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Camellia cattienensis: phytochemical and biological properties from the leaf extract



Camellia cattienensis: propiedades fitoquímicas y biológicas del extracto de hojas

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ABSTRACT

Keywords:

Antibacterial Antioxidant Cat Tien camellia GC/MS Camellia consists of many plants of high economic importance that are used in different fields, especially in the food industry. Camellia cattienensis is a rare species and is native to Vietnam. Studies on the phytochemical and biological properties of this species have been unknown so far. In this study, the chemical components of the acetone extract from C. cattienensis leaves and its fractions, including n-hexane, chloroform, and ethyl acetate, were investigated using gas chromatography-mass spectrometry assay. Accordingly, 2-pentanone, 4-hydroxy-4-methyl- was the most abundant compound in the acetone extract and the ethyl acetate fraction, while phenol, 2,4-bis(1,1-dimethylethyl)-, phosphite (3:1) and hexanedioic acid, bis(2-ethylhexyl) ester are the richest components in the chloroform and n-hexane fractions, respectively. Furthermore, the acetone extract was active against four tested bacteria, including Klebsiella pneumoniae, Staphylococcus aureus, and Staphylococcus saprophyticus BAA750. The acetone extract of the C. cattienensis leaves also possessed DPPH, and ABTS free radical scavenging with the IC₅₀ values of 91.63±1.88 and 13.32±0.49 µL, respectively. The outcomes of this study hold promise for potential applications of C. cattienensis leaves in food product development, especially in the future beverage industry.

RESUMEN

Palabras clave: Antibacterianas Antioxidantes Cat tien camellia GC/MS La Camelia se compone de muchas plantas de alto valor económico y se utilizan en diferentes campos, especialmente en la industria alimentaria. Camellia cattienensis es una especie rara y originaria de Vietnam. Hasta el momento se desconocen los estudios sobre las propiedades fitoquímicas y biológicas de esta especie. En este estudio, se investigaron los componentes químicos del extracto de acetona de las hojas de C. cattienensis y sus fracciones, como n-hexano, acetato de etilo y cloroformo, mediante un ensayo de cromatografía de gases/espectrometría de masas. En consecuencia, 2-pentanona, 4-hidroxi-4-metil-fue el compuesto más abundante en el extracto de acetona y la fracción de acetato de etilo, mientras que fenol, 2,4-bis(1,1-dimetiletil)-, fosfito (3:1) y el ácido hexanodioico, el éster bis(2-etilhexílico) son los componentes más ricos en las fracciones de cloroformo y n-hexano, respectivamente. Además, se descubrió que el extracto de acetona era eficaz contra cuatro bacterias analizadas, incluidas Klebsiella pneumoniae, Staphylococcus aureus, y Staphylococcus saprophyticus. El extracto de acetona de las hojas de C. cattienensis también poseía eliminación de radicales libres DPPH y ABTS con valores de IC₅₀ de 91,63±1,88 y 13,32±0,49 μL, respectivamente. Los resultados de este estudio son prometedores para aplicaciones potenciales de las hojas de C. cattienensis en el desarrollo de productos alimenticios, especialmente en la industria de bebidas en el futuro.

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amellia is a large genus in the family of Theaceae, comprising over 280 species distributed in many countries around the world, and about 95 species have been found in Vietnam (Quach et al. 2021). Many members of the Camellia genus are plants of high economic value due to their byproducts, including oil seeds, tea, ornamental plants, and iconic flowering shrubs (Yang et al. 2016). Additionally, various solvent extracts obtained from Camellia plants have been reported to possess numerous pharmaceutical properties, including antibacterial, antifungal, antitumor, and antioxidant activities (Yang et al. 2016). Notably, Camellia sinensis, commonly known as the tea plant, is cultivated in tropical and subtropical regions. Additionally, leaf extracts from this species are recognized as the second most consumed beverage in the world (Chitsazan 2015). Moreover, they contain numerous beneficial compounds, such as catechins (Gaur and Bao 2021), steroids, alkaloids, polyphenols, and terpenoids (Anand et al. 2015). Furthermore, the oil extracted from certain Camellia species, particularly C. oleifera and C. japonica, is known as 'Asian olive oil' due to its high content of major chemical compounds, such as oleic acid and neutral lipids (Kim et al. 2014).

Camellia cattienensis Orel was first reported as a new species for the flora of Vietnam by Orel and Wilson (2011). This species, whose type specimen was collected from Cat Tien National Park, Dong Nai Province, Vietnam. To date, it is a rare plant and is found only in the type collection (Orel and Wilson 2011). The phytochemical and biological effects of this species have been unknown so far. This study, thus, provided the chemical constituents, antioxidants, and antibacterial activities of *C. cattienensis* for the first time.

MATERIALS AND METHODS

Materials

The leaf specimens of *Camellia cattienensis* were collected from Bau Sau station, Cat Tien National Park, (Dong Nai Province, Vietnam) by Van Hop Nguyen, where its type specimen was collected. The voucher specimen was CT22092022, and it was deposited in the herbarium of the Faculty of Natural Resources and Environment, Vietnam National University of Forestry-Dong Nai Campus, Vietnam.

