

**Design, synthesis, *in silico* studies, and antidiabetic activity of several sulfanilamide incorporated 2,3-disubstituted thiazolidin-4-ones****Diseño, síntesis, estudios *in silico* y actividad antidiabética de algunas sulfanilamidas 2,3-disustituidas tiazolidina-4-onas incorporadas****Concepção, síntese, em estudos de *silico* e actividade antidiabética de poucas sulfanilamidas incorporado 2,3-disubstituído tiazolidina-4-ones****Abstract**

A panel of 2,3-disubstituted thiazolidin-4-ones **4a-n** was synthesised from Schiff bases **3a-n** derived from sulfanilamide, by reaction with thioglycolic acid. The compounds were characterised by means of IR, NMR, and Mass spectral data. Compounds **4a-n** were screened for DPPH scavenging assay and compounds **4e, 4h, 4i**, and **4n** exhibited moderate activity. Compounds **4e, 4h**, and **4i** were tested at 200 mg/kg and **4e** at 50 mg/kg b.w. orally for antidiabetic activity in fructose induced diabetic rats. They exhibited significant antidiabetic activity compared to the control group. Pioglitazone was used as a standard drug. The tested compounds exhibited better and significant serum cholesterol lowering activity when compared with the control and standard groups. They also reduced the triglyceride level after the 21st day; however, it was insignificant when compared to the control group. Compound **4n** displayed the highest binding energy when docked with PPAR- γ followed by compounds **4e, 4h**, and **4i** when compared to pioglitazone. The physicochemical, drug likeness and ADME properties of the title compounds were found to be satisfactory.

Keywords: Schiff bases; DPPH; antioxidant; serum glucose; total serum cholesterol; serum triglycerides; PPAR- γ ; ADME; Drug Likeness.

Resumen

Se sintetizó un panel de tiazolidinas-4-onas 2,3-disustituidas **4a-n** a partir de las bases de Schiff **3a-n** derivadas de la sulfanilamida por reacción con ácido tioglicólico. Los compuestos se caracterizaron por IR, RMN y datos espectrales de masa. Los compuestos **4a-n** se analizaron para DPPH y los compuestos **4e, 4h, 4i** y **4n** mostraron una actividad moderada. Los compuestos **4e, 4h** y **4i** se probaron a 200 mg/kg y **4e** a 50 mg/kg b.w. oralmente para la actividad antidiabética en ratas diabéticas, inducida por fructosa. Los compuestos mostraron una actividad antidiabética muy significativa en comparación con el grupo control. La pioglitazona se utilizó como fármaco estándar. Los compuestos ensayados mostraron una mejor y significativa actividad reductora del colesterol sérico en comparación con los grupos control y estándar. Estos compuestos también redujeron el nivel de triglicéridos después del 21^o día, aunque fue insignificante en comparación con el grupo control. El compuesto **4n** mostró la mayor afinidad de unión cuando se acopló a PPAR- γ , seguido de **4e, 4h** y **4i** en comparación con la pioglitazona. Las propiedades fisicoquímicas, la similitud con el fármaco y las propiedades ADME de los compuestos fueron satisfactorias, lo que los convierte en útiles agentes antidiabéticos.

Palabras clave: Bases de Schiff; DPPH; antioxidante; glucosa sérica; colesterol total sérico; triglicéridos séricos triglicéridos séricos; PPAR- γ ; ADME; similitud de fármacos.

Resumo

Um painel de 2,3-disubstituído tiazolidina-4-ones **4a-n** foram sintetizados a partir de bases Schiff **3a-n** derivado da sulfanilamida por reação com ácido tioglicólico. Os compostos eram caracterizado por IR, NMR e dados espectrais de massa. Os compostos **4a-n** foram rastreados para O ensaio DPPH de limpeza radical e os compostos **4e, 4h, 4i** e **4n** exibiram actividade moderada. Os compostos **4e, 4h** e **4i** foram testados a 200 mg/kg e **4e** a 50 mg/kg de peso corporal por via oral para antidiabéticos. actividade em ratos diabéticos induzidos por fructose. Exibiram uma actividade antidiabética altamente significativa actividade em comparação com o controlo. A pioglitazona foi utilizada como droga padrão. Os compostos testados exibiu uma melhor e significativa actividade de redução do colesterol sérico quando comparado com de triglicéridos após o 21^o dia; no entanto, foi insignificante quando comparado com o controlo. O composto **4n** mostrou a maior afinidade de ligação quando acoplado com PPAR- γ seguido de **4e, 4h, 4i** quando comparado com pioglitazona. O propriedades físico-químicas, de semelhança com drogas e ADME dos compostos do título de propriedade também foram encontrados paraser satisfatórios, tornando-os agentes antidiabéticos úteis.

Palavras-chave : bases Schiff; DPPH; antioxidante; glucose sérica; colesterol total sérico; soro triglicéridos; PPAR- γ ; ADME; Probabilidade de drogas.



Introduction

Diabetes mellitus is a spectrum of metabolic disorders resulting from myriad pathogenic mechanisms, all of which lead to hyperglycemia [1]. In recent years, diabetes has become one of the leading causes of death worldwide. In 2016, the World Health Organisation reported around 1.6 million deaths globally due to diabetes. The number of diabetes cases worldwide in 2019 was 463 million and by 2045 it could increase to 629 million. In India, 77 million suffer from diabetes and could be further increased and India has the second highest number of cases in the world [2]. Oxidative stress plays an important role in the pathogenesis of typical long-term diabetic complications such as neuropathy and microangiopathy. Further, dynamic modification of intracellular redox sensors by way of reactive oxygen species in addition to other post-translational modifications such as protein phosphorylation, acetylation or ubiquitination also play a role in the development of the diabetes [3]. Thiazolidinone is one of the important and fundamental central structures for the synthesis of biologically active compounds [4]. Thiazolidin-4-one is a well-known scaffold for producing the desired biological activities: Antidiabetic [5], antioxidant [6], anticancer [7], antitubercular [8], antimicrobial [9], anticonvulsant [10], analgesic and anti-inflammatory [11] and antihyperlipidemic [12].

The abovementioned facts triggered an interest in taking up this investigation, wherein molecular architecture having a 2,3-disubstituted 4-thiazolidinone core derived from sulfanilamide was designed with an anticipation of good antidiabetic activity. Spectral characterisation, docking studies with peroxisome proliferative activated receptor- γ (PPAR- γ), and *in silico* ADME parameter studies were also undertaken.

Materials and Methods

Chemicals employed were of laboratory reagent grade and used as received. Sulphanilamide, 1,1-diphenyl-2-picryl-hydrazyl (DPPH), and *p*-hydroxybenzaldehyde were purchased from Loba Chemie Pvt Limited, Mumbai, India. The other substituted aldehydes, 1,4-dioxane, thioglycolic acid, anhydrous zinc chloride, ascorbic acid, and D-fructose were procured from SD Fine Chemicals, Mumbai, India. *p*-Tolualdehyde was purchased from Sigma Aldrich, Steinheim, USA. *p*-Fluorobenzaldehyde was obtained from Spectrochem, Mumbai, India; and 2,4-dichlorobenzaldehyde, 2,6-dichlorobenzaldehyde, and *p*-bromobenzaldehyde from HiMedia Laboratories Pvt. Ltd. Mumbai, India. The glucose, cholesterol, and triglyceride kits were purchased from Erba Mannheim, Sikkim, India. Pioglitazone hydrochloride was provided by USV Private Limited, Solan, India. Thin layer chromatography using pre-coated aluminium sheets with silica gel 60 F₂₅₄ (Merck, Darmstadt, Germany) was performed using a combination of solvents in different ratios as mobile phase. Spots were visualised by iodine vapor and/or UV light. Yields refer to crude products.

Melting points were determined on an open capillary apparatus (Veego VMP-DS Mumbai, India) and are uncorrected. The compounds were characterised by ultraviolet (UV) spectra (UV 1601 Shimadzu, Kyoto, Japan) and infrared (IR) spectra as KBr pellets (Shimadzu and Thermo Scientific Nicolet iS5, Japan). Proton nuclear magnetic resonance (¹H NMR) was recorded by means of DMSO-*d*₆ (Bruker Avance Neo 500 MHz, Germany) while ¹³C NMR was recorded at 125 MHz. Mass spectra (MS) were registered by chemical ionisation technique (Waters-SynaptG2, Massachusetts, USA).

Animals

Sprague Dawley rats of either sex weighing between 250 and 350 g and Swiss albino female mice weighing from 25 to 40 g were obtained from the central animal house of the HSK College of Pharmacy, Bagalkote. The animals were housed under standard conditions (temperature 25 ± 1°C,

relative humidity (50-55%)) for 12 h dark and light cycle. They were given standard laboratory feed (Pranava Agro Industries Ltd., Sangli, India) and water *ad libitum* and acclimatised for 7 days before experimentation. Food was withdrawn 18 h before the experiment, but free access to water was allowed. The experiments were performed on randomly formed groups during the light phase of the cycle and the animals were used for one experiment only. The test compounds and standard drug were administered *p.o.* as aqueous suspension in 0.5% Tween-80. The animals in the vehicle control group received 0.5% aqueous Tween-80. Blood samples were collected from retro orbital flexus under ether anaesthesia. The study was approved and conducted under the guidelines of institutional animal ethics committee constituted as per committee for the control and supervision of experiments on animals, Ministry of Environment and Forestry, Government of India (F.No. HSK College of Pharmacy Bagalkot/IAEC// HSKCOP/AUG2021/PG13).

Synthesis of Schiff bases 3a-n

Sulfanilamide (40 mmol) 1 and substituted aldehydes (40 mmol) 2a-n were dissolved in 20 mL ethanol and the reaction mixture was refluxed for 4-5 h. Completion of reaction was monitored by TLC (n-hexane: ethyl acetate 1:2). The excess of ethanol was distilled, and the product 3a-n was settled after several hours and was filtered and washed with cold ethanol multiple times [13].

4-(benzylideneamino)benzenesulfonamide 3a

White crystals; Yield: 38%; MP: 192-194 °C; *R*_f: 0.77; IR (KBr, cm⁻¹): 3296.35 (NH str), 3012.81 (Ar-CH str), 1624 (C = N str), 1575 (Ar-C = C str), 1334 (SO₂ str).

4-((4-methylbenzylidene)amino)benzenesulfonamide 3b

White crystals; Yield: 43.93%; MP: 198-200 °C; *R*_f: 0.78; IR (KBr, cm⁻¹): 3277 (NH str), 2993 (Ar-CH str), 1622 (C = N str), 1581 (Ar-C = C str), 1330 (SO₂ str).

4-((4-methoxybenzylidene)amino)benzenesulfonamide 3c

White crystals; Yield: 46.11%; MP: 200 °C; *R*_f: 0.69; IR (KBr, cm⁻¹): 3277 (NH str), 2954 (Ar-CH str), 1610 (C = N str), 1577 (Ar-C = C str), 1322 (SO₂ str).

