapenem-resistant *A. baumannii* (CRAB) is very high in emerging countries, such as Brazil [2, 4, 18]. These isolates typically exhibit resistance to other classes of antimicrobials, such as quinolones, aminoglycosides, and other β-lactam compounds [19]. Thus, with few therapeutic options to treat infections by CRAB, the only option is the use of tigecycline or polymyxins. Here, we investigated the susceptibility profile to tigecycline and polymyxins of CRAB recovered over the last 10 years in Brazil, and described the main mechanisms of resistance to carbapenems that circulate among these isolates.

The search process in the databases resulted in 882 articles; 433 in Pubmed, 317 in Scopus, 6 in Cochrane Library, 95 in Virtual Health Library, and 31 in SciELO (figure 1). After identifying and excluding repeated articles between the databases, 69 studies were obtained and analyzed according to the eligibility criteria. From these articles, 48 were excluded, and the main exclusion moieties were studies that did not identify
Acinetobacter at the species level, or included Acinetobacter non-baumannii (n=13); articles prior to 2009 (n=26); and studies designed without inclusion criteria as revision articles (n=6) (figure 1). Furthermore, two studies that did not include the resistance profile dates and one article which studied only isolates from colonized patients were also excluded from this review. Finally, 21 studies that met the eligibility criteria were selected for extraction of the variables of interest. The degree of agreement between two authors was considered substantial, according to kappa test (kappa=0.764).

Figure 1. Flowchart of the selected studies for the systematic review according to the Prisma criteria.

Studies on carbapenem-resistant *A. baumannii* have been further explored in recent years around the world. For instance, the number of studies in PubMed reporting
CRAB has increased from a single report in 2000 to over 266 reports in 2018, highlighting the global dissemination and good adaptation of this pathogen [23-43]. In the present study, most articles were published in 2016 and 2018. The increased interest in studying and investigating CRAB isolates, in response to the great clinical impact and advancing microbial identification techniques over the last five years (particularly with the introduction of MALDI-TOF/MS in clinical microbiology research centers), justifies the number of publications peaking in those years.

A total of 1,096 CRAB isolates, obtained from patients between 2009 and 2019, were analyzed in the selected studies (table 1). All included studies recovered one isolate per patient. Most studies were published in 2016 (6/21; 28.6 %) [11, 23-27] and 2018 (5/21; 23.8 %) [12, 13, 28-30] and the majority were conducted in the south region of the country (7/21; 33.3%) [12, 13, 23, 31-34], followed by the southeast (5/21; 23.8 %) [24, 27, 29, 31, 34], mid-west (3/21, 14.3 %) [30, 35, 36] and northeast (3/21, 14.3 %) [25, 26, 37]. Two studies involved different Brazilian regions (9.5 %) [38, 39]. The prospective design (9/21; 42.8 %) [13, 23, 25, 27, 29, 36, 37, 40, 41] was more common than retrospective (5/21; 23.8 %) [31, 32, 34, 38, 39]. Five case reports were also included (5/21; 23.8 %) [11, 24, 26, 30, 33].

Regarding the states where the studies were conducted, the most frequent were Parana [13, 33, 34] and Minas Geraias [27, 29, 35], each with 3 studies (3/21; 14.3 %). The state of Paraná was where the first CRAB outbreak occurred in two different hospitals, making this a “model state” to study the resistance dynamics of this XDR bacterium in Brazil (Anvisa, 2013).

The most-used method for microbial identification was the detection of the gene \( \text{bla}_{OXA-51} \) (18/21; 85.7 %) [12, 13, 23-25, 27, 29-37, 39, 40], which is intrinsic in \( A. \text{baumannii} \). The detection of \( \text{bla}_{OXA-51-like} \) can be used as a simple and reliable way of identifying \( A. \text{baumannii} \), as this gene is found in virtually every isolate of this species [42-44]. However, the \( \text{bla}_{OXA-51-like} \) gene has also been found in other species from the \( \text{Acinetobacter calcoaceticus-A. baumannii} \) (Acb) complex, such as \( \text{Acinetobacter nosocomialis} \) [44].

Therefore, it is necessary to incorporate additional identification methods, and the combination of the automated VITEK-2\(^* \) (bioMerieux, France) system with molecular biology (i.e., identification of \( \text{gyrA}, \text{rpoB}, \) or \( \text{bla}_{OXA-51-like} \) gene) was the second-most used form of bacterial identification in the studies included in this review (15/21; 71.4 %) [12, 13, 23-25, 27, 30-35, 39, 40]. In contrast to the VITEK-2\(^* \) system alone, which could not correctly identify \( A. \text{baumannii} \) and failed in predicting carbapenem suscep-
Table 1. Main characteristics of included studies.