Bacterial strains

Ten bacterial strains were used to identify the antibacterial

activity of the studied species, including six Gram-negative strains (*Escherichia coli* ATCC 25922, *Salmonella typhimurium* ATCC 13311, *Klebsiella pneumoniae* ATCC 13883, *Klebsiella pneumoniae* ATCC 700603, *Shigella flexneri* ATCC 9199, *Enterobacter hormaechei* ATCC 700323) and four Gram-positive strains (*Staphylococcus saprophyticus* BAA750, *Staphylococcus aureus* ATCC 29213, *Staphylococcus aureus* ATCC 25923, *Bacillus cereus* ATCC 13883).

Extraction procedures

The leaves of *C. cattienensis* were freshly harvested, well-washed, air-dried at 50 °C, and evenly ground into fine powder. 500 mL of acetone solution (99%, Thermo Fisher Scientific, USA) was used to immerse 100 g of powder for 72 h; after that, the supernatant was collected and filtered. The remaining solid matter underwent two more rounds of acetone extraction, and the end product was recovered by combining all the filtered fractions. The solvent was then eliminated in vacuum condition at 45 °C.

Three grams of acetone extract were dissolved in 30 mL of distilled water in an extraction flask. Then, 30 mL n-hexane was added, shaken, and left standing for layer formation. The hexane layer on the top was collected, and this procedure was performed again two more times to pick up 90 mL of the n-hexane extract. This extract was also eliminated under vacuum conditions at 45 °C to obtain the hexane fraction. The same process done above was used to obtain ethyl acetate and chloroform (Le et al. 2021).

Gas chromatography-mass spectrometry assays

The chemical constituents of the acetone extract and its fractions (n-hexane, ethyl acetate, and chloroform) from *C. cattienensis* leaves were determined using the TRACE 1310 Gas Chromatograph in conjunction with the ISQ 7000 mass spectrometer (Thermo Fisher Scientific, USA). The DB-5MS column (Agilent, USA) was used as a stationary phase with GC/MS, and run parameters were configured as previously described by Nguyen et al. (2023). Acquired mass spectral data were used to compare with the NIST 2017 library to determine the exact chemical compositions.

Determination of antibacterial activity

The disk diffusion method was employed to assess the antibacterial efficacy of the acetone extract from the

studied species, following the Clinical and Laboratory Standards Institute guidelines (CLSI 2016). Acetone extracts at concentrations of 100, 150, and 200 mg mL⁻¹ in 15% dimethyl sulfoxide were used for the test. As a positive control, a 10-µg gentamicin disk (Nam Khoa BioTek, Vietnam) was used, while 15% DMSO served as the negative control. The experiment was conducted in triplicate, and Fisher's least significant difference (LSD) and one-way analysis of variance (ANOVA) were employed for statistical analysis.

DPPH radical scavenging assay

DPPH radical scavenging activity of the acetone extract from the studied species was determined by DPPH assay (Nguyen et al. 2023) with slight modifications. Methanol 99.8% was used to dissolve the extract into different concentrations. The mixture, composed of 0.3 mL of extract and 3.7 mL of DPPH 0.1 mM was incubated at room temperature for 30 min in the dark. Absorbance measurement was done using a UV-Vis spectrophotometer (Genesys 20, USA) at 517 nm. DPPH radical scavenging activity was calculated as follows the Equation 1:

DPPH(%) =
$$\frac{A_0 - A_i}{A_0} \times 100$$
 (1)

Where A_{\circ} and A_{\circ} are the absorbance of the DPPH solution and the sample-DPPH mixture, respectively. The DPPH radical scavenging effect was determined by IC $_{50}$ value in comparison with the control ascorbic acid.

ABTS radical scavenging assay

Antioxidant activity was determined by ABTS radical scavenging assay described by Re et al. (1999) with slight modifications. Initially, solution A comprised 7 mM ABTS and 2.45 mM $\rm K_2S_2O_8$, was prepared and incubated at 37 °C for 18 h in the dark. Subsequently, a mixture of 3 mL of solution A, 0.1 mL of the studied extract, and 1.9 mL of acetone was made, followed by 15 min incubation in the dark. To assess the ABTS radical scavenging activity, the absorbance of the solution was measured at 734 nm using a UV-VIS spectrophotometer (UVS 2800, Labome, USA) equipped with UVWin6 Software. Ascorbic acid served as the reference standard, and the standard curve for

ascorbic acid (0 to 15 ppm) was constructed. The sample concentration was determined from the standard curve equation and expressed as µg mL⁻¹ ascorbic acid.