4-((2-hydroxybenzylidene)amino)benzenesulfonamide 3d

Yellowish orange crystals; Yield: 45.0%; MP: 218 °C; *R*_f: 0.76; IR (KBr, cm⁻¹): 3342 (OH str), 3230 (NH str), 3012.81 (Ar-CH str), 1618 (C = N str), 1562 (Ar-C = C str), 1301 (SO₂ str).

4-((4-hydroxybenzylidene)amino)benzenesulfonamide 3e

Light yellowish powder; Yield: 69.05%; MP: 212 °C; *R*_f: 0.67; IR (KBr, cm⁻¹): 3346 (OH str), 3266 (NH str), 3011.66 (Ar-CH str), 1620 (C = N str), 1576 (Ar-C = C str), 1326 (SO₂ str).

4-((4-(dimethylamino)benzylidene)amino)benzenesulfonamide 3f

Yellowish powder; Yield: 6.82%; MP: 220 °C; *R*_f: 0.83; IR (KBr, cm⁻¹): 3277.06 (NH str), 2900 (Ar-CH str), 1604.77 (C = N str), 1571 (Ar-C = C str), 1328 (SO₂ str).

4-((3,4-dimethoxybenzylidene)amino)benzenesulfonamide 3g

White powder; Yield: 57.98%; MP: 200-202 °C; *R*_f: 0.55; IR (KBr, cm⁻¹): 3277 (NH str), 2954 (Ar-CH str), 1608 (C = N str), 1568 (Ar-C = C str), 1330 (SO₂ str), 1278 (Ar-C-O-C str).

4-((4-chlorobenzylidene)amino)benzenesulfonamide 3h

White crystals; Yield: 86.16%; MP: 206 °C; *R*_f: 0.75; IR (KBr, cm⁻¹): 3320 (NH str), 3010 (Ar-CH str), 1615 (C = N str), 1566 (Ar-C = C str), 1296 (SO₂ str), 846 (C-Cl str).

4-((2,4-dichlorobenzylidene)amino)benzenesulfonamide 3i

Light yellowish crystals; Yield: 44.93%; MP: 210 °C; R_f : 0.80; IR (KBr, cm^{-1}): 3348 (NH str), 3050 (Ar-CH str), 1618 (C = N str), 1565 (Ar-C = C str), 1298 (SO_2 str), 831 (C-Cl str).

4-((2,6-dichlorobenzylidene)amino)benzenesulfonamide 3j

White powder; Yield: 34.55%; MP: 178 °C; R_f : 0.83; IR (KBr, cm^{-1}): 3347 (NH str), 3056 (Ar-CH str), 1619 (C = N str), 1568 (Ar-C = C str), 1299 (SO_2 str), 833 (C-Cl str).

4-((4-bromobenzylidene)amino)benzenesulfonamide 3k

White crystals; Yield: 98.3%; MP: 222 °C; R_f : 0.75; IR (KBr, cm^{-1}): 3290 (NH str), 2987 (Ar-CH str), 1622 (C = N str), 1577 (Ar-C = C str), 1330 (SO_2 str), 840 (C-Br str).

4-((4-nitrobenzylidene)amino)benzenesulfonamide 3l

Yellowish orange powder; Yield: 18.83%; MP: 178-180 °C; R_f : 0.85; IR (KBr, cm^{-1}): 3263 (NH str), 3091 (Ar-CH str), 1627 (C = N str), 1591 (Ar-C = C str), 1340 (SO_2 str), 1294 (C-NO₂).

4-((4-fluorobenzylidene)amino)benzenesulfonamide 3m

Yellowish white crystals; Yield: 55.58%; MP: 156 °C; R_f : 0.76; IR (KBr, cm^{-1}): 3286 (NH str), 2996 (Ar-CH str), 1622 (C = N str), 1582 (Ar-C = C str), 1310 (SO_2 str), 1130 (C-F).

4-((3-phenylallylidene)amino)benzenesulfonamide 3n

White crystals; Yield: 30.64%; MP: 220-222 °C; R_f : 0.56; IR (KBr, cm^{-1}): 3283.53 (NH str), 2992.18 (Ar-CH str), 1614 (C = N str), 1580 (Ar-C = C str), 1335 (SO_2 str).

Synthesis of 2,3-disubstituted thiazolidin-4-ones (4a-n)

The substituted Schiff bases **3a-n** (10 mmol), thioglycolic acid (20 mmol), and anhydrous zinc chloride (0.2 g) were added to 10 mL 1,4-dioxane and the mixture was refluxed for 20 h. The progress of the reaction was monitored by TLC (n-hexane: ethyl acetate 3:2). The reaction mixture was allowed to cool and poured into crushed ice. The crude solid of 2,3-disubstituted thiazolidin-4-ones **4a-n** obtained was filtered, washed thoroughly with sodium bicarbonate followed by ice cold brine solution. The compounds were recrystallised with aqueous alcohol (30:70) [14].

4-(4-oxo-2-phenylthiazolidin-3-yl)benzenesulfonamide 4a

Orange powder, Yield: 28.74%; MP: 122 °C; R_f : 0.33; IR (KBr, cm^{-1}): 3422.20 (NH str), 1630.63 (C = O str), 1384.70 (SO_2 str), 618 (C-S str), ¹H NMR (500 MHz, δ , ppm): 4.10 (q, 2H, 5-CH₂, Thiaz) 6.39 (s, 1H, 2-CH, Thiaz) 7.03 (s, 2H, SO_2NH_2) 7.10 (m, 1H, 4" ArH) 7.21 (m, 2H, 3" & 5" ArH) 7.35 (m, 2H, 2" & 6" ArH) 7.42 (d, 2H, 2' & 6' ArH) 7.76 (d, 2H, 3' & 5' ArH); ¹³C NMR (125 MHz, δ , ppm): 32.53 (1C, 5C, Thiaz) 62.64 (1C, 2C, Thiaz) 121.5 (2C, 2' & 6' ArC) 125.8 (2C, 2" and 6" ArC) 126.2 (1C, 4" ArC) 128.4 (2C, 3" and 5" ArC) 129.3 (2C, 3' & 5' ArC) 135.4 (1C, 4' ArC) 140.6 (1C, 1" ArC) 143.8 (1C, 1' ArC) 170.40 (1C, C = O); Mass (m/z): 335.01 (M+ +1), 336.01 (M++2).

4-(4-oxo-2-(p-tolyl)thiazolidin-3-yl)benzenesulfonamide 4b

Orange powder, Yield: 31%; MP: 182-184 °C; R_f : 0.37; IR (KBr, cm^{-1}): 3406.18 (NH str), 2850 (Ali CH str), 1650 (C = O str), 1384.71 (SO_2 str), 618.52 (C-S str), ¹H NMR (500 MHz, δ , ppm): 2.39 (s, 3H, CH₃) 4.4 (q, 2H, 5-CH₂, Thiaz) 5.7 (s, 1H, 2-CH, Thiaz) 6.86 (s, 2H, SO_2NH_2) 7.34 (m, 2H, 3" & 5" ArH) 7.38 (m, 2H, 2" & 6" ArH) 7.5 (m, 2H, 2' & 6' ArH) 7.8 (m, 2H, 3' & 5' ArH); ¹³C NMR (125 MHz, δ , ppm): 20.4 (1C, CH₃) 32.76 (1C, 5C, Thiaz) 65.68 (1C, 2C, Thiaz) 121.6 (2C, 2' & 6' ArC) 127.9 (2C, 2" and 6" ArC) 128.4 (2C, 3" and 5" ArC) 129.5 (2C, 3' & 5' ArC) 136.4 (1C, 4' ArC) 137.1 (1C, 1" ArC) 137.6 (1C, 4" ArC) 144.8 (1C, 1' ArC) 169.38 (1C, C = O); Mass (m/z): 349.07 (M+ +1).

4-(2-(4-methoxyphenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4c

Orange powder, Yield: 23.07%; MP: 220 °C; R_f : 0.77; IR (KBr, cm^{-1}): 3418.04 (NH str), 2852 (Ali CH str), 1655 (C = O str), 1384.86 (SO_2 str), 1333.29 (Ar-C-O-C str), 618.33 (C-S str), ¹H NMR (500 MHz, δ , ppm): 3.76 (s, 3H, OCH₃) 4.4 (q, 2H, 5-CH₂, Thiaz) 6.36 (s, 1H, 2-CH, Thiaz) 6.71 (m, 2H, SO_2NH_2) 6.72 (m, 2H, 3" & 5" ArH) 7.40 (d, 2H, 2' & 6' ArH) 7.72 (d, 2H, 3' & 5' ArH) 7.82 (d, 2H, 2" & 6" ArH); ¹³C NMR (125 MHz, δ , ppm): 33.76 (1C, 5C, Thiaz) 55.8 (1C, OCH₃) 70.53 (1C, 2C, Thiaz) 113.8 (2C, 3" and 5" ArC) 120.2 (2C, 2' & 6' ArC) 128.3 (2C, 3' & 5' ArC) 129.9 (2C, 2" and 6" ArC) 130.3 (1C, 1" ArC) 135.4 (1C, 4' ArC) 144.8 (1C, 1' ArC) 157.6 (1C, 4" ArC) 170.38 (1C, C = O); Mass (m/z): 365.02 (M+ +1), 366 (M+2).

4-(2-(2-hydroxyphenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4d

Orange powder, Yield: 47.14%; MP: 202-204 °C; R_f : 0.7; IR (KBr, cm^{-1}): 3345.60 (OH str), 3248.17 (NH str), 2923 (Ali CH str) 1618.84 (C = O str), 1384.77 (SO_2 str), 618.57 (C-S str), ¹H NMR (500 MHz, δ , ppm): 4.4 (q, 2H, 5-CH₂, Thiaz) 5.7 (s, 1H, 2-CH, Thiaz) 6.99 (m, 2H, 3" & 5" ArH) 7.0 (m, 1H, 4" ArH) 7.4 (m, 1H, 6" ArH) 7.39 (s, 2H, SO_2NH_2) 7.56 (d, 2H, 2' & 6' ArH) 7.8-7.9 (d, 2H, 3' & 5' ArH); ¹³C NMR (125 MHz, δ , ppm): 32.6 (1C, 5C, Thiaz) 65.3 (1C, 2C, Thiaz) 115.8 (1C, 5" ArC) 119.2 (1C, 1" ArC) 121.9 (1C, 3" ArC) 122.2 (2C, 2' & 6' ArC) 127.8 (1C, 2" ArC) 128.3 (1C, 4" ArC) 129.4 (2C, 3' & 5' ArC) 136.4 (1C, 4' ArC) 145.8 (1C, 1' ArC) 152.9 (1C, 6" ArC) 170.6 (1C, C = O); Mass (m/z): 351.05 (M+ +1).