<table>
<thead>
<tr>
<th>Reference</th>
<th>Regions, period and design of study</th>
<th>Dates of isolates</th>
<th>Total of CRAB</th>
<th>Total of PRAB</th>
<th>Total of TRAB</th>
<th>Molecular mechanisms of resistance (n)</th>
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</thead>
<tbody>
<tr>
<td>da Silva et al., 2018 [30]</td>
<td>· Dourados, MS · Case-control · 2013-2015</td>
<td>· Vitek2 · MALDI-TOF MS · Identification of ( \text{bla}_{\text{OXA-51}} )</td>
<td>41</td>
<td>0</td>
<td>18</td>
<td>· ( \text{ISAba}<em>1 ) was found upstream both ( \text{bla}</em>{\text{OXA-23}} ) and ( \text{bla}_{\text{OXA-51}} ) genes in all isolates</td>
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<td>· Absence of ( \text{bla}<em>{\text{IMP}}, \text{bla}</em>{\text{NDM}}, \text{bla}<em>{\text{VIM}}, \text{bla}</em>{\text{KPC}}, \text{bla}<em>{\text{OXA-48}}, \text{bla}</em>{\text{OXA-58}}, \text{bla}<em>{\text{OXA-143}} ) and ( \text{bla}</em>{\text{OXA-24/40}} )</td>
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<tr>
<td>Castilho, et al., 2017 [36]</td>
<td>· Goiânia, GO · Prospective · 2010</td>
<td>· Vitek2 · Gram staining · Biochemical test · Identification of ( \text{bla}_{\text{OXA-51}} )</td>
<td>49</td>
<td>6</td>
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<td>· ( \text{ISAba}<em>1/\text{bla}</em>{\text{OXA-23}} ) (n=17)</td>
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<td>· ( \text{bla}_{\text{OXA-58}} ) (n=3)</td>
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<td>· ( \text{ISAba}<em>1/\text{bla}</em>{\text{OXA-51}} ) (n=17)</td>
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<tr>
<td>Cortivo et al., 2015 [31]</td>
<td>· Joinville, SC · Retrospective · 2011-2013</td>
<td>· AutoScan™ · Gram staining · Identification of ( \text{bla}_{\text{OXA-51}} )</td>
<td>118</td>
<td>0</td>
<td>14 (1)</td>
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<td>· ( \text{bla}_{\text{OXA-23}} ) (n=103)</td>
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<td>· Absence of ( \text{bla}<em>{\text{OXA-58}}, \text{bla}</em>{\text{OXA-143}} ) and ( \text{bla}_{\text{OXA-24/40}} )</td>
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<tr>
<td>Rocha et al., 2017 [39]</td>
<td>· Porto Alegre, RS (n=31); Rio de Janeiro, RJ (n=28); Curitiba, PR (n=21); São Paulo, SP (n=12) · Retrospective · 2010</td>
<td>· Vitek2 · MALDI-TOF MS · Identification of ( \text{gyrB} ) · Identification of ( \text{bla}_{\text{OXA-51}} )</td>
<td>91</td>
<td>3</td>
<td>0</td>
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<td>· ( \text{bla}_{\text{OXA-23}} ) (n=80)</td>
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<td>· ( \text{bla}_{\text{OXA-24/40}} ) (n=12)</td>
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<td>· Absence of ( \text{bla}<em>{\text{OXA-58}} ) and ( \text{bla}</em>{\text{OXA-143}} )</td>
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<td>Neves et al., 2016 [27]</td>
<td>· Belo Horizonte, MG · Prospective · 2009-2010</td>
<td>· Vitek™ · Identification of ( \text{bla}_{\text{OXA-51}} )</td>
<td>56</td>
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<td></td>
<td>· ( \text{bla}_{\text{OXA-23}} ) (n=22)</td>
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<td>· ( \text{bla}_{\text{OXA-143}} ) (n=8)</td>
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<td>· Absence of ( \text{bla}<em>{\text{IMP}}, \text{bla}</em>{\text{VIM}}, \text{bla}<em>{\text{OXA-58}} ) and ( \text{bla}</em>{\text{OXA-24/40}} )</td>