RESULTS AND DISCUSSION

Chemical components of the acetone extract and chloroform, ethyl acetate, and hexane fractions from *Camellia cattienensis*

The chemical components of the acetone extract and chloroform, ethyl acetate, and hexane fractions of C. cattienensis are shown in Table 1 and Figure 1. A total of 54 compounds were found in the four extracts studied. The acetone extract was found to be rich in 2-pentanone, 4-hydroxy-4-methyl (22.85%); 3,7,11,15-tetramethyl-2-hexadecen-1-ol (17.14%); neophytadiene (16.97%); stigmasterol (12.78%); α -amyrin (5.29%); and n-hexadecanoic acid (4.75%). The chloroform fraction possessed phenol, 2,4-bis(1,1-dimethylethyl)-, phosphite (3:1) (74.91%); n-hexadecanoic acid (8.30%); and 7,9-di-tertbutyl-1-oxaspiro (4.5) deca-6,9-diene-2,8-dione (5.08%) as the major compounds. The ethyl acetate fraction was mainly composed of 2-pentanone, 4-hydroxy-4-methyl (54.68%); phenol, 2,4-bis(1,1-dimethylethyl)-, phosphite (3:1) (9.17%); n-hexadecanoic acid (8.25%); and 5-hydroxymethylfurfural (8.08%) whereas hexanedioic acid, bis(2-ethylhexyl) ester (30.89%); neophytadiene (25.21%); 3,7,11,15-tetramethyl-2-hexadecen-1-ol (11.58%); n-hexadecanoic acid (10.26%); and 9,12,15-octadecatrienoic acid, (Z,Z,Z)- (8.44%) were the major compounds in the n-hexane fraction.

Previous studies have also demonstrated the chemical compositions of various *Camellia* species collected from Vietnam. For example, the essential oil of the *C. longii* flower was found to be rich in α -eudesmol (16.1%), (E)-nerolidol (13.0%), and β -eudesmol (8.9%) (Tran et al. 2023). Hoang et al. (2014) determined 35 volatile compounds that could be the main factor responsible for the black tea's aroma quality (*Camellia sinensis*), of which α -ionone, ethyl caprylate, 3-hydroxy- β -damascone, β -ionone, 2(4H)-benzofuranone were the main factors contributing for this (Hoang et al. 2014). The seed oil of the *Camellia ninhii* was characterized by the predominance of oleic acid (45.43%), palmitic acid (27.83%), and *trans*-cinnamic acid (4.83%) (Tran et al. 2022).

Table 1. Chemical components of the acetone extract and its fractions.

STT	RT	Compounds	Relative percentage (%)				
			CAT	CATC	CATE	CATH	
1	2.20	Acetic acid, butyl ester	-	-	1.55	-	
2	2.41	3-Furaldehyde	-	-	0.74	-	
3	2.43	3-Penten-2-one, 4-methyl-	1.77	-	-	-	
4	2.48	2-Pentanone, 4-hydroxy-4-methyl-	22.85	-	54.68	0.24	
5	2.82	Benzene, 1,3-dimethyl	-	-	0.15	-	
6	3.04	2-Propenoic acid, butyl ester	-	-	1.65	-	
7	3.09	1,3,5,7-Cyclooctatetraene	-	-	0.23	-	
8	3.81	2-Heptanol, acetate	0.47	-	-	-	
9	5.81	Ethanol, 2,2'-oxybis-, diacetates	0.10	-	-	-	
10	6.33	Dodecane	-	-	1.35	-	
11	6.43	2,6-Dimethyldecane	-	-	0.22	-	
12	7.44	Linalool	0.38	-	-	-	
13	7.96	Octanoic acid	-	-	0.42	-	
14	8.03	2,6-Dimethyl-6-nitro-2-hepten-4-one	0.35	-	-	-	
15	8.14	Benzoic acid	0.18	-	-	-	
16	8.27	Azulene	-	-	0.69	-	
17	8.51	Undecane, 2,6-dimethyl	-	-	0.22	-	
18	8.61	Dodecane, 4-methyl	-	-	0.17	-	
19	8.74	5-Hydroxymethylfurfural	0.20	-	8.08	-	
20	9.02	Tetradecane, 5-methyl-	-	-	0.50	-	
21	9.14	Dodecane, 2,6,11-trimethyl-	-	-	1.38	-	
22	10.08	Undecane, 4,7-dimethyl-	-	0.56	-	-	
23	10.84	1-lodo-2-methylundecane	-	-	1.45	-	
24	10.97	Pentanoic acid, 5-hydroxy-, 2,4-di-t-butylphenyl esters	-	0.30	-	-	
25	11.53	Phenol, 2,4,6-tri-tert-butyl-	-	0.62	-	-	
26	12.19	Tetradecane, 2,6,10-trimethyl-	-	0.93	1.40	-	
27	12.66	Benzyl Benzoate	-	0.79	-	-	
28	12.96	Neophytadiene	16.97	2.13	-	25.21	
29	13.36	7,9-Di-tert-butyl-1-oxaspiro (4,5) deca-6,9-diene-2,8-dione	-	5.08	1.43	-	
30	13.37	3,7,11,15-Tetramethyl-2-hexadecen-1-ol	17.14	-	-	11.58	
31	13.57	n-hexadecenoic acid	4.75	8.30	8.25	10.26	
32	14.45	9,12,15-Octadecatrienoic acid, 2,3-dihydroxypropyl ester, (Z,Z,Z)-	-	-	2.01	-	
33	14.46	9,12,15-Octadecatrienoic acid, (Z,Z,Z)-	-	-	-	8.44	
34	14.55	Octadecanoic acid	0.56	2.53	2.25	1.97	
35	14.63	Phytol	1.34		-	1.01	
36	14.72	9,12-Octadecadienoic acid (Z,Z)-	0.28	-	-	-	
37	15.36	Octadecane, 3-ethyl-5-(2-ethylbutyl)-	-	0.85	-	-	
38	15.50	17-Pentatriacontene	0.35	-	_	_	