4-(2-(4-hydroxyphenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4e

Brownish powder, Yield: 2.5%; MP: 230 °C; R_f : 0.28; IR (KBr, cm^{-1}): 3358.68 (OH str), 3253 (NH str), 1650 (C = O str), 1384.23 (SO_2 str), 618.34 (C-S str), ¹H NMR (500 MHz, δ , ppm): 3.98 (q, 2H, 5-CH₂, Thiaz) 6.46 (s, 1H, 2-CH, Thiaz) 6.69 (d, 2H, 3" & 5" ArH) 6.88 (s, 2H, SO_2NH_2) 7.46 (d, 2H, 2' & 6' ArH) 7.62 (d, 2H, 2" & 6" ArH) 7.73 (d, 2H, 3' & 5' ArH) 9.02 (s, 1H, OH); ¹³C NMR (125 MHz, δ , ppm): 31.5 (1C, 5C, Thiaz) 70.6 (1C, 2C, Thiaz) 114.8 (2C, 3" and 5" ArC) 121.2 (2C, 2' & 6' ArC) 129.3 (2C, 3' & 5' ArC) 131.1 (2C, 2" and 6" ArC) 131.8 (1C, 1" ArC) 136.3 (1C, 4' ArC) 143.8 (1C, 1' ArC) 156.6 (1C, 4" ArC) 168.38 (1C, C = O); Mass (m/z): 351.04 (M+ +1).

4-(2-(4-(dimethylamino)phenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4f

Dark brownish, Yield: 19.09%; MP: 158 °C; R_f : 0.57; IR (KBr, cm^{-1}): 3424.72 (NH str), 2870 (Ali CH str), 1606 (C = O str), 1384.46 (SO_2 str), 618.21 (C-S str), ¹H NMR (500 MHz, δ , ppm): 3.01 (s, 6H, N(CH₃)₂) 3.92 (q, 2H, 5-CH₂, Thiaz) 6.38 (s, 1H, 2-CH, Thiaz) 6.70 (d, 2H, 3" & 5" ArH) 6.82 (s, 2H, SO_2NH_2) 7.08 (d, 2H, 2" & 6" ArH) 7.40 (d, 2H, 2' & 6' ArH) 7.74 (d, 2H, 3' & 5' ArH); ¹³C NMR (125 MHz, δ , ppm): 32.6 (1C, 5C, Thiaz) 41.6 (2C, N(CH₃)₂) 70.8 (1C, 2C, Thiaz) 112.8 (2C, 3" and 5" ArC) 122.5 (2C, 2' & 6' ArC) 126.4 (2C, 2" and 6" ArC) 127.9 (1C, 1" ArC) 128.6 (2C, 3' & 5' ArC) 136.8 (1C, 4' ArC) 143.9 (1C, 1' ArC) 148.2 (1C, 4" ArC) 170.42 (1C, C = O); Mass (m/z): 378.01 (M+ +1).

4-(2-(3,4-dimethoxyphenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4g

Dark brownish, Yield: 1.26%; MP: 90 °C; R_f : 0.46; IR (KBr, cm^{-1}): 3370.80 (NH str), 2924.94 (Ali CH str), 1674.77 (C = O str), 1384.68 (SO_2 str), 1333.29 (Ar-C-O-C str), 618.80 (C-S str), ¹H NMR (500 MHz, δ , ppm): 3.70 (s, 3H, OCH₃) 3.81 (s, 3H, OCH₃) 4.02 (q, 2H, 5-CH₂, Thiaz) 6.46 (s, 1H, 2-CH, Thiaz) 6.82 (m, 1H, 3" ArH) 6.83 (m, 2H, SO_2NH_2) 7.45 (m, 2H, 2' & 6' ArH) 7.52 (m, 1H, 2" ArH) 7.63 (m, 1H, 6" ArH) 7.74 (d, 2H, 3' & 5' ArH); ¹³C NMR (125 MHz, δ , ppm): 34.46 (1C, 5C, Thiaz) 56.7 (2C, (OCH₃)₂) 72.33 (1C, 2C, Thiaz) 112.1 (1C, 3" ArC) 113.2 (1C, 6" ArC) 121.2 (2C, 2' & 6' ArC) 122.4 (1C, 2" ArC) 129.3 (2C, 3' & 5' ArC) 132.6 (1C, 1" ArC) 136.6 (1C, 4' ArC) 144.2 (1C, 1' ArC) 144.4 (1C, 4" ArC) 149.5 (1C, 5" ArC) 171.88 (1C, C = O); Mass (m/z): 395.07 (M+ +1).

4-(2-(4-chlorophenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4h

Orange powder, Yield: 47.55%; MP: 90 °C; R_f : 0.42; IR (KBr, cm^{-1}): 3405.55 (NH str), 1630 (C = O str), 1384.76 (SO_2 str), 618.03 (C-S str), 843.25 (C-Cl str), ¹H NMR (500 MHz, δ , ppm): 3.99 (q, 2H, 5-CH₂, Thiaz) 6.2 (s, 1H, 2-CH, Thiaz) 6.8 (m, 2H, SO_2NH_2) 7.34 (m, 2H, 2" & 6" ArH)

7.36 (d, 2H, 3" & 5" ArH) 7.63 (d, 2H, 2' & 6' ArH) 7.85 (d, 2H, 3' & 5' ArH); ¹³C NMR (125 MHz, δ , ppm): 31.33 (1C, 5C, Thiaz) 71.49 (1C, 2C, Thiaz) 121.22 (2C, 2' & 6' ArC) 128.28 (2C, 3" and 5" ArC) 128.43 (2C, 3' & 5' ArC) 130.9 (2C, 2" and 6" ArC) 132.1 (1C, 4" ArC) 136.3 (1C, 4' ArC) 136.2 (1C, 1" ArC) 144.3 (1C, 1' ArC) 168.26 (1C, C = O); Mass (m/z): 368.97 (M⁺+1), 370.97 (M⁺+3).

4-(2-(2,4-dichlorophenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4i

Yellow powder, Yield: 57.07%; MP: 110 °C; R_f: 0.54; IR (KBr, cm⁻¹): 3360.66 (NH str), 1680.92 (C = O str), 1384.52 (SO₂ str), 833.81 (C-Cl str), 618.10 (C-S str), ¹H NMR (500 MHz, δ , ppm): 4.0 (q, 2H, 5-CH₂, Thiaz) 5.94 (s, 1H, 2-CH, Thiaz) 6.86 (s, 2H, SO₂NH₂) 7.21 (m, 1H, 6" ArH) 7.27 (m, 1H, 5" ArH) 7.5 (m, 2H, 2' & 6' ArH) 7.74 (m, 1H, 3" ArH) 7.79 (d, 2H, 3' & 5' ArH); ¹³C NMR (125 MHz, δ , ppm): 32.69 (1C, 5C, Thiaz) 67.8 (1C, 2C, Thiaz) 100.4 (1C, 1" ArC) 120.6 (2C, 2' & 6' ArC) 126.4 (1C, 5" ArC) 128.4 (2C, 3' & 5' ArC) 130.2 (1C, 3" ArC) 131.9 (2C, 6" ArC) 134.6 (1C, 4" ArC) 135.9 (1C, 2" ArC) 136.4 (1C, 4' ArC) 143.7 (1C, 1' ArC) 171.38 (1C, C = O); Mass (m/z): 402.93 (M⁺+1), 404.92 (M⁺+3).

4-(2-(2,6-dichlorophenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4j

Pale yellow powder, Yield: 44.66%; MP: 138 °C; R_f: 0.43; IR (KBr, cm⁻¹): 3314.79 (NH str), 3081.21 (Ar-CH str), 1643.96 (C = O str), 1384.73 (SO₂ str), 833.56 (C-Cl str), 618.11 (C-S str), ¹H NMR (500 MHz, δ , ppm): 4.1 (q, 2H, 5-CH₂, Thiaz) 5.8 (s, 1H, 2-CH, Thiaz) 7.38 (m, 2H, SO₂NH₂) 7.4 (m, 2H, 2' & 6' ArH) 7.52 (m, 1H, 4" ArH) 7.61 (m, 2H 3" & 5" ArH) 7.9 (d, 2H, 3' & 5' ArH); ¹³C NMR (125 MHz, δ , ppm): 32.55 (1C, 5C, Thiaz) 62.43 (1C, 2C, Thiaz) 121.6 (2C, 2' & 6' ArC) 126.73 (2C, 3" and 5" ArC) 128.4 (1C, 4" ArC) 128.9 (2C, 3' & 5' ArC) 135.4 (2C, 2" and 6" ArC) 136.4 (1C, 4' ArC) 140.3 (1C, 1" ArC) 143.86 (1C, 1' ArC) 170.38 (1C, C = O); Mass (m/z): 402.93 (M⁺+1), 404.92 (M⁺+3).

4-(2-(4-bromophenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4k

Orange powder, Yield: 84.74%; MP: 212-214 °C; R_f: 0.44; IR (KBr, cm⁻¹): 3424.85 (NH str), 1640 (C = O str), 1384.77 (SO₂ str), 752.53 (C-Br str), 618.35 (C-S str), ¹H NMR (500 MHz, δ , ppm): 4.02 (q, 2H, 5-CH₂, Thiaz) 6.47 (s, 1H, 2-CH, Thiaz) 6.93 (s, 2H, SO₂NH₂) 7.21 (d, 2H, 2" & 6" ArH) 7.53 (d, 2H, 2' & 6' ArH) 7.79 (d, 2H, 3' & 5' ArH) 7.86 (d, 2H, 3" & 5" ArH); ¹³C NMR (125 MHz, δ , ppm): 32.41 (1C, 5C, Thiaz) 71.52 (1C, 2C, Thiaz) 121.6 (1C, 4" ArC) 121.4 (2C, 2' & 6' ArC) 129.3 (2C, 3' & 5' ArC) 130.9 (2C, 2" and 6" ArC) 131.8 (2C, 3" and 5" ArC) 136.4 (1C, 4' ArC) 138.3 (1C, 1" ArC) 144.6 (1C, 1' ArC) 171.37 (1C, C = O); Mass (m/z): 412.96 (M⁺+1).