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<th>Dates of isolates</th>
<th>Molecular mechanisms of resistance (n)</th>
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<tr>
<td>Pagano et al., 2018 [12]</td>
<td>Porto Alegre, RS · Vitek&lt;sup&gt;*&lt;/sup&gt; · Identification of bla&lt;sub&gt;OXA-51&lt;/sub&gt;</td>
<td>Total of CRAB: 49 Total of PRAB: - Total of TRAB: -</td>
<td>OXA-23</td>
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<td>Moreira et al., 2018 [13]</td>
<td>Maringá, PR · BD Phoenix system · Identification of bla&lt;sub&gt;OXA-51&lt;/sub&gt;</td>
<td>Total of CRAB: 31 Total of PRAB: 0 Total of TRAB: -</td>
<td>bla&lt;sub&gt;OXA-23&lt;/sub&gt; (n=29) · Absence of bla&lt;sub&gt;IMP&lt;/sub&gt;, bla&lt;sub&gt;VIM&lt;/sub&gt;, bla&lt;sub&gt;GIM&lt;/sub&gt;, bla&lt;sub&gt;SPM&lt;/sub&gt;, bla&lt;sub&gt;SIM&lt;/sub&gt;, bla&lt;sub&gt;OXA-58&lt;/sub&gt;, and bla&lt;sub&gt;OXA-24/40&lt;/sub&gt;</td>
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<tr>
<td>Tavares et al., 2019 [28]</td>
<td>Botucatu, SP · Biochemical test&lt;sup&gt;3&lt;/sup&gt; · Gram staining · Identification of bla&lt;sub&gt;OXA-51&lt;/sub&gt; · Identification of gltA · Identification of rpoB</td>
<td>Total of CRAB: 107 Total of PRAB: 0 Total of TRAB: 41 (1)</td>
<td>ISAba1/bla&lt;sub&gt;OXA-23&lt;/sub&gt; (n=104) · bla&lt;sub&gt;OXA-41&lt;/sub&gt; (n=2) · bla&lt;sub&gt;OXA-24/40&lt;/sub&gt; (n=1) · Absence of bla&lt;sub&gt;KPC&lt;/sub&gt;, bla&lt;sub&gt;NDM&lt;/sub&gt;, bla&lt;sub&gt;IMP&lt;/sub&gt;, bla&lt;sub&gt;VIM&lt;/sub&gt;, bla&lt;sub&gt;SPM&lt;/sub&gt;, bla&lt;sub&gt;OXA-58&lt;/sub&gt;, and bla&lt;sub&gt;OXA-48&lt;/sub&gt;</td>
</tr>
<tr>
<td>Pagano et al., 2016 [11]</td>
<td>Porto Alegre, RS · Vitek&lt;sub&gt;2&lt;/sub&gt; · MALDI-TOF MS · Identification of gyrB</td>
<td>Total of CRAB: 2 Total of PRAB: 0 Total of TRAB: 0</td>
<td>bla&lt;sub&gt;OXA-24/40&lt;/sub&gt; (n=2) · bla&lt;sub&gt;OXA-72&lt;/sub&gt; (n=2) · Absence of bla&lt;sub&gt;KPC&lt;/sub&gt;, bla&lt;sub&gt;NDM&lt;/sub&gt;, bla&lt;sub&gt;IMP&lt;/sub&gt;, bla&lt;sub&gt;VIM&lt;/sub&gt;, bla&lt;sub&gt;SPM&lt;/sub&gt;, bla&lt;sub&gt;OXA-58&lt;/sub&gt;, and bla&lt;sub&gt;OXA-48&lt;/sub&gt;</td>
</tr>
<tr>
<td>Fonseca et al., 2019 [40]</td>
<td>Boa Vista, RR · Vitek&lt;sub&gt;2&lt;/sub&gt; · Identification of bla&lt;sub&gt;OXA-51&lt;/sub&gt; · 16S rRNA gene</td>
<td>Total of CRAB: 32 Total of PRAB: 0 Total of TRAB: -</td>
<td>bla&lt;sub&gt;OXA-41&lt;/sub&gt; (n=2) · bla&lt;sub&gt;OXA-72&lt;/sub&gt; (n=2)</td>
</tr>
<tr>
<td>Pillonetto et al., 2014 [32]</td>
<td>Londrina, PR · Vitek&lt;sub&gt;2&lt;/sub&gt; · Identification of bla&lt;sub&gt;OXA-51&lt;/sub&gt; · 16S rRNA gene</td>
<td>Total of CRAB: 1 Total of PRAB: 0 Total of TRAB: 1</td>
<td>bla&lt;sub&gt;NDM&lt;/sub&gt;</td>
</tr>
<tr>
<td>Cavalcanti et al., 2016 [26]</td>
<td>Recife, PE · Vitek&lt;sub&gt;2&lt;/sub&gt; · Identification of bla&lt;sub&gt;OXA-51&lt;/sub&gt; · 16S rRNA gene</td>
<td>Total of CRAB: - Total of PRAB: 45 Total of TRAB: -</td>
<td>bla&lt;sub&gt;OXA-25&lt;/sub&gt; (n=31)</td>
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<table>
<thead>
<tr>
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<th>Regions, period and design of study</th>
<th>Dates of isolates</th>
<th>Molecular mechanisms of resistance (n)</th>
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<tr>
<td>França et al., 2018 [29]</td>