Table 1

STT	RT	Compounds	Relative percentage (%)				
			CAT	CATC	CATE	CATH	
40	15.66	Oleic Acid	0.29	-	-	-	
41	16.18	Benzenepropanoic acid, 3,5-bis(1,1-dimethylethyl)-4-hydroxy-, 2-ethylhexyl ester	-	2.08	-	-	
42	16.41	Bis(2-ethylhexyl) phthalate	-	-	-	1.61	
43	16.59	Octadecane, 3-ethyl-5-(2-ethylbutyl)-	0.51	-	-	-	
44	16.70	Hexadecenoic acid, 1-(hydroxymethyl)-1,2-ethanediyl ester	0.72	-	-	-	
45	16.80	Oleic acid, 3-(octadecyloxy)propyl ester	0.73	-	0.18	-	
46	16.83	Octadecanal, 2-bromo	0.65	-	-	-	
47	17.26	Octadecane, 3-ethyl-5-(2-ethylbutyl)-	0.58	-	-	-	
48	18.15	1,2-Cyclohexanedicarboxylic acid, dinonyl ester	-	-	-	2.35	
49	18.54	Squalene	2.42	-	-	2.84	
50	24.05	dl-α-Tocopherol	2.26	-	-	-	
51	27.26	Stigmasterol	12.78	-	-	3.42	
52	27.63	Phenol, 2,4-bis(1,1-dimethylethyl)-, phosphite (3:1)	-	74.91	9.17	-	
53	28.03	α-Amyrin	5.29	-	-	-	
54	28.30	β-Sitosterol	4.30	-	-	-	
		Total	98.22	99.08	98.17	98.82	

Note: CAT: acetone extract, CATC: chloroform, CATE: ethyl acetate, and CATH: hexane.

Furthermore, a recent study reported that the leaf and flower of *C. tonkinensis* consisted of Zn (10.20 and 13.40 mg kg⁻¹) and saponin (58.30 and 87.10 mg g⁻¹) whereas various amino acids were also present in two organs of this plant, including aspartate, glutamate, serine, histamine, glycine, arginine, threonine, alanine, cysteine, tyrosine, valine, phenylalanine, methionine, leucine, isoleucine, proline, and lysine with the contents ranging from 3.512 to 42.087 g kg⁻¹ (Dang et al. 2022).

Studies have identified the chemical compounds in various *Camellia* species extracts using different solvents and analyzed them with gas chromatography-mass spectrometry. For instance, the methanol extract isolated from *Camellia sinensis* leaves collected from Bangladesh mainly contained caffeine (27.44%), hexadecanoic acid, methyl ester (14.02%), 9,12,15-octadecatrienoic acid methyl ester (Z, Z, Z) (3.95%) (Hasan et al. 2024). Similarly, the chloroform: methanol extract of the *C. sinensis* leaves grown in Ambala Cantt., India was found to be rich in caffeine (48.21%), *trans*-13-octadecenoic acid (18.30%), *cis*-11-eicosenoic acid (15.15%) (Gupta and Kumar

2017). In addition, the methanol extract of *C. sinensis* leaves from Dehradun, India, was mainly composed of *n*-heptadecanol-1 (68.63%); 2-pentanone, 4-hydroxy-4methyl- (3.82%); and 7-hexadecanoic acid, methyl ester, (Z) (2.32%) (Pradhan and Dubey 2021). The phytochemistry of the methanol extract of *C. sinensis* leaves collected from six regions of India was also investigated. Accordingly, the sample extracts from Moonar (Kerala) and Kodaikanal (Tamil Nadu) were characterized by the predominance of caffeine (46.25-57.17%); 1,2,3-benzenetriol (18.40-16.28%); and 1,3,4,5-tetrahydroxycyclohexanecarbonyl (13.96-9.64%). Meanwhile, the extracts grown in Ootacamund (Tamil Nadu) and Bengaluru (Karnataka) contained caffeine 1,3,7-trimethyl-3,7-dihydro-1H-purine-2,6-dione (56.48-57.52%);1,2,3-benzenetriol (28.36-21.55%) as the major compounds. Finally, 1,3,7-trimethyl-3,7-dihydro-1H-purine-2,6-dione (42.75-59.79%) as the most abundant compound in the extracts from Assam (Assam) and Kolkata (West Bengal) (Senthilkumar et al. 2015). Apart from that, the major constituents of the ethanol extract of the C. sinensis leaves from Uganda were caffeine (82.69%); naphthacene-5,12-dione, 6,11