4-(2-(4-nitrophenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4l

Pale brownish powder, Yield: 98.59%; MP: 96 °C; R_f: 0.77; IR (KBr, cm⁻¹): 3380.52 (NH str), 1650 (C = O str), 1596.22 (NO₂ Asy str), 1384.77 (SO₂ str), 1346.91 (NO₂ sym str), 617.93 (C-S str), ¹H NMR (500 MHz, δ , ppm): 3.9 (q, 2H, 5-CH₂, Thiaz) 6.81 (s, 1H, 2-CH, Thiaz) 7.32 (s, 2H, SO₂NH₂) 7.59 (m, 2H, 2' & 6' ArH) 7.72 (m, 2H, 2" & 6" ArH) 7.76 (m, 2H, 3' & 5' ArH) 8.16 (m, 2H, 3" & 5" ArH); ¹³C NMR (125 MHz, δ , ppm): 32.53 (1C, 5C, Thiaz) 61.64 (1C, 2C, Thiaz) 121.38 (2C, 2' & 6' ArC) 123.96 (2C, 3" and 5" ArC) 130.53 (4C, 3', 5', 2" & 6" ArC) 139.96 (1C, 4' ArC) 141.44 (1C, 1" ArC) 147.18 (1C, 1" ArC) 147.29 (1C, 4" ArC) 170.65 (1C, C = O); Mass (m/z): 380 (M⁺+1).

4-(2-(4-fluorophenyl)-4-oxothiazolidin-3-yl)benzenesulfonamide 4m

Pale brownish powder, Yield: 32.65%; MP: 128 °C; R_f: 0.81; IR (KBr, cm⁻¹): 3385.74 (NH str), 1680 (C = O str), 1385 (SO₂ str), 1110.32 (C-F str), 618.28 (C-S str), ¹H NMR (500 MHz, δ , ppm): 3.97 (q, 2H, 5-CH₂, Thiaz) 6.41 (s, 1H, 2-CH, Thiaz) 6.94 (s, 2H, SO₂NH₂) 7.14 (m, 2H, 3" & 5" ArH) 7.22 (m, 2H, 2" & 6" ArH) 7.49 (d, 2H, 2' & 6' ArH) 7.8 (d, 2H, 3' & 5' ArH); ¹³C NMR (125 MHz, δ , ppm): 33.48 (1C, 5C, Thiaz) 72.9 (1C, 2C, Thiaz) 115.5 (2C, 3" and 5" ArC) 121.4 (2C, 2' & 6' ArC) 129.3 (2C, 3' & 5' ArC) 130.5 (2C, 2" and 6" ArC) 134.6 (1C, 1" ArC) 136.4 (1C, 4' ArC) 143.8 (1C, 1' ArC) 160.7 (1C, 4" ArC) 172.83 (1C, C = O); Mass (m/z): 353.01 (M⁺+1), 354.01 (M⁺+2).

4-(4-oxo-2-styrylthiazolidin-3-yl)benzenesulfonamide 4n

Pale brownish powder, Yield: 24.44%; MP: 202 °C; R_f: 0.70; IR (KBr, cm⁻¹): 3384.84 (NH str), 1670 (C = O str), 1382 (SO₂ str), 618.27 (C-S str), ¹H NMR (500 MHz, δ , ppm): 4.4 (q, 2H 5-CH₂, Thiaz) 5.7 (d, 1H 2-CH Thiaz) 6.53 (m, 1H CH) 6.88 (d, 1H CH) 6.89 (m, 2H SO₂NH₂) 7.29 (m, 2H 3" & 5" ArH) 7.30 (m, 1H 4" ArH) 7.44 (m, 2H 2' & 6' ArH) 7.72 (m, 2H 2" & 6" ArH) 7.8 (m, 2H 3' & 5' ArH); ¹³C NMR (125 MHz, δ , ppm): 34.1 (1C, 5C, Thiaz) 58.5 (1C, 2C, Thiaz) 121.3 (2C, 2' & 6' ArC) 123.2 (1C, CH) 127.8 (1C, 4" ArC) 128.6 (2C, 2" and 6" ArC) 128.8 (2C, 3" and 5" ArC) 129.6 (2C, 3' & 5' ArC) 129.8 (1C, CH) 136.5 (1C, 1" ArC) 136.6 (1C, 4' ArC) 144.7 (1C, 1' ArC) 171.63 (1C, C = O); Mass (m/z): 361.06 (M⁺+1).

Biological activity

DPPH scavenging assay

A stock solution of DPPH (0.1 mM) in ethanol (95%) was prepared. The concentrations of 40, 80, 160, 320, and 640 μ g of 2,3-disubstituted thiazolidin-4-ones **4a-n** were prepared in ethanol. For the different concentration sample solutions, 2 mL DPPH stock solution was added and the volume was completed to 4 mL with ethanol. The reaction mixtures were allowed to stand for 30 min in the dark and absorbance was recorded at 517 nm against blank. Ascorbic acid was used as reference. The experiment was carried out in triplicate. The free radical scavenging activity was calculated by

$$\% \text{ Scavenging} = \frac{\text{Absorbance of control} - \text{Absorbance of test}}{\text{Absorbance of control}} \times 100 \quad \text{Eq 1}$$

The effective concentration of the sample required to scavenge the DPPH radical by 50% (IC₅₀) was obtained by linear regression analysis between concentration and absorbance [15].

Acute oral toxicity

Acute oral toxicity of 2,3-disubstituted thiazolidin-4-ones (**4a-n**) was performed as per OECD guidelines 425 [16]. The Swiss albino female mice were fasted before dosing for 18 h. The animals were administered experimental compounds at a dose of 2000 mg/kg b.w. as aqueous suspension of 0.5% Tween 80 orally and observed for 4 h after dosing and for 14 days until death.

Antidiabetic activity

The adult healthy Sprague Dawley rats of either sex was fed 30% D-fructose in water for 30 days and their serum glucose level was estimated using a glucose kit at 505 nm on the 0th, 15th and 30th days. The animals which exhibited hyperglycemia and hyperlipidemia were selected for the study. The rats were allowed to continue to receive fructose until the end of the study. The animals were divided into six groups, each consisting of six as follows: Group 1 (normal): received the water orally, Group 2 (control): received 30% fructose solution and 0.5% aqueous Tween 80 p.o., Group 3 (standard): received 30% fructose solution and pioglitazone (15 mg/kg b.w.) aqueous suspension with 0.5% Tween 80 p.o. and Groups 4, 5, 6, and 7 (Diabetic): received 30% fructose solution, compound **4e** (50 mg/kg b.w.) in 0.5% Tween 80 aqueous suspension p.o., compound **4h**, **4i**, and **4n** (200 mg/kg b.w.) in 0.5% Tween 80 aqueous suspension p.o., respectively [17]. Approximately 2 mL of blood was collected and centrifuged at 3500 RPM for 15 min to obtain the serum. The serum glucose was estimated on the 0th, 3rd, 7th, 14th and 21st days. The serum triglyceride and total cholesterol was estimated before and after treatment.

Estimation of serum glucose

The serum glucose was estimated by means of Trinder's method [18]. Glucose reagent 1 mL was added into blank, standard and test. 10 μ L of distilled water was appended to blank, standard with 10 μ L to standard solution and 10 μ L sample serum was added to test. The mixture was mixed well and incubated for 10 min at 37 °C. The absorbance was read against blank at 505 nm. The glucose level was expressed as:

$$\text{Concentration of glucose (mg/dL)} = \frac{\text{absorbance of sample}}{\text{absorbance of glucose standard}} \times \text{concentration of standard} \quad \text{Eq. 2}$$

Estimation of total serum cholesterol

The total serum cholesterol was estimated by the CHOD-PAP method [19]. Into each blank, standard and test 1000 μ L of working reagent was added. Blank was supplemented with 10 μ L distilled water, standard with 10 μ L standard solution and test was appended 10 μ L sample serum, mixed well and incubated for 10 min at 37 °C. The absorbance was read against blank at 505 nm. The cholesterol level was calculated by:

$$\text{Cholesterol} \left(\frac{\text{mg}}{\text{dL}} \right) = \frac{\text{absorbance of test}}{\text{absorbance of standard}} \times \text{Concentration of standard} \quad \text{Eq 3}$$

Estimation of serum triglycerides

The serum triglycerides level was estimated by means of GPO-Trinder's method [20]. Working reagent 1000 μ L was added into each blank, standard, and test. Blank was supplemented with 10 μ L distilled water, standard with 10 μ L standard solution, and test was supplemented with 10 μ L sample serum, mixed well and incubated for 10 min at 37 °C. The absorbance was read at 546 nm against blank. Triglycerides level was calculated by:

$$\text{Triglycerides} \left(\frac{\text{mg}}{\text{dL}} \right) = \frac{\text{absorbance of test}}{\text{absorbance of standard}} \times \text{Concentration of standard} \quad \text{Eq 4}$$

Histopathology

The rats were anaesthetised and exsanguinated. The pancreas from control, standard, and test groups were fixed with 10% formalin and embedded in paraffin wax and cut into longitudinal sections of 4 mm thickness. The sections were stained with haematoxylin and eosin dye for histopathological observation.

Molecular docking

The three-dimensional crystal structure of PPAR- γ with PDB ID: 4PRG [22] was downloaded from the RCSB protein data bank. Water molecules were removed followed by the addition of Kollaman's charges and polar hydrogens using AutoDock 1.5.6 tools [23]. The structures of ligands **4a-n** were drawn by ChemDraw 16.0 and copied as SMILES. The Avogadro tool [24] was used to generate the .pdb format of ligand with energy minimisation. The .pdb file was converted into .pdbqt format for input into AutoDock 1.5.6 tools. The grid file was generated and saved as .gpf. The Lamarckian Genetic Algorithm was used to find the conformers with the

lowest binding energies and the .dpf file was generated to dock. The final docking task was performed by Autodock vina [25]. The docked file was saved as .dlg format and analysed for the binding interaction of ligand with target in different conformations. The highest binding energy conformation of ligand was selected and its interactions were visualised in Biovia discovery studio 2021 [26].

Physicochemical, drug likeness and ADME properties

The ADME properties and drug likeness of 2,3-disubstituted thiazolidin-4-ones (**4a-n**) was predicted by Swiss Prediction web tool [27]. The structures of compounds **4a-n** and pioglitazone were drawn in ChemDraw 16.0 and copied as SMILES and then submitted one by one to web tool to predict the physicochemical, ADME, and drug likeness properties.

Statistics

The data of animal experiments were expressed as mean \pm standard error of mean (SEM) and analysed by means of Graphpad prism version 8.0.0 (Graphpad Software, San Diego, USA) [21]. The statistical difference between the treatments and various groups were tested by analysis of variance (ANOVA), followed by Dunnett's multiples comparison test. Data with *p*-value < 0.01 was considered significant.

Results and discussion

The hypoglycemic side effect of antimicrobial sulfonamides led to the development of the sulfonyl urea class of hypoglycemic agents such as tolbutamide and chlorpropamide. Glitazone antidiabetics such as rosiglitazone and pioglitazone show their activity due to the presence of acidic thiazolidinedione head. Enthused by this fact, the title compounds which have both the features of sulfonamide and thiazolidin-4-one (that closely resembles the thiazolidinedione present in glitazone antidiabetics) in the same molecule were designed by way of the molecular hybridisation strategy [28] with an anticipation of better hypoglycemic activity [29].