<td>Belo Horizonte, MG · Prospective · 2016</td>
<td>Identification of $\text{bla}_{OXA-51}$</td>
<td>$\text{bla}<em>{OXA-23}$ (n=61) · $\text{bla}</em>{VIM-1}$ (n=52) · Absence of $\text{bla}<em>{OXA-58}$, $\text{bla}</em>{OXA-143}$ and $\text{bla}_{OXA-24/40}$</td>
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<tr>
<td>de Azevedo et al., 2019 [35]</td>
<td>Cuiabá, MS · Prospective · 2011-2015</td>
<td>· Bact/Alert 3D · Vitek2 · Identification of $\text{bla}_{OXA-51}$</td>
<td>80 · 1 · 26 · $\text{bla}<em>{OXA-23}$ (n=68) · ISAba1/$\text{bla}</em>{OXA-51}$ (n=48) · $\text{bla}<em>{OXA-58}$ (n=5) · $\text{bla}</em>{OXA-143}$ (n=25) · $\text{bla}<em>{OXA-24/40}$ (n=48) · $\text{bla}</em>{KPC}$ (n=1) · $\text{bla}_{NDM}$ (n=3)</td>
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<tr>
<td>Kobs et al., 2016 [23]</td>
<td>Joinville, SC · Prospective · 2009</td>
<td>· Microscan Walkaway · Identification of $\text{bla}_{OXA-51}$</td>
<td>69 · 0 · - · $\text{bla}<em>{OXA-23}$ (n=63) · Absence of $\text{bla}</em>{OXA-58}$, $\text{bla}<em>{OXA-143}$ and $\text{bla}</em>{OXA-24/40}$</td>
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<tr>
<td>Gomes et al., 2016 [24]</td>
<td>Rio de Janeiro, RJ · Case report · 2014</td>
<td>· Vitek2 · Identification of $\text{bla}_{OXA-51}$ · Identification of $\text{rpoB}$</td>
<td>1 · - · - · $\text{bla}_{OXA-72}$</td>
</tr>
<tr>
<td>Ribeiro et al., 2016 [25]</td>
<td>São Luis, MA · Prospective · 2012-2013</td>
<td>· Vitek2 · Identification of $\text{bla}_{OXA-51}$</td>
<td>128 · 0 · 0 · $\text{bla}_{KPC}$ (n=21)</td>
</tr>
<tr>
<td>Vasconcelos et al., 2015 [38]</td>
<td>Brazil (PA, RN, SP, DF and RS) · Retrospective · 2014</td>
<td>· MALDI-TOF MS · Amplificação do gene $\text{rpoB}$</td>
<td>43 · 3 · 0 · $\text{bla}<em>{OXA-23}$ (n=32) · $\text{bla}</em>{OXA-72}$ (n=10) · Absence of $\text{bla}_{OXA-143}$</td>
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<th>Reference</th>
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<th>Dates of isolates</th>
<th>Method to identification of isolates</th>
<th>Total of CRAB</th>
<th>Total of PRAB</th>
<th>Total of TRAB</th>
<th>Molecular mechanisms of resistance (n)</th>
</tr>
</thead>
</table>
| Cavalcanti et al., 2016 [26] | · Recife, PE  
· Case report  
· 2010-2011 |                  | · API ID 32 E  
· Identification of bla\text{\textsubscript{OXA-51}} | 2             | 0            | -            | · bla\text{\textsubscript{OXA-72}} (n=2)  
· Absence of bla\text{\textsubscript{IMP}}, bla\text{\textsubscript{VIM}}, bla\text{\textsubscript{SPM}}, bla\text{\textsubscript{OXA-58}}, and bla\text{\textsubscript{OXA-48}} |
| Dias et al., 2017 [34] | · Juiz de Fora, MG  
· Cross-sectional  
· 2013 |                  | · Vitek2  
· Identification of bla\text{\textsubscript{OXA-51}} | 44            | 0            | 0            | · bla\text{\textsubscript{OXA-23}} (n=32)  
· Absence of bla\text{\textsubscript{KPC}}, bla\text{\textsubscript{IMP}}, bla\text{\textsubscript{VIM}}, bla\text{\textsubscript{GIM}}, bla\text{\textsubscript{SIM}}, bla\text{\textsubscript{NDM}}, bla\text{\textsubscript{OXA-48}}, and bla\text{\textsubscript{OXA-24/40}} |
| Cieslinski et al., 2013 [33] | · Paraná, PR  
· Retrospective  
· 2009-2011 |                  | · Vitek2  
· Identification of bla\text{\textsubscript{OXA-51}}  
· Identification of gyrB | 46            | -            | -            | · bla\text{\textsubscript{OXA-23}} (n=46) |