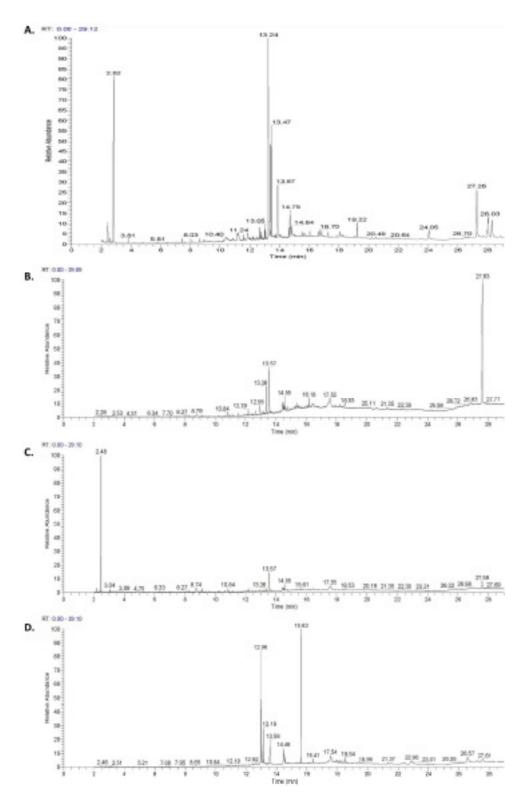


Figure 1. The GC chromatogram of **A.** acetone extract from *Camellia cattienensis* and **B.** chloroform, **C.** ethyl acetate, and **D.** hexane fractions from *Camellia cattienensis*.

-dihydroxy-2,3,8,9-tetramethyl- (3.73%); and estra-1,3,5(10)-trien-17-ol, 2,3,4-trimethoxy-, (17. β .)- (3.73%) (Hope et al. 2022).

The benzene-ethanol extracts of *Camellia oleifera* leaves collected from Hunan, China, were found rich in butyraldehyde, semicarbazone (11.58%); hexatriacontane (8.04%); and 1,6-anhydro- β -D-glucopyranose (7.54%) (Liu et al. 2009). The chemical constituents of different extracts of *C. oleifera* fruit grown in Hunan, China, were also reported. For instance, the methanol extract contained a mixture of γ -sitosterol (43.32%); 5-hydroxymethylfurfural (22.07%); and *cis*-vaccenic acid (12.71%). The ethanol extract was mainly composed of 5-hydroxymethylfurfural (58.78%) and furfural (4.74%), while *cis*-vaccenic acid (45.20%); *n*-hexadecanoic acid (10.98%); and 9-octadecenamide, (Z)- (5.82%) were the main compounds in the ethyl acetate extract (Xie et al. 2018). Moreover, the acetone extract of *Camellia assamica*

leaves collected from Dehradun, India was characterized by the predominance of 2',6'dihydroxyacetophenone, bis(trimethylsilyl) ether (17.58%); *N*(trifluoracetyl) *O,O',O"*tris(trimethylsilyl) epinephrine (15.83%); and tetracosamethyl cyclododecasiloxane (10.62%) (Pradhan and Dubey 2021).

Antibacterial activity of acetone extract from *Camellia* cattienensis

The antibacterial property of *C. cattienensis* acetone extract was presented in Table 2. Accordingly, the studied extract was found to be effective against four out of eight bacterial strains, including *K. pneumoniae* ATCC 700603, *S. aureus* ATCC 29213, *S. aureus* ATCC 25923, and *S. saprophyticus* BAA750. Overall, at a concentration of 200 mg mL⁻¹, the studied extract exhibits a stronger antibacterial effect against four bacterial strains in comparison with the remaining concentrations (Table 2).

Table 2. Antibacterial property of acetone extracts from Camellia cattienensis.

	Zone of inhibition (mm)					
Bacterial strains	100 (mg mL ⁻¹)	150 (mg mL ⁻¹)	200 (mg mL ⁻¹)	Gentamycin	Negative control	
B. cereus ATCC 13883	-	-	-	14.17±0.29	-	
E. coli ATCC 25922	-	-	-	13.33±0.58	-	
E. hormaechei ATCC 700323	-	-	-	8.83±0.76	-	
K. pneumoniae ATCC 700603	4.00±0.87a	4.83±0.29ª	5.33±1.15ª	12.83±0.76b	-	
K. pneumoniae ATCC 13883	-	-	-	13.50±0.87	-	
S. aureus ATCC 29213	2.50±0.50 ^a	4.00±0.50 ^b	4.33±0.58b	13.67±1.53°	-	
S. aureus ATCC 25923	2.67±1.15 ^a	4.33±0.58 ^a	6.67±0.76 ^b	13.00±1.00°	-	
S. flexneri ATCC 9199	-	-	-	12.00±0.87	-	
S. saprophyticus BAA750	2.33±0.58 ^a	2.50±0.50 ^a	3.67±0.29b	13.33±0.58°	-	
S. typhimurium ATCC 13311	-	-	-	14.50±0.87	-	

Different superscript lower-case letters in the same row denote significant difference (P<0.05). (-) Not active.