Chemistry

The design and synthesis of the title compounds 2,3-disubstituted thiazolidin-4-ones (**4a-n**) are illustrated in Figure 1. A series of Schiff bases **3a-n** was prepared by condensing substituted aromatic aldehydes with sulphanilamide in absolute alcohol with moderate to excellent yield.

The IR spectra of compounds **3a-n** showed the characteristic C = N stretching frequency between 1604.77 and 1627 cm⁻¹ among others. Further, compounds **3a-n** were reacted with thioglycolic acid in presence of anhydrous zinc chloride to yield compounds **4a-n** in moderate to excellent yield. The 2,3-disubstituted thiazolidinones **4a-n** displayed the methylene protons of the C-5 thiazolidinone ring at δ 3.9-4.4 ppm as quartet in ¹H NMR. The proton of thiazolidinone C-2 resonated at δ 5.7-6.81 ppm among other aromatic protons. The thiazolidinone C-2 appeared at δ ~ 62 ppm while C-5 resonated at δ ~ 33 ppm and C-4 showed signal at δ ~ 171 ppm amidst other carbons in ¹³C NMR. Further, the presence of C-S stretching at 617.93-618.80 cm⁻¹, SO₂ stretching at 1382-1385 cm⁻¹ and C = O stretching at 1606-1680.92 cm⁻¹ in IR spectrum confirmed their structure. Compounds **4a-n** displayed M⁺ and M⁺+1 ion peaks in their required value in mass spectrum suggesting their formation.

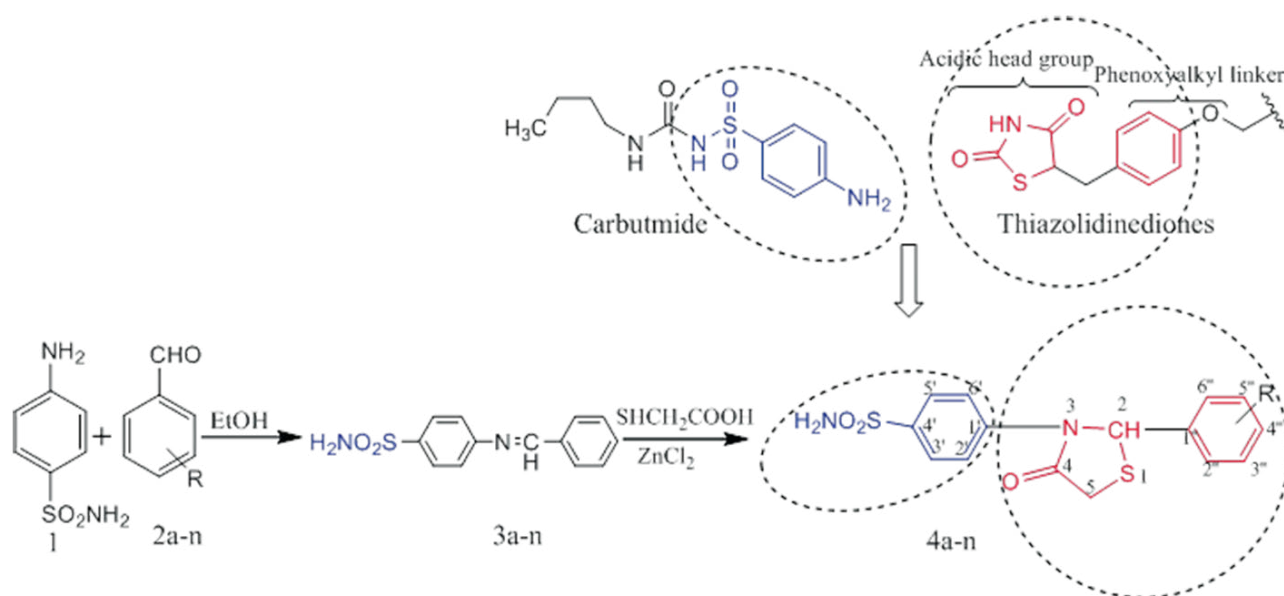


Figure 1. Design and synthesis of 2,3-disubstituted thiazolidin-4-ones.

R = a : H; b : 4-CH₃; c : 4-OCH₃; d : 2-OH; e : 4-OH; f : 4-N(CH₃)₂; g : 3,4-(OCH₃)₂; h : 4-Cl; i : 2,4-Cl₂; j : 2,6-Cl₂; k : 4-Br; l : 4-NO₂; m : 4-F.

DPPH scavenging assay

The DPPH radical scavenging activity of 2,3-disubstituted thiazolidin-4-ones 4a-n is presented on Table 1. The DPPH radical scavenging % was calculated as per equation 1. The *p*-chloro substituted compound 4h exhibited weak DPPH scavenging activity (IC₅₀ = 355.83 µg) followed by 2,4-dichloro substituted 4i (IC₅₀ = 359.101 µg), 4n with vinyl group (IC₅₀ = 372.28 µg), and *p*-hydroxy substituted 4e (IC₅₀ = 440.84 µg) when compared with standard, ascorbic acid (IC₅₀ = 9.25 µg). The remainder of the compounds did not exhibit good DPPH scavenging activity. Compounds with electron attracting substituents exhibited better DPPH scavenging than those with electron releasing substituents.

Table 1. DPPH Scavenging Activity of 2,3-disubstituted thiazolidin-4-ones (4a-n).

Compound	IC ₅₀ , µg
4a	9920
4b	1349.70
4c	2280
4d	1016.77
4e	440.84
4f	1857.36
4g	952.28
4h	355.83
4i	359.101
4j	1975.95
4k	6556
4l	786.85
4m	2737.27
4n	372.28
Asc acid	9.25

Acute oral toxicity

Compounds **4e**, **4h**, **4i**, and **4n** were found to be safe at the dose of 2000 mg/kg b.w. Thus, they were tested at 200 mg/kg b.w. and compound **4e** at 50 mg/kg b.w. orally for *in vivo* antidiabetic activity.

Antidiabetic activity

It is a well-known fact that, diabetes mellitus is associated with oxidative stress [30] and compounds having the antioxidant property will be of great value in the treatment of diabetes. Therefore, the thiazolidin-4-ones that have exhibited good antioxidant activity in the DPPH assay namely, **4e**, **4h**, **4i** and **4n** were selected for antidiabetic activity. The antidiabetic activity was carried out by estimating the serum glucose level as per equation 2 in rats with fructose induced diabetes. The estimation of the serum glucose level was carried out by Trinder's method in which, glucose oxidase reacts with glucose, water, and oxygen to form gluconic acid and hydrogen peroxide. The enzyme peroxidase catalyses the oxidative coupling of 4-aminoantipyrine with phenol in the presence of hydrogen peroxide to yield a purple coloured quinoneimine complex with absorbance proportional to the concentration of glucose [31].

The administration of fructose resulted in the induction of diabetes in rats when compared to the normal group. The tested compounds exhibited highly significant antidiabetic activity when compared with the control group. Compound **4e** with *p*-hydroxy substitution reduced serum glucose by 67.93% followed by compound **4h** (*p*-Cl) by 66.03%, compound **4i** (2,4-Cl₂) by 65.29%, and compound **4n** by 62.68% on the 21st day of the treatment. All the compounds exhibited higher glucose lowering activity than the standard drug pioglitazone. Contrary to DPPH scavenging activity, *p*-OH substitution seems to favour the activity more than *p*-Cl and 2,4-Cl₂. The reduction in the serum glucose level by way of compounds was found to be significant when compared with pioglitazone on the 3rd day, compound **4e** exhibited a significant reduction in the serum glucose level on the 14th day. The results are tabulated on Table 2.

Table 2. Antidiabetic Activity of Compounds **4e**, **4h**, **4i** and **4n**.

Groups	Serum glucose, mg/dl				
	0 th Day	3 rd Day	7 th Day	14 th Day	21 st Day
Normal	85.74±7.09	89.26±5.49	85.28±4.05	90.08±3.60	90.56±4.38
Control	186.5±19.37	202.6±11.11a	193.5±8.39a	191.7±7.66a	199.0±13.95a
Compound 4e (% reduction of serum glucose)	129.3±7.06 (30.67)	83.71±4.64*** ### (58.68)	70.47±0.98*** # (63.58)	67.37±0.48*** ## (64.85)	63.80±0.90*** (67.93)
Compound 4h (% reduction of serum glucose)	110.8±3.85 (40.58)	79.21±2.34*** ### (60.90)	77.52±2.23*** (59.93)	72.52±1.26*** (62.17)	67.59±1.18*** (66.03)
Compound 4i (% reduction of serum glucose)	128.7±10.69 (30.99)	110.2±5.15*** ## (45.60)	82.18±0.98*** (57.52)	76.20±1.70*** (60.25)	69.06±1.79*** (65.29)
Compound 4n (% reduction of serum glucose)	135.3±10.93 (27.45)	112.1±5.50*** ## (44.66)	93.88±3.43*** (51.48)	83.99±2.12*** (56.15)	74.26±0.93*** (62.68)
Pioglitazone (% reduction of serum glucose)	151.0±20.97 (19.03)	144.8±9.25*** (28.52)	89.66±3.96*** (53.66)	86.40±4.18*** (54.92)	85.29±3.47*** (57.14)

All the values are expressed as mean ± SEM, n = 6, ^a*p* < 0.001 as compared to the normal group, ****p* < 0.001 as compared to the control group and ###*p* < 0.001, ##*p* < 0.01 and #*p* < 0.05 as compared to the standard group [ANOVA followed by multiple comparison Dunnett's test].

Total serum cholesterol

Cholesterol esterase catalyses the conversion of cholesterol ester to cholesterol in the presence of water. The released cholesterol is then converted into cholest-4-en-3-one and hydrogen peroxide in the presence of oxygen by cholesterol oxidase. The released hydrogen peroxide reacts with 4-aminoantipyrine and phenol to yield a purple complex of quinoneimine that was measured spectrophotometrically [19]. The serum cholesterol was calculated as per equation 3. The tested compounds exhibited better and significant serum cholesterol lowering activity than pioglitazone, when compared with the control group. Compound **4e** possessed the highest activity followed by **4h**, **4i**, and **4n**. The activity pattern displayed by the compounds is the same as that of antidiabetic activity. The reduction in total cholesterol by tested compounds was found to be insignificant when compared with pioglitazone; only compound **4e** has shown significant activity on the 21st day. The results are tabulated on Table 3.

Table 3. Effect of compounds **4e**, **4h**, **4i** and **4n** on serum total cholesterol and triglycerides.