- : No identified or reported; IMP: Imipenem; MEP: Meropenem; POL: Polymyxin B; MALDI-TOF MS: MALDI-TOF mass spectrometry; OXA: Oxacillinase; KPC: Klebsiella pneumoniae carbapenemase; NDM: New-delhi metallo-beta-lactamase.

1 Motility, growth at 42 °C, citrate utilization, oxidase and urease production, oxidative/fermentation, (OF)-glucose test, bile esculin hydrolysis test, decarboxylation of amino acids (i.e., lysine, ornithine and arginine) and OF-lactose test at 10%.

2 Oxidation/fermentation test activity, motility, oxidase production, catalase, glucose fermentation, urease activity and hemolysis of sheep blood.

3 Oxidase-negative, catalase-positive, glucose oxidation, ability to grow at 42 ° and 44 °C.
tibility [45], the combination of VITEK-2® with genetic techniques increased significantly the performance of this identification method.

According to this study, the main determinant of resistance in CRAB isolated over the last decade in Brazil was the presence of OXA-type carbapenemases. The frequency calculated by the meta-analysis was 98 % [95 % CI: 0.91; 0.99; I² = 95 %] (figure 2). During the meta-analysis, a high level of heterogeneity between the included studies, as shown by the I-squared (I²) index (I² = 95 %), was observed, and thus all data were analyzed following the random effect model.

![Figure 2. Meta-analysis of the oxacillinase-like resistance mechanism.](image)

In fact, several other reviews point out that class D carbapenemases (OXAs) are the principal mechanism of resistance to carbapenem among isolates of *A. baumannii* [42, 46, 47]. Although all *A. baumannii* possess the *bla*OXA-51-like* gene [48, 49], it is only expressed at levels compatible with carbapenem resistance after insertion sequence acquisition, specifically the ISAba1 element [45, 50]. In this review, 62 % of isolates
Mechanisms in resistance to carbapenems among *A. baumannii* isolates

were reported to carry this element upstream of the *bla*<sub>OXA-51-like</sub> gene, revealing its importance among *A. baumannii* in Brazil.

Insertion sequences, also described as insertion sequence elements, are considered to be the smallest mobile DNA elements (not exceeding 2500 bp), and are very rapidly transferred between several microorganisms [51].

In addition, a subgroup analysis was performed to assess the most frequent OXA variant. As shown in table 2, the OXA-23 was the most frequent carbapenemase in CRABs from Brazil, showing a frequency of 91% among the isolates included in this review [95% CI: 0.76; 0.97; I²= 97 %]. A similar rate was observed in South Africa, where >90% of *A. baumannii* isolates from HAIs were shown to produce OXA-23, while only 4% were found to have other types of carbapenemases (such as OXA-58) [52].

The *bla*<sub>OXA-23</sub> gene is considered a virulence biomarker, and its presence in *A. baumannii* strains has been observed since 1985, when the first report was made in Scotland [46]. Since then, this gene has been spread to many hospitals worldwide, making it the most frequent carbapenemase found in *A. baumannii* [43]. In Brazil, the first report of the spread of OXA-23 appears to have started in Curitiba (Parana state) in 1999, when the first outbreak associated with CRAB-related infections occurred in Brazil [18]. Due to its importance, the Agência Nacional de Vigilância Sanitária [53] considers OXA-23 to be the main mechanism responsible for resistance to carbapenems in *A. baumannii* in country, which is in accordance with this meta-analysis.

The presence of metallo-beta-lactamase (MBL) in CRAB isolates recovered in Brazil between 2009 and 2019 was also analyzed. Here, 467 isolates were included, with 56 of them presenting some MBL type. The meta-analysis revealed that, for this carbapenemase, a 12% frequency was found [95% CI: 0.09; 0.15; I²=99 %] (figure 3), evidencing that MBLs are not frequently reported in *A. baumannii* isolates [54], these enzymes have recently started appearing in sporadic cases, in several parts of the world.

Currently, some MBLs have been reported in *A. baumannii*, such as imipenemase (IMP), German imipenemase (GIM), Verona imipenemase (VIM), Seoul imipenemase (SIM), and São Paulo metallo-β-lactamase (SPM) [55, 56]. In the analysis of carbapenemase subgroups (table 2), the VIM type (Verona Imipenemase) was the most common in Brazil, and a 15% frequency was found [95% CI: 0.12; 0.19; I²= 99 %] which is in concordance with isolates from patients admitted in ICUs in Iran, India, Saudi Arabia, and Korea [54, 56, 57].
Table 2. Frequency of different types of carbapenemases subgroup identified between carbapenem-resistant *Acinetobacter baumannii* recovered in Brazil (2009-2019).