Studies provided the antimicrobial activities of different solvent extracts from *Camellia* species. For example, the chloroform: methanol extract of the *Camellia sinensis* leaves grown in Ambala Cantt., India had an inhibitory effect on *Staphylococcus aureus, Escherichia coli, Pseudomonas aeruginosa*, and *Candida albicans* (Gupta and Kumar 2017). The methanol extract of the *C. sinensis* and *C. assamica* leaves from Dehradun, India and its various fractions such as water, ethanol, methanol, chloroform, and petroleum ether were reported

to possess antimicrobial effects against *Escherichia coli, Salmonella typhi, Klebsiella pneumoniae, Listeria monocytogenes, Staphylococcus aureus, Pseudomonas aeruginosa*, and *Candida albicans* (Pradhan and Dubey 2021). In another report, the antibacterial effects of the hot water extract isolated from the green, herbal, and black teas (*C. sinensis*) were also investigated. Accordingly, the first extract was found to be effective against three bacterial strains, including *Micrococcus luteus, Bacillus cereus*, and *Staphylococcus aureus*, while the latter

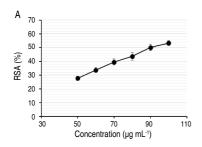
extracts exhibited an inhibitory effect on *Micrococcus luteus* and *Bacillus cereus* (Chan et al. 2011).

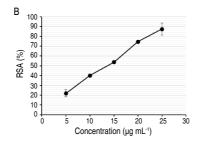
The ethanol extract of *C. sinensis* leaves from Uganda was demonstrated to be effective against different pathogenic bacteria, especially *Salmonella* and *Escherichia coli* (Hope et al. 2022). Also, the leaf ethanol extract of this plant collected from Chennai, India, displayed activity against *Streptococcus mutans* and *Lactobacillus acidophilus* (Anita et al. 2015). The ethanol extract of *C. sinensis* leaves grown in Iran was effective against 30 *Escherichia coli* strains isolated from the urine cultures of patients in three hospitals in Zabol, southeastern Iran (Sepehri et al. 2014). Additionally, the organic acids and phenolic components from the seeds of *Camellia oleifera* cake, collected in Jiangxi, China, exhibited antimicrobial activity against six fungal and bacterial strains, including *Aspergillus oryzae*, *Rhizopus stolonifer*, *Mucor racemosus*,

Bacillus subtilis, Staphylococcus aureus, and Escherichia coli (Zhang et al. 2020). Furthermore, another study demonstrated the antibacterial effects of the methanol extract fractioned into basic, neutral, and acid fractions obtained from Camellia japonica grown in Yeosoo, Korea. Accordingly, the methanol extract, neutral, and acid fractions displayed activity against some bacterial strains, including Escherichia coli, Salmonella typhimurium, Listeria monocytogenes, and Staphylococcus aureus (Kim et al. 2001).

Antioxidant activity of acetone extract from Camellia cattienensis

The antioxidant effects of the tested extract from C. cattienensis were shown in Figure 2. Accordingly, the extract had the DPPH, ABTS free radical scavenging with the IC₅₀ values of 91.63 \pm 1.88 and 13.32 \pm 0.49 μ g mL⁻¹, respectively.





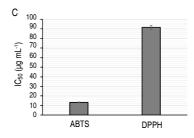


Figure 2. The free radical scavenging activity of the acetone extract from *Camellia catienensis* was measured using **A.** the DPPH assay, **B.** the ABTS assay, and **C.** the corresponding IC_{50} values.

Compared to other Camellia species, the results show moderate antioxidant potential. For example, the methanol extract of C. sinensis leaves from Bangladesh showed a stronger DPPH scavenging effect with an IC₅₀ value of 69.51 μg mL⁻¹, indicating better antioxidant activity compared to C. cattienensis (Hasan et al. 2024). Conversely, the acetone extract of C. oleifera seed oil from Taiwan exhibited a much lower antioxidant effect, with only 4.46% inhibition at 200 μg mL⁻¹, significantly weaker than *C. cattienensis* (Lee and Yen 2006). Similarly, the ethyl acetate extract of C. oleifera seed oil showed only 5.92% inhibition, while the methanol extract performed better, with 66.50% inhibition at the same concentration (Lee and Yen 2006). In another study, the organic acids and phenolic components of C. oleifera seed cake from Jiangxi, China, demonstrated weaker antioxidant activity than C. cattienensis, with IC₅₀ values of 184 mg L-1 and 103 mg L-1, respectively (Zhang et al. 2020). Furthermore, the ethanol extract of C. japonica flowers from Korea demonstrated a DPPH inhibition of up to 60% at 50 μ g mL $^{-1}$ (Piao et al. 2011), while the ethanol extract of C. japonica leaves from Jeonnam, Korea, showed a stronger antioxidant effect with an IC $_{50}$ value of 38.53 μ g mL $^{-1}$ (Yoon et al. 2017).