Group	Total cholesterol level (mg/dl)		Triglycerides level (mg/dl)	
	Before treatment (0 th day)	After treatment (21 st day)	Before treatment (0 th day)	After treatment (21 st day)
Normal	35.96±7.99	22.05±2.05	51.01±9.50	95.75±13.40
Control	65.18±3.58	98.20±4.50a	91.15±9.55	119.5±12.78b
Compound 4e	86.25±8.03	41.20±5.41*** ##	175.7±22.67	91.05±16.07
Compound 4h	69.78±5.01	51.86±0.94***	163.6±19.71	77.47±4.61
Compound 4i	71.82±10.42	52.79±3.76***	156.6±29.25	104.0±15.16#
Compound 4n	71.43±7.26	52.54±6.54***	139.3±19.97	86.03±5.63
Pioglitazone	97.65±20.84	64.10±4.94***	153.0±11.68	52.32±4.51**

All the values are expressed as mean ± SEM, n = 6, ^a*p* < 0.001 as compared to the normal group, ****p* < 0.001 as compared to the control group and ##*p* < 0.01 as compared to the standard group. [One way ANOVA followed by multiple comparison Dunnett's test].

Serum triglycerides

The determination of serum triglycerides was done by means of the GPO-Trinder method. Lipase hydrolyses serum triglycerides to glycerol and free fatty acids. In the presence of ATP and glycerol kinase, the glycerol

is converted to glycerol-3-phosphate. The glycerol-3-phosphate is then oxidised by glycerol phosphate oxidase to dihydroxy acetone phosphate and hydrogen peroxide. The condensation of hydrogen peroxide with TOOS reagent (sodium 3-(ethyl(m-tolyl)amino)-2-hydroxypropane-1-sulfonate) and 4-aminoantipyrine yielded a coloured complex of quinoneimine which absorbs at 546 nm [20]. The serum triglycerides level was calculated as per equation 4. The tested compounds reduced the triglyceride level after the 21st day. However, it was found to be insignificant, whereas pioglitazone significantly reduced triglycerides when compared to the control group. Compound **4h** exhibited the highest serum triglyceride reduction followed by **4n**. Compound **4e** displayed moderate activity. Compound **4i** with 2,4-Cl₂ substitution has significantly reduced serum triglycerides after the 21st day when compared with standard pioglitazone. The results are tabulated on Table 3.

Body weight

The tested compounds were found to decrease the elevated body weight when tested at different time intervals. However, it was insignificant when compared with the control and standard groups. The results are tabulated on Table 4.

Table 4. Effects of compounds **4e**, **4h**, **4i** and **4n** on the body weight of animals.

Groups	Body Weight (gm)				
	0 th Day	3 rd Day	7 th Day	14 th Day	21 st Day
Normal	288.2 ± 9.35	295.8 ± 10.00	282.7 ± 11.74	283.3 ± 12.38	287.7 ± 13.10
Control	317.5 ± 9.90	325.8 ± 9.404	316.3 ± 10.19	322.3 ± 11.86	321.3 ± 10.62
Compound 4e	308.3 ± 14.45	291.0 ± 6.70	285.3 ± 6.03	285.3 ± 7.72	282.7 ± 10.57
Compound 4h	308.7 ± 18.82	299.3 ± 18.99	297.0 ± 19.28	292.7 ± 22.16	291.3 ± 21.57
Compound 4i	294.8 ± 24.19	283.0 ± 21.61	282.7 ± 24.97	285.2 ± 29.32	281.0 ± 273.0
Compound 4n	280.0 ± 19.0	278.7 ± 21.24	276.7 ± 20.88	275.0 ± 21.98	273.0 ± 23.18
Pioglitazone	316.8 ± 7.59	320.2 ± 10.22	295.5 ± 12.53	311.3 ± 10.79	326.0 ± 12.37

All the values are expressed as mean ± SEM, n = 6, as compared to the control and standard group. [One way ANOVA followed by multiple comparison Dunnett's test].

Histopathology

The histopathological investigation of the pancreas of rats belonging to the normal group showed a normal architecture of islets of Langerhans with pale, rounded and ovoid β cells in the centre embedded in the exocrine portion of the pancreas. Whereas, shrinkage of islets of Langerhans with degeneration and necrosis of cells with their nucleus appearing densely basophilic with predominant karyolysis was observed in rats of the control group. The pancreas of diabetic rats treated with pioglitazone showed a normal architecture of islets of Langerhans with pale, large, round to ovoid shape containing cells embedded in the exocrine portion of the pancreas. In the case of a pancreas treated with compounds **4h** and **4i** normal sized islets of Langerhans but some degeneration of β cells in the centre were noticed, suggesting a protective role of compounds **4h** and **4i** on the islets of Langerhans. The histopathological changes are presented in Figure 2.

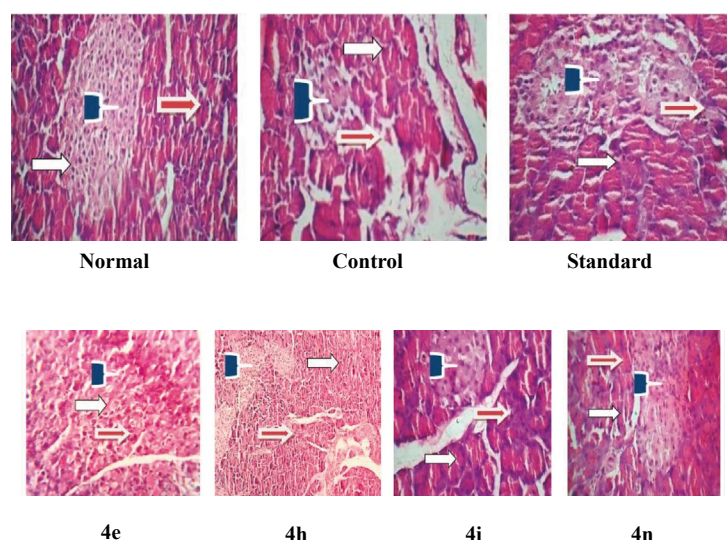


Figure 2. Effect of 2,3-disubstituted thiazolidin-4-ones on pancreatic cell.

Normal: Pancreatic islet of rat showed intact cluster of β -cells and architecture of islets of Langerhans.

Control: Showed destruction of pancreatic islets of β -cells, islet shape and number of islets are reduced.

Standard: Showed apparently reduced degranulation architecture, moderately restored pancreatic cells, islet shape and increased number of islets.

Treatment: 4e, 4h, 4i and 4n treated showing regeneration of β -cells, restoration of architecture, shape and increase in the number of islets.

→ Architecture of β cells of Islets

■ Islets shape

→ Number of β cells of islets

Molecular docking

The compounds that were tested for antidiabetic activity were docked with PPAR- γ (PDB ID: 4PRG) using autodock tools 1.5.6 to explore their interaction. The PPAR- γ plays an important role in the regulation of glucose and homeostasis and was used as a molecular target for the 2,4-thiazolidinedione class (glitazones) of antidiabetic agents [32]. The binding energies and amino acid residues with which compounds **4e**, **4h**, **4i**, **4n** and pioglitazone interacted are presented on Table 5 and in Figure 3. The compounds exhibited more binding energy with PPAR- γ than with pioglitazone.

Table 5. Interactions of compounds **4e**, **4h**, **4i** and **4n** with PPAR- γ .

Compound	Binding energy Kcal/mol	Interacting amino acid residues
4e	-7.6	GLU(B:343), ARG(B:288), LEU(B:340), LEU(B:228), LEU(B:330)
4h	-7.3	GLU(B:460), LEU(B:453), VAL(B:450)
4i	-8.3	ASP(B:475), ILE(B:279), GLU(B:460)
4n	-10.3	ILE(A:267), ARG(A:280), PHE(A:264), LYS(A:265), PHE(A:287)
Pioglitazone	-7.12	GLU(B:343), LYS(C:263), LEU(C:255), MET(C:256), ILE(C:281), ARG(C:280), PHE(C:264), PHE(C:287)

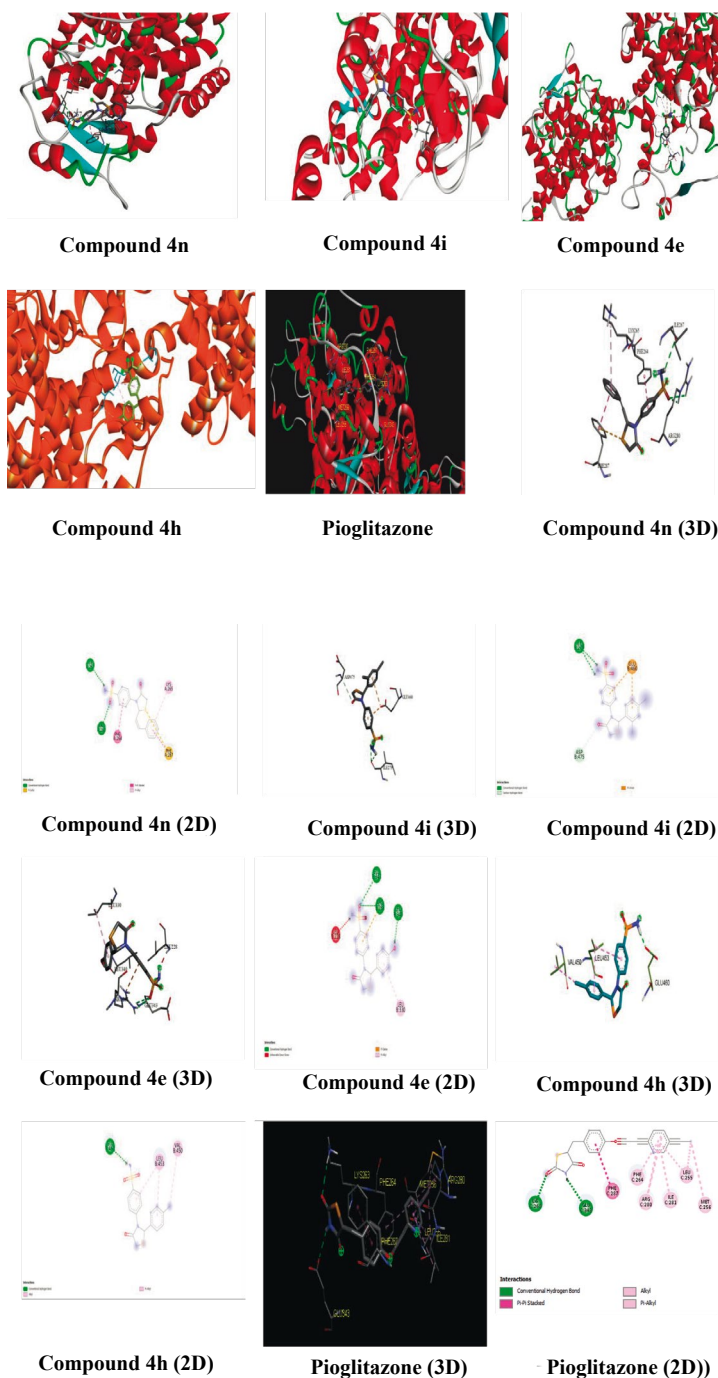


Figure 3. Docking of Compounds and Standard drug with PPAR- γ .