<table>
<thead>
<tr>
<th>Subgroup</th>
<th>events/n</th>
<th>Proportion (%)</th>
<th>95% IC</th>
<th>Heterogeneity (I²)</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>OXA type</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>OXA-23</td>
<td>754/887</td>
<td>91</td>
<td>0.76; 0.97</td>
<td>High (96)</td>
<td>Castilho et al., 2017 [37]; Cieslinski et al., 2013 [34]; Cortivo et al., 2015 [32]; da Silva et al., 2018 [30]; de Azevedo et al., 2019 [36]; Dias et al., 2017 [35]; França et al., 2018 [29]; Kobs et al., 2016 [23]; Moreira et al., 2018 [13]; Neves et al., 2016 [27]; Pagano et al., 2016 [11]; Pagano et al., 2018 [12]; Rocha et al., 2017 [40]; Tavares et al., 2018 [28]; Vasconcelos et al., 2015 [39]</td>
</tr>
<tr>
<td>OXA-51/ISAba1</td>
<td>106/170</td>
<td>84</td>
<td>0.15; 0.99</td>
<td>High (98)</td>
<td>Castilho et al., 2017 [37]; da Silva et al., 2018 [30]; de Azevedo et al., 2019 [36]</td>
</tr>
<tr>
<td>OXA-72</td>
<td>47/80</td>
<td>58</td>
<td>0.48; 0.69</td>
<td>High (93)</td>
<td>Cavalcanti et al., 2013 [26]; Fonseca et al., 2019 [41]; Gomes et al., 2016 [24]; Pagano 2016 [11]; Vasconcelos et al., 2015 [39]</td>
</tr>
<tr>
<td>OXA-24/40</td>
<td>60/749</td>
<td>8</td>
<td>0.00; 0.14</td>
<td>High (98)</td>
<td>Castilho et al., 2017 [37]; Cortivo et al., 2015 [32]; da Silva et al., 2018 [30]; de Azevedo et al., 2019 [36]; Dias et al., 2017 [35]; França et al., 2018 [29]; Kobs et al., 2016 [23]; Moreira et al., 2018 [13]; Neves et al., 2016 [27]; Pagano et al., 2016 [11]; Rocha et al., 2017 [40]; Tavares et al., 2018 [28]</td>
</tr>
<tr>
<td>OXA-143</td>
<td>34/561</td>
<td>6</td>
<td>0.00; 0.14</td>
<td>High (98)</td>
<td>Cortivo et al., 2015 [32]; da Silva et al., 2018 [30]; de Azevedo et al., 2019 [36]; França et al., 2018 [29]; Kobs et al., 2016 [23]; Neves et al., 2016 [27]; Pagano et al., 2016 [11]; Rocha et al., 2017 [40]; Vasconcelos et al., 2015 [39]</td>
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</table>

(Continued)
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<thead>
<tr>
<th>Subgroup</th>
<th>events/n</th>
<th>Proportion (%)</th>
<th>95% IC</th>
<th>Heterogeneity (I²)</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>OXA-58</td>
<td>8/751</td>
<td>1</td>
<td>0.00; 0.07</td>
<td>High (88)</td>
<td>Castilho et al., 2017 [37]; Cavalcanti et al., 2013 [26]; Cortivo et al., 2015 [32]; da Silva et al., 2018 [30]; de Azevedo et al., 2019 [36]; Dias et al., 2017 [35]; França et al., 2018 [29]; Kobs et al., 2016 [23]; Moreira et al., 2018 [13]; Neves et al., 2016 [27]; Pagano et al., 2016 [11]; Rocha et al., 2017 [40]; Tavares et al., 2018 [28]</td>
</tr>
<tr>
<td>OXA-48</td>
<td>0/150</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Cavalcanti et al., 2013 [26]; da Silva et al., 2018 [30]; Tavares et al., 2018 [28]</td>
</tr>
<tr>
<td>OXA-253</td>
<td>31/45</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Cavalcanti et al., 2016 [38]</td>
</tr>
<tr>
<td>MBL type</td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NDM</td>
<td>4/273</td>
<td>1</td>
<td>0.07; 0.12</td>
<td>High (99)</td>
<td>da Silva et al., 2018 [30]; de Azevedo et al., 2019 [36]; Dias et al., 2017 [35]; Pillonetto et al., 2014 [33]; Tavares et al., 2018 [28]</td>
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<td>VIM</td>
<td>52/342</td>
<td>15</td>
<td>0.12; 0.19</td>
<td>High (99)</td>
<td>Cavalcanti et al., 2013 [26]; da Silva et al., 2018 [30]; Dias et al., 2017 [35]; França et al., 2018 [29]; Moreira et al., 2018 [13]; Neves et al., 2016 [27]; Tavares et al., 2018 [28]</td>
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<td>IMP</td>
<td>0/281</td>
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<td>-</td>
<td>-</td>
<td>Cavalcanti et al., 2013 [26]; da Silva et al., 2018 [30]; Dias et al., 2017 [35]; Moreira et al., 2018 [13]; Neves et al., 2016 [27]; Tavares et al., 2018 [28]</td>
</tr>
<tr>
<td>SIM</td>
<td>0/75</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Dias et al., 2017 [35]; Moreira et al., 2018 [13]</td>
</tr>
<tr>
<td>SPM</td>
<td>0/140</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Cavalcanti et al., 2013 [26]; Moreira et al., 2018 [13]; Tavares et al., 2018 [28]</td>
</tr>
<tr>
<td>GIM</td>
<td>0/75</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Dias et al., 2017 [35]; Moreira et al., 2018 [13]</td>
</tr>
</tbody>
</table>

*: As only one study was dedicated to assessing the presence of this carbapenemases, subgroup analysis cannot be performed.
However, the presence of MBL in *A. baumannii* was lower in Brazil compared to Asian countries. For instance, a study from Iran which included 100 isolates of *A. baumannii*, collected from different clinical specimens of inpatients admitted to the largest teaching hospital in the north-west region of the country, showed that among 63 carbapenem (imipenem and meropenem) non-susceptible isolates, 31 (49 %) were found to be MBL producers. Of 31 MBL-producing isolates, 19 (61 %) carried the *bla* *IMP* gene and 9 (29%) carried the *bla* *VIM* gene [54].