Overall, the acetone extract of *C. cattienensis* leaves displayed a moderate antioxidant effect, stronger than that of *C. oleifera* extracts but weaker compared to the more potent antioxidant effects of *C. sinensis* and *C. japonica* ethanol and methanol extracts. These differences likely reflect the variations in solvent choice, plant parts used, and regional factors influencing the phytochemical composition.

CONCLUSION

Camellia cattienensis is a rare species that, to date, has

only been documented from the type collection at Nam Cat Tien National Park, Dong Nai Province, Vietnam. In the present study, a limited number of samples of this species were utilized for experimental purposes to prioritize its conservation. Furthermore, phytochemical, antioxidant, and antibacterial properties are recognized as prominent features in Camellia species. Therefore, the present study provides these characteristics for the first time from the acetone extract of *C. cattienensis*. As a result, the various acetone sub-extracts were found to contain several bioactive compounds, with 2-pentanone, 4-hydroxy-4-methyl-phenol, 2,4-bis(1,1-dimethylethyl)-, phosphite (3:1); and hexanedioic acid, bis(2-ethylhexyl) identified as the most abundant compounds. The acetone extracts of C. cattienensis exhibited moderate effects against bacterial strains, including Klebsiella pneumoniae, Staphylococcus aureus, and Staphylococcus saprophyticus. Notably, this extract demonstrated potent antioxidant activity, as evaluated using the DPPH and ABTS assays, with IC50 values of 91.63±1.88 and 13.32±0.49 µg mL⁻¹, respectively. The results of analyses, such as the antioxidant and antibacterial activities of the n-hexane, chloroform, and ethyl acetate fractions, will be reported in a subsequent study. Given the versatile applications of various Camellia species in the food industry, the findings of this study contribute to a deeper understanding of the potential food uses of *C. cattienensis*, particularly in the beverage industry.

CONFLICT OF INTERESTS

The author declares no conflict of interest.

REFERENCES

Anand J, Upadhyaya B, Rawat P and Rai N (2015) Biochemical characterization and pharmacognostic evaluation of purified catechins in green tea (*Camellia sinensis*) cultivars of India. 3 Biotech 5(3): 285–294. https://doi.org/10.1007/s13205-014-0230-0

Anita P, Balan In, Ethiraj S et al (2015) In vitro antibacterial activity of *Camellia sinensis* extract against cariogenic microorganisms. Journal of Basic and Clinical Pharmacy 6(1): 35-39. https://www.jbclinpharm.org/articles/in-vitro-antibacterial-activity-of-camellia-sinensis-extract-against-cariogenic-microorganisms.pdf

Chan EW, Tie P, Soh E and Law Y (2011) Antioxidant and antibacterial properties of green, black, and herbal teas of *Camellia sinensis*. Pharmacognosy Research 3(4): 266–272. https://doi.org/10.4103/0974-8490.89748

Chitsazan A (2015) Anti-cancer properties of green Tea probed via quantum mechanics calculations. Oriental Journal of Chemistry 31(1): 393–408. https://doi.org/10.13005/ojc/310147

CLSI - Clinical & Laboratory Standards Institute (2016) Methods

for antimicrobial dilution and disk susceptibility testing of infrequently isolated or fastidious bacteria. 3rd ed. CLSI guideline M45. Wayne PA. Clinical and Laboratory Standard Institute.

Dang VH, Nguyen TC, Le VQ et al (2022) Content of some chemical components from Ba Vi yellow *Camellia* plant (*Camellia tonkinensis* (Pit.) Cohen-Stuart). Vietnam Journal of Agriculture and Rural Development 434: 72-76.

Gaur R and Bao GH (2021) Chemistry and pharmacology of natural catechins from camellia sinensis as anti-MRSA agents. Current Topics in Medicinal Chemistry 21(17): 1519-1537. https://doi.org/10.2174/1568026621666210524100632

Gupta D and Kumar M (2017) Evaluation of in vitro antimicrobial potential and GC–MS analysis of *Camellia sinensis* and *Terminalia arjuna*. Biotechnology Reports 13: 19–25. https://doi.org/10.1016/j. btre.2016.11.002

Hasan R, Haque MM, Hoque A et al (2024) Antioxidant activity study and GC-MS profiling of *Camellia sinensis* Linn. Heliyon 10(1): e23514. https://doi.org/10.1016/j.heliyon.2023.e23514

Hoang QT, Tham BHP, Vu HS et al (2014) Sensory aroma and related volatile flavor compound profiles of different black tea grades (*Camellia sinensis*) produced in Northern Vietnam. In: Valentin D, Chollet S, Le S et al (2014) Summer Program in Sensory Evaluation 2014, Vietnam. VNU-HCM Publishing House, 113-119.