Table 6. *In silico* parameters of 2,3-disubstituted thiazolidin-4-ones (4a-n).

Compounds	Physicochemical properties							Lipophilicity						Drug likeness					Pharmacokinetics	
	MW	Fsp3	RB	HBA	HBD	MR	TPSA	iLOGP	X LOGP3	W LOGP	M LOGP	SILICOS-IT	Consensus Log P	Lipinski	Ghose	Veber	Egan	Muegge	Log KP (cm/s)	F
4a	334.41	0.13	3	4	1	89.55	114.15	1.45	1.93	2.49	1.51	1.12	1.70	0	0	0	0	0	-6.97	0.55
4b	348.44	0.19	3	4	1	94.52	114.15	1.62	2.29	2.80	1.76	1.62	2.02	0	0	0	0	0	-6.80	0.55
4c	364.44	0.19	4	5	1	96.04	123.38	2.13	1.90	2.50	1.21	1.16	1.78	0	0	0	0	0	-7.17	0.55
4d	350.41	0.13	3	5	2	91.57	134.38	1.51	1.57	2.19	0.96	0.64	1.37	0	0	0	1	0	-7.32	0.55
4e	350.41	0.13	3	5	2	91.5	134.38	1.49	1.57	2.19	0.96	0.64	1.37	0	0	0	1	0	-7.32	0.55
4f	377.48	0.24	4	4	1	103.76	117.39	1.76	2.05	2.55	1.45	0.77	1.72	0	0	0	0	0	-7.15	0.55
4g	394.47	0.24	5	6	1	102.53	132.61	2.26	1.87	2.51	0.92	1.21	1.75	0	0	0	1	0	-7.38	0.55
4h	368.86	0.13	3	4	1	94.56	114.15	2.13	2.55	3.14	2.02	1.75	2.32	0	0	0	0	0	-6.74	0.55
4i	403.30	0.13	3	4	1	99.57	114.15	2.41	3.18	3.80	2.53	2.40	2.86	0	0	0	0	0	-6.50	0.55
4j	403.30	0.13	3	4	1	99.57	114.15	1.77	3.18	3.80	2.53	2.40	2.74	0	0	0	0	0	-6.50	0.55
4k	413.31	0.13	3	4	1	97.25	114.15	1.84	2.62	3.25	2.15	1.79	2.33	0	0	0	0	0	-6.96	0.55
4l	379.41	0.13	4	6	1	98.37	159.97	1.54	1.75	2.40	0.59	-1.04	1.04	0	0	1	1	1	-7.37	0.55
4m	352.40	0.13	3	5	1	89.51	114.15	1.94	2.03	3.05	1.90	1.54	2.09	0	0	0	0	0	-7.01	0.55
4n	360.45	0.12	4	4	1	99.48	114.15	1.86	2.57	3.04	1.66	1.68	2.16	0	0	0	0	0	-6.67	0.55
Pioglitazone	356.44	0.32	7	4	1	99.48	93.59	2.61	3.75	2.78	2.01	4.28	3.09	0	0	0	0	0	-5.81	0.55

MW: Molecular Weight, Fsp3: Fraction of sp³ Carbon atoms, RB: Rotatable Bonds, HBA: Hydrogen Bond Acceptor, HBD: Hydrogen Bond Donor, MR: Molar Refractivity, TPSA: Topological Polar Surface Area, LogP: Indicator of Lipophilicity, Log KP: Skin Permeation, F: Bioavailability Score

It is interesting that, the tested compounds exhibited higher binding energy with PPAR- γ . The compound **4n** demonstrated the highest binding energy with -10.3 *kcal/mol* among the tested compounds and formed two hydrogen bonds with amino acid residues ILE (A:267) and ARG (A:280), and also formed other bonds by interacting with other amino acid residues PHE (A:264), LYS (A:265), and PHE (A:287). Likewise, compound **4i** showed good binding energy of -8.3 *kcal/mol* and formed two hydrogen bonds with ASP (B:475) and ILE (B:279), and also formed another bond with GLU (B:460). Compound **4e** displayed good binding energy of -7.6 *kcal/mol* and formed three hydrogen bonds with GLU (B:343), ARG (B:288), and LEU (B:340), and formed other bonds with LEU (B:228) and LEU (B:330). Compound **4h** also showed binding energy of -7.3 *kcal/mol* and formed a hydrogen bond with GLU (B:460), and other bonds with LEU (B:453) and VAL (B:450). Compound **4n** interacted with the A chain while, others with the B chain of PPAR- γ . All the results are compared with pioglitazone which had binding energy of -7.12 *kcal/mol* and formed two hydrogen bonds with GLU (B:343) and LYS (C:263), and other bonds with PHE (C:287), LEU (C:255), MET (C:256), ILE (C:281), PHE (C:264), and ARG (C:280) with the B and C chains. Thus, compound **4n** exhibited excellent binding energy and interaction with PPAR- γ followed by **4i**, **4e** and **4h** when compared with pioglitazone. It is interesting to note that, the compounds which showed good antidiabetic activity interacted with a lower binding energy with PPAR- γ .

Physicochemical, ADME, and drug likeness properties

For a molecule to be effective as a potent drug, it must reach its target in the body in sufficient concentration and stay there in a bioactive form long enough for the expected biologic events to occur. Drug development involves assessments of pharmacokinetic properties such as absorption, distribution, metabolism, and excretion (ADME); in this context computer models constitute valid alternatives to experiments. The Swiss ADME web

tool provides easy efficient inputs and free access to a pool of past yet robust predictive models for the physicochemical properties, pharmacokinetics, and drug likeness of compounds. The ADME and drug likeness of 2,3-disubstituted thiazolidin-4-ones (**4a-n**) were determined by the Swiss ADME web tool and the results are tabulated on Table 6.

The filters used to evaluate the drug likeness of the title compounds were i) Lipinski filter: MW \leq 500, MLogP \leq 4.15, HBA \leq 10, HBD \leq 5 [33] ii) Ghose filter: 160 \leq MW \leq 480, -0.4 \leq WlogP \leq 5.6, 40 \leq MR \leq 130, 20 \leq atoms \leq 70 [34] iii) Veber (GSK) filter: RB \leq 10, TPSA \leq 140 [35] iv) Egan (Pharmacia) filters: WLogP \leq 5.88, TPSA \leq 131.6 [36] v) Muegge (Bayer) filter : 200 \leq MW \leq 600, -2 \leq XLogP \leq 5, TPSA \leq 157; HBD \leq 5, RB \leq 15, number of rings \leq 7; number of carbons $>$ 4, number of heteroatoms \leq 1 [37]. The compounds did not violate the Lipinski and Ghose rule but, some violated the Veber, Egan, and Muegge rules. To summarise, 2,3-disubstituted thiazolidin-4-ones met the criteria of drug likeness and exhibited satisfactory ADME parameters.

Conclusions

A series of 2,3-disubstituted thiazolidinones **4a-n** were synthesised from Schiff base intermediates **3a-n** that have been derived from sulfanilamide. The final products were characterised by physicochemical parameters and spectral data.

Compounds **4e**, **4h**, **4i**, and **4n** exhibited weak DPPH radical scavenging activity. The acute toxicity study revealed the safety of compounds **4e**, **4h**, **4i** and **4n** at the dose of 2000 mg/kg b.w. p.o. Compounds **4h**, **4i** and **4n** were tested at 200 mg/kg b.w. and compound **4e** at 50 mg/kg b.w. orally for in vivo antidiabetic activity. Compounds **4e** with p-hydroxy substitution, **4h** with p-chloro substitution, **4i** with 2,4-dichloro substitution, and **4n** with vinyl substitution exhibited highly significant antidiabetic activity when compared with the control group. The tested compounds exhibited better and significant serum cholesterol lowering activity with reference to the control group. Compound **4e** possessed the highest cholesterol lowering activity followed by **4h**, **4i**, and **4n**. The tested compounds also

reduced the serum triglyceride level after the 21st day, however this was found to be insignificant in comparison to the control group. Compound **4h** exhibited the highest serum triglyceride reducing activity followed by **4n** and **4e**. The tested compounds reduced elevated body weight but it was found to be insignificant when compared with the control group. The histopathology demonstrated the protective role of compounds **4h** and **4i** on the β cells of islets of Langerhans. The molecular docking showed compound **4n** binds with high energy more than pioglitazone with PPAR- γ . The physicochemical, ADME, and drug likeness properties of most of the synthesised compounds were found to be within permitted limits. Thus, the molecular architecture developed and synthesised in this study would be of great interest with good antidiabetic potential warranting further work.