The presence of New Delhi metallo-beta-lactamase (NDM) was also observed in only 1 % [95 % CI: 0.01; 0.04; $\Gamma^2=99\%$] of isolates, corroborating other studies from the United Arab Emirates and China, which also reported a low prevalence of this enzyme between *A. baumannii* isolates (1.2 % and 0.2 %, respectively) [56, 57]. However, Adler et al. (2018) [58] showed that, in an Israeli hospital, the prevalence of CRAB with the presence of the NDM enzyme was slightly higher (5.1 %), highlighting the regional differences in mechanisms of resistance to carbapenems circulating in *A. baumannii*.

Finally, for *Klebsiella pneumoniae* carbapenemase (KPC), 400 isolates were included, in which 22 presented this type of resistance mechanism. The frequency calculated by the meta-analysis was 6 %, as reported by other authors [59, 60] [95 % CI: 0.04; 0.08; $\Gamma^2=87\%$] (figure 4), and the random effect model was used. Subgroup analyses to variants of KPC were not performed, due to no studies being dedicated to typing this enzyme.
Mechanisms in resistance to carbapenems among *A. baumannii* isolates

Although the estimated prevalence remains lower, since 2009 when it was first reported in Puerto Rico [44], this type of carbapenemase has increased in prevalence in recent years, with rates increasing from 3.4% to 14% in beta-lactam–resistant isolates [59]. In addition, this carbapenemase has already been associated with resistance to colistin and tigecycline for *A. baumannii* isolated in Portugal [60].

In the analysis of resistance to polymyxins, 955 isolates were included, with only 17 among them presenting resistance. The calculated frequency was 1% [95% CI: 0.00; 0.0, 3; I²= 76 %], thus being antibiotic class with the highest activity against CRAB (figure 5). This observation is in accordance with reports from other countries, in which a low rate of resistance for colistin (2%) and polymyxin B (3%) was reported in different studies carried out in Iran [14, 61, 62]. These antibiotics are generally reliable agents against most *A. baumannii* isolates; however, resistance of *A. baumannii* to this class is emerging.

The use of colistin monotherapy has been associated with suboptimal clinical and microbiological outcomes. Furthermore, the *A. baumannii* hetero-resistance phenotype is currently reported in much higher frequency than the resistance rate to carbapenems in this bacterium, and the emergence of a determinant of colistin resistance carried by plasmids (*i.e.*, the *mcr* gene) has already been described [17, 63, 64].

To analyze resistance to tigecycline between CRAB, 821 isolates were included. Among them, 147 presented intermediate or complete resistance to tigecycline, with a calculated frequency of 7% [95% CI: 0.01; 0.28; I²=98 %], and the random effect model was also used (figure 6). In Middle Eastern [66-68], south and southwest Asia [65-71] and Europe [72, 73], resistance to tigecycline can vary from 2–82%, considering that Germany (6%) [73] and Saudi Arabia (6.6%) [65] have rates similar to those found in Brazil. In the Americas, non-susceptibility to tigecycline was found to be 5
% in North America [74], and between 0 – 20% in South America [75]. Thus, rates of non-susceptibility to tigecycline were lower in this review compared to other studies reported in Latin America.

<table>
<thead>
<tr>
<th>Study</th>
<th>Events</th>
<th>Total</th>
<th>Proportion</th>
<th>95%-CI</th>
</tr>
</thead>
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<td>da Silva 2018</td>
<td>0</td>
<td>41</td>
<td>0.00</td>
<td>[0.00; 0.09]</td>
</tr>
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<td>Castilho 2017</td>
<td>6</td>
<td>49</td>
<td>0.12</td>
<td>[0.05; 0.25]</td>
</tr>
<tr>
<td>Cortivo 2015</td>
<td>0</td>
<td>118</td>
<td>0.00</td>
<td>[0.00; 0.03]</td>
</tr>
<tr>
<td>Rocha 2017</td>
<td>3</td>
<td>91</td>
<td>0.03</td>
<td>[0.01; 0.09]</td>
</tr>
<tr>
<td>Neves 2016</td>
<td>3</td>
<td>56</td>
<td>0.05</td>
<td>[0.01; 0.15]</td>
</tr>
<tr>
<td>Moreira 2018</td>
<td>0</td>
<td>31</td>
<td>0.00</td>
<td>[0.00; 0.11]</td>
</tr>
<tr>
<td>Tavares 2018</td>
<td>0</td>
<td>107</td>
<td>0.00</td>
<td>[0.00; 0.03]</td>
</tr>
<tr>
<td>Pagano 2016</td>
<td>0</td>
<td>2</td>
<td>0.00</td>
<td>[0.00; 0.84]</td>
</tr>
<tr>
<td>Fonseca 2019</td>
<td>0</td>
<td>32</td>
<td>0.00</td>
<td>[0.00; 0.11]</td>
</tr>
<tr>
<td>Pillonetto 2014</td>
<td>0</td>
<td>1</td>
<td>0.00</td>
<td>[0.00; 0.98]</td>
</tr>
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<td>França 2018</td>
<td>1</td>
<td>61</td>
<td>0.02</td>
<td>[0.00; 0.09]</td>
</tr>
<tr>
<td>de Azevedo 2019</td>
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<td>80</td>
<td>0.01</td>
<td>[0.00; 0.07]</td>
</tr>
<tr>
<td>Kobs 2016</td>
<td>0</td>
<td>69</td>
<td>0.00</td>
<td>[0.00; 0.05]</td>
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<tr>
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<td>0.00</td>
<td>[0.00; 0.03]</td>
</tr>
<tr>
<td>Vasconcelos 2015</td>
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<td>43</td>
<td>0.07</td>
<td>[0.01; 0.19]</td>
</tr>
<tr>
<td>Cavalcanti 2013</td>
<td>0</td>
<td>2</td>
<td>0.00</td>
<td>[0.00; 0.84]</td>
</tr>
<tr>
<td>Dias 2017</td>
<td>0</td>
<td>44</td>
<td>0.00</td>
<td>[0.00; 0.08]</td>
</tr>
</tbody>
</table>

Random effects model 955 0.01 [0.00; 0.03]

Figure 5. Meta-analysis of resistance to polymyxins between CRAB isolates.

Furthermore, four studies (4/21; 19.0%) demonstrated, concomitantly, isolates with resistance to carbapenems (CRAB), tigecycline (TRAB) and polymyxin (PRAB) [27, 29, 35, 36], characterizing a pan-drug resistance phenotype (PDR).

This review faces some limitations that should be considered when interpreting data. Data regarding characteristics of patients, antimicrobial use, and profile of health units are limited in the included articles, which the roles of these factors on the susceptibility profile of isolates. Also, the heterogeneity of the included articles was high. May be justified, samples from different origins were included in the studies (e.g., blood culture, sputum, urine, or wounds). Lastly, as data in most regions in the country were not available, our findings do not completely represent the profile of resistance of CRAB isolates from Brazil. In addition, due to the lack of identification of some carbapenemases in studies, relevant data may have been excluded.
Mechanisms in resistance to carbapenems among *A. baumannii* isolates

**Conclusions**

In summary, we revealed that resistance to tigecycline in Brazil is lower, compared to in other countries in Latin America and in Asian countries. Regarding polymyxins, susceptibility remains high, and does not appear to be impacted by resistance to carbapenem in these isolates. In addition, oxacillinase is the main mechanism of resistance found in CRAB isolates in Brazil, with a major detachment for the presence of OXA-23, which is also considered an important virulence biomarker in this pathogen. Therefore, in order to prevent further dissemination of resistant isolates, appropriate diagnostic methods and infection control measures must be implemented. In addition, to prevent treatment failure, regular monitoring of antibiotic resistance by standard guidelines and methods is necessary, particularly for last-line antibiotics such as tigecycline and colistin.

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Disclosure statement

All authors report that they do not have any conflicts of interest.

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Mechanisms in resistance to carbapenems among A. baumannii isolates


determined by the broth microdilution and disk diffusion methods, Int. J. Anti-


How to cite this article

Details of the search strategy

1. Search in Pubmed

#1 “Acinetobacter [MeSH]” OR “Acinetobacter” OR “acinetobacter” OR “Acinetobacterbaumannii [MeSH]” OR “Acinetobacterbaumannii” OR “baumannii, Acinetobacter” OR “acinetobacterbaumannii” OR “Acinetobacter Infections [MeSH]” OR “Acinetobacter Infections” OR “Infections, Acinetobacter” in Title/Abstract

#2 “Brazil [MeSH]” OR “Brazil” in All fields

Combination to search: #1 AND #2

2. Search in Web of Science

#1 TI= “Acinetobacter” OR “Acinetobacterbaumannii” OR “Acinetobacter Infection*”

#2 TS=Brazil

Combination to search: #1 AND #2

3. Search in Scopus

TITLE-ABS-KEY “Acinetobacter” OR “Acinetobacterbaumannii” OR “Acinetobacter Infection*” OR INDEXTERMS “Acinetobacter” OR “Acinetobacterbaumannii” OR “Acinetobacter Infection*” AND (TITLE-ABS-KEY “Brazil” OR INDEXTERMS (“Brazil”))

4. Search in Cochrane library

“Acinetobacterbaumannii” AND (Brazil) in Title Abstract Keyword

5. Search in Biblioteca Virtual em Saúde

“Acinetobacterbaumannii” OR “Acinetobacter” AND (Brazil) in Title Abstract Keyword

6. Search in SciELO

“Acinetobacterbaumannii” AND (Brazil) in Todososcampos

7. Search in LILACS

“Acinetobacterbaumannii” AND (Brazil) in Todososcampos