Hope O, Bright IE and Alagbonsi AI (2022) GC-MS biocomponents characterization and antibacterial potency of ethanolic crude extracts of *Camellia sinensis*. SAGE Open Medicine 10: 1-13. https://doi.org/10.1177/20503121221116859

Kim JK, Park HG, Kim CR et al (2014) Quality evaluation on use of *Camellia* oil as an alternative method in dried seaweed preparation. Preventive Nutrition and Food Science 19(3): 234–241. https://doi.org/10.3746/pnf.2014.19.3.234

Kim KY, Davidson PM and Chung HJ (2001) Antibacterial activity in extracts of *Camellia japonica* L. petals and its application to a model food system. Journal of Food Protection 64(8): 1255–1260. https://doi.org/10.4315/0362-028X-64.8.1255

Le HT, Luu TN, Nguyen HMT et al (2021) Antibacterial, antioxidant and cytotoxic activities of different fractions of acetone extract from flowers of *Dipterocarpus intricatus* Dyer (Dipterocarpaceae). Plant Science Today 8(2): 273–277. https://doi.org/10.14719/pst.2021.8.2.1086

Lee CP and Yen GC (2006) Antioxidant activity and bioactive compounds of tea seed (*Camellia oleifera* Abel.) oil. Journal of Agricultural and Food Chemistry 54(3): 779–784. https://doi.org/10.1021/if052325a

Liu QM, Peng WX, Wu YX et al (2009) Analysis of biomedical components of *Camellia oleifera* Leaf and kernel hull by GC/MS. 2009 3rd International Conference on Bioinformatics and Biomedical Engineering IEEE 1-4.

Nguyen ND, Trinh NN, Nguyen QH et al (2023) Chemical profiles and biological activities of acetone extracts of nine Annonaceae plants. Journal of Applied Botany & Food Quality 96: 148-156. https://doi.org/10.5073/JABFQ.2023.096.019

Orel G and Wilson PG (2011) *Camellia cattienensis*: a new species of *Camellia* (sect. Archaecamellia: Theaceae) from Vietnam. Kew Bulletin 66(4): 565–569. https://www.jstor.org/stable/23216738

Piao MJ, Yoo ES, Koh YS et al (2011) Antioxidant effects of the ethanol extract from flower of *Camellia japonica* via scavenging of reactive oxygen species and induction of antioxidant enzymes. International Journal of Molecular Sciences 12(4): 2618–2630. https://doi.org/10.3390/ijms12042618

Pradhan S and Dubey RC (2021) GC–MS analysis and molecular docking of bioactive compounds of *Camellia sinensis* and *Camellia assamica*. Archives of Microbiology 203(5): 2501–2510. https://doi.org/10.1007/s00203-021-02209-6

Quach V H, Luong VD, Doudkin RV et al (2021) Diversity of the genus Camellia L. (Theaceae) in Lam Dong Province. Academia Journal of Biology 43: 129-138. https://www.researchgate.net/publication/357451755

Re R, Pellegrini N, Proteggent, A et al (1999) Antioxidant activity applying an improved ABTS radical cation decolorization assay. Free radical biology and medicine 26: 1231-1237. https://doi.org/10.1016/s0891-5849(98)00315-3

Senthilkumar SR, Sivakumar T, Arulmozhi KT and Mythili N (2015) Gas chromatography-mass spectroscopy evaluation of bioactive phytochemicals of commercial green teas (*Camellia sinensis*) of India. Asian Journal of Pharmaceutical and Clinical Research 8: 278-282

Sepehri Z, Hassanshahian M, Shahi Z et al (2014) Antibacterial effect of ethanol extract of *Camellia sinensis* L. against *Escherichia coli*. Asian Pacific Journal Microbiology Research 2: 6-8.

Tran TTQ, Trinh TD, Nguyen TTU and Le VS (2022) Fatty acid

composition and antioxidant activity of *Camellia ninhii* seed oil collected in Lam Dong province. Dalat University Journal of Science 12(3): 27–33. https://doi.org/10.37569/DalatUniversity.12.3.993(2022)

Tran TH, Nguyen TC, Dung VC et al (2023) Characterization and evaluation of the in vitro antioxidant, α -glucosidase inhibitory activities of *Camellia longii* Orel and Luu. (Theaceae) llower essential oil and extracts from Vietnam. Natural Product Communications 18(11). https://doi.org/10.1177/1934578X231208348

Xie Y, Ge S, Jiang S et al (2018) Study on biomolecules in extractives of *Camellia oleifera* fruit shell by GC–MS. Saudi Journal of Biological Sciences 25(2): 234–236. https://doi.org/10.1016/j.sjbs.2017.08.006

Yang C, Liu X, Chen Z et al (2016) Comparison of oil content and fatty acid profile of ten new *Camellia oleifera* cultivars. Journal of Lipids 2016, 3982486. https://doi.org/10.1155/2016/3982486

Yoon IS, Park DH, Kim JE et al (2017) Identification of the biologically active constituents of *Camellia japonica* leaf and anti-hyperuricemic effect in vitro and in vivo. International Journal of Molecular Medicine 39(6): 1613–1620. https://doi.org/10.3892/ijmm.2017.2973

Zhang D, Nie S, Xie M and Hu J (2020) Antioxidant and antibacterial capabilities of phenolic compounds and organic acids from *Camellia oleifera* cake. Food Science and Biotechnology 29(1): 17–25. https://doi.org/10.1007/s10068-019-00637-1