References

- [1] Louis Sanford Goodman, Alfred Zack Gilman, and L. L. Brunton, *Goodman and Gilman's the pharmacological basis of therapeutics*, 13th ed. New York, N.Y.: McGraw-Hill, 2018, pp. 865-869.
- [2] Statista, "Diabetics number top countries 2019 | Statista," *Statista*, 2019. <https://www.statista.com/statistics/281082/countries-with-highest-number-of-diabetics/> (accessed Mar. 27, 2022).
- [3] P. Zhang, T. Li, X. Wu, E. C. Nice, C. Huang, and Y. Zhang, "Oxidative stress and diabetes: antioxidative strategies," *Frontiers of Medicine*, vol. 14, no. 5, pp. 583-600, Apr. 2020, DOI: <https://doi.org/10.1007/s11684-019-0729-1>.
- [4] N. Agrawal, "Synthetic and therapeutic potential of 4-thiazolidinone and its analogs," *Current Chemistry Letters*, vol. 10, no. 2, pp. 119-138, 2021, doi: <https://doi.org/10.5267/j.ccl.2020.11.002>.
- [5] R. Bhutani, D. P. Pathak, G. Kapoor, A. Husain, and Md. A. Iqbal, "Novel hybrids of benzothiazole-1,3,4-oxadiazole-4-thiazolidinone: Synthesis, in silico ADME study, molecular docking and in vivo anti-diabetic assessment," *Bioorganic Chemistry*, vol. 83, pp. 6-19, Mar. 2019, DOI: <https://doi.org/10.1016/j.bioorg.2018.10.025>.
- [6] A. Abdul Hameed, and F. Hassan, "Synthesis, Characterization and Antioxidant Activity of Some 4-Amino-5-Phenyl-4h-1, 2, 4-Triazole-3-Thiol Derivatives," *International Journal of Applied Science and Technology*, vol. 4, no. 2, pp. 202-211, Mar. 2014.
- [7] M. Mohamed Ghorab, D. A. Abou El Ella, H. I. Heiba, and A. M. Soliman, "Synthesis of Certain New Thiazole Derivatives Bearing a Sulfonamide Moiety with Expected Anticancer and Radiosensitizing Activities," *Journal of Materials Science and Engineering A*, vol. 1, no. 5, pp. 684-691, 2011.
- [8] M. A. Bhat, M. A. Al-Omar, A. M. Naglah, and A. A. Khan, "Synthesis of Novel Sulfamethoxazole 4-Thiazolidinone Hybrids and Their Biological Evaluation," *Molecules*, vol. 25, no. 16, p. 3570, Aug. 2020, DOI: <https://doi.org/10.3390/molecules25163570>.
- [9] T. Nasr, S. Bondock, and S. Eid, "Design, synthesis, antimicrobial evaluation and molecular docking studies of some new 2,3-dihydrothiazoles and 4-thiazolidinones containing sulfoxazole," *Journal of Enzyme Inhibition and Medicinal Chemistry*, vol. 31, no. 2, pp. 236-246, Mar. 2015, DOI: <https://doi.org/10.3109/14756366.2015.1016514>.
- [10] R. V. Shingalapuri, K. M. Hosamani, R. S. Keri, and M. H. Hugar, "Derivatives of benzimidazole pharmacophore: Synthesis, anticonvulsant, antidiabetic and DNA cleavage studies," *European Journal of Medicinal Chemistry*, vol. 45, no. 5, pp. 1753-1759, May 2010, DOI: <https://doi.org/10.1016/j.ejmech.2010.01.007>.
- [11] P. P. Prabhu, A. Pai, Padmashree, and R. Shabaraya, "Analgesic and Anti-Inflammatory Activity Studies of Some new Aryl 4-Thiazolidinones in Experimental Mice," *Pharmacologyonline*, vol. 1, pp. 1132-1139, 2010.
- [12] K. I. Bhat, A. Kumar, and P. Kumar, "Synthesis, Cytotoxic and Antihyperlipidemic Activities of Some New coumarinyl 4-Thiazolidinone Derivatives," *Indian Journal of Pharmaceutical Education and Research*, vol. 55, no. 3s, pp. S807-S813, Nov. 2021, DOI: <https://doi.org/10.5530/ijper.55.3s.188>.
- [13] C. Supuran, A. Nicolae, and A. Popescu, "Carbonic anhydrase inhibitors. Part 35. Synthesis of Schiff bases derived from sulfanilamide and aromatic aldehydes: the first inhibitors with equally high affinity towards cytosolic and membrane-bound isozymes," *European Journal of Medicinal Chemistry*, vol. 31, no. 6, pp. 431-438, Jan. 1996, DOI: [https://doi.org/10.1016/0223-5234\(96\)85163-4](https://doi.org/10.1016/0223-5234(96)85163-4).
- [14] E. H. ZIMAM, "Synthesis and characterization of some new azetidinone, thiazolidinone and imidazolidinone derivatives from 2-aminopyridine," *Acta Chim. Pharm. Indica*, vol. 4, no. 4, pp. 180-190, 2014.
- [15] S.-F. Barbuceanu, D. Ilies, G. Saramet, V. Uivarosi, C. Draghici, and V. Radulescu, "Synthesis and Antioxidant Activity Evaluation of New Compounds from Hydrazinecarbothioamide and 1,2,4-Triazole Class Containing Diarylsulfone and 2,4-Difluorophenyl Moieties," *International Journal of Molecular Sciences*, vol. 15, no. 6, pp. 10908-10925, Jun. 2014, DOI: <https://doi.org/10.3390/ijms150610908>.
- [16] "OECD/OCDE 425 OECD GUIDELINE FOR TESTING OF CHEMICALS Acute Oral Toxicity-Up-and-Down Procedure INTRODUCTION," 2001. Accessed: Mar. 22, 2022. [Online]. Available: https://ntp.niehs.nih.gov/iccvm/suppdocs/fedddocs/oecd/oecd_gl425-508.pdf
- [17] M. O. Germoush, H. A. Elgebaly, S. Hassan, and A. M. Mahmoud, "Anti - Diabetic Effects of Padina Pavonia in Fructose - Induced Diabetic Rats," *Aljouf Science and Engineering Journal*, vol. 2, no. 2, pp. 10-16, Jun. 2015, DOI: <https://doi.org/10.12816/0023935>.
- [18] "GLUCOSE GOD -POD METHOD, END POINT INTENDED USE Diagnostic reagent for quantitative in vitro determination of Glucose in human serum, plasma and urine. CLINICAL SIGNIFICANCE," <https://labsakha.com/wpcontent/uploads/2019/02/ErbaGlucose.pdf?189db0&189db0>
- [19] "Cholesterol," https://janissahealthcare.com/downloads/CHOLESTEROL_BLT00034_35_36.pdf (accessed Apr. 28, 2022).
- [20] B. Winter, G. Fiskum, and L. Gallo, "Effects of L-carnitine on serum triglyceride and cytokine levels in rat models of cachexia and septic shock," *British Journal of Cancer*, vol. 72, no. 5, pp. 1173-1179, Nov. 1995, DOI: <https://doi.org/10.1038/bjc.1995.482>.
- [21] J. Xun *et al.*, "Histone demethylase KDM6B inhibits breast cancer metastasis by regulating Wnt/ β -catenin signaling," *FEBS Open Bio*, vol. 11, no. 8, pp. 2273-2281, Jul. 2021, DOI: <https://doi.org/10.1002/2211-5463.13236>.
- [22] R. P. D. Bank, "RCSB PDB - 4PRG: 0072 PARTIAL AGONIST PPAR GAMMA COCRYSTAL," www.rcsb.org. <https://www.rcsb.org/structure/4PRG> (accessed Jul. 12, 2022).
- [23] G. M. Morris *et al.*, "AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility," *Journal of Computational Chemistry*, vol. 30, no. 16, pp. 2785-2791, Dec. 2009, DOI: <https://doi.org/10.1002/jcc.21256>.
- [24] M. D. Hanwell, D. E. Curtis, D. C. Lonie, T. Vandermeersch, E. Zurek, and G. R. Hutchison, "Avogadro: an advanced semantic chemical editor, visualization, and analysis platform," *Journal of Cheminformatics*, vol. 4, no. 1, Aug. 2012, DOI: <https://doi.org/10.1186/1758-2946-4-17>.
- [25] O. Trott, and A. J. Olson, "AutoDock Vina: Improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading," *Journal of Computational Chemistry*, vol. 31, no. 2, p. NA-NA, 2009, DOI: <https://doi.org/10.1002/jcc.21334>.
- [26] "BIOVIA Discovery Studio - BIOVIA - Dassault Systèmes®," www.3ds.com. <https://www.3ds.com/products-services/biovia/products/molecular-modeling-simulation/biovia-discovery-studio/> (accessed Jul. 15, 2022).
- [27] A. Daina, O. Michielin, and V. Zoete, "SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules," *Scientific Reports*, vol. 7, no. 1, Mar. 2017, DOI: <https://doi.org/10.1038/srep42717>.
- [28] Claudio Viegas-Junior, Eliezer J. Barreiro, and Carlos Alberto Manssour Fraga, "Molecular Hybridization: A Useful Tool in the Design of New Drug Prototypes," *Current Medicinal Chemistry*, vol. 14, no. 17, pp. 1829-1852, Jul. 2007, DOI: <https://doi.org/10.2174/092986707781058805>.

- [29] C. O. Wilson, O. Gisvold, J. H. Block, and J. M. Beale, *Wilson and Gisvold's Textbook of organic medicinal and pharmaceutical chemistry*, 12th ed. Philadelphia, Pa.: Wolters Kluwer Health/Lippincott Williams & Wilkins, 2011, pp. 670-684.
- [30] Kenneth Maiese, Zhao Zhong Chong, and Yan Chen Shang, "Mechanistic Insights Into Diabetes Mellitus and Oxidative Stress," *Current Medicinal Chemistry*, vol. 14, no. 16, pp. 1729-1738, Jul. 2007, DOI: <https://doi.org/10.2174/092986707781058968>.
- [31] J. A. Lott, and K. Turner, "Evaluation of Trinder's Glucose Oxidase Method for Measuring Glucose in Serum and Urine," *Clinical Chemistry*, vol. 21, no. 12, pp. 1754-1760, Nov. 1975, DOI: <https://doi.org/10.1093/clinchem/21.12.1754>.
- [32] S. S. Jangam, and S. B. Wankhede, "Synthesis, Molecular Docking, and Biological Evaluation of the New Hybrids of 4-Thiazolidinone and 4(3H)-Quinazolinone Against Streptozotocin Induced Diabetic Rats," *Russian Journal of General Chemistry*, vol. 89, no. 5, pp. 1029-1041, May 2019, DOI: <https://doi.org/10.1134/s1070363219050256>.
- [33] C. A. Lipinski, F. Lombardo, B. W. Dominy, and P. J. Feeney, "Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings," *Advanced Drug Delivery Reviews*, vol. 23, no. 1-3, pp. 3-25, Jan. 1997, DOI: [https://doi.org/10.1016/s0169-409x\(96\)00423-1](https://doi.org/10.1016/s0169-409x(96)00423-1).
- [34] A. K. Ghose, V. N. Viswanadhan, and J. J. Wendoloski, "A Knowledge-Based Approach in Designing Combinatorial or Medicinal Chemistry Libraries for Drug Discovery. 1. A Qualitative and Quantitative Characterization of Known Drug Databases," *Journal of Combinatorial Chemistry*, vol. 1, no. 1, pp. 55-68, Jan. 1999, DOI: <https://doi.org/10.1021/cc9800071>.
- [35] D. F. Veber, S. R. Johnson, H.-Y. Cheng, B. R. Smith, K. W. Ward, and K. D. Kopple, "Molecular Properties That Influence the Oral Bioavailability of Drug Candidates," *Journal of Medicinal Chemistry*, vol. 45, no. 12, pp. 2615-2623, Jun. 2002, DOI: <https://doi.org/10.1021/jm020017n>.
- [36] W. J. Egan, K. M. Merz, and J. J. Baldwin, "Prediction of Drug Absorption Using Multivariate Statistics," *Journal of Medicinal Chemistry*, vol. 43, no. 21, pp. 3867-3877, Oct. 2000, DOI: <https://doi.org/10.1021/jm000292e>.
- [37] I. Muegge, S. L. Heald, and D. Brittelli, "Simple Selection Criteria for Drug-like Chemical Matter," *Journal of Medicinal Chemistry*, vol. 44, no. 12, pp. 1841-1846, Jun. 2001, DOI: <https://doi.org/10.1021/jm015507e>.

Article citation:

S. R. Deshpande, M. S. Wader, P.V. Malagi & P. C. Jaliha, "Design, synthesis, *in silico* studies, and antidiabetic activity of several sulfanilamide incorporated 2,3-disubstituted thiazolidin-4-ones", *Rev. Colomb. Quim.*, vol. 51, no. 3, pp. 3-13 2022. DOI:<https://doi.org/10.15446/rev.colomb.quim.v51n3.105